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In-vivo anti-tumor activities of curcumin-solid-dispersion loaded floating tablets on stomach and lung cancers

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ABSTRACT

Curcumin, a natural compound commonly found in turmeric (Curcuma longa L.), has been proven effective in stomach and lung cancer treatment. However, due to its poor solubility in water and instability in alkaline medium, curcumin oral application is still limited. Thus, in this study, a curcumin-solid-dispersion (CSD) loaded floating tablet was successfully developed and evaluated for its anti-cancer properties against lung and stomach cancer in an in-vivo setting in mice. The results demonstrated that the CSD-loaded floating tablets did not exhibit acute oral toxicity in healthy mice at maximum doses of ~10 g/kg body weight. Moreover, tested in a carcinogenic mice model induced with 7,12-dimethyl benz[a]anthracene, the tablets could significantly inhibit the growth of stomach/lung cancer cells and reduce the cancer rate up to ~40%, comparable to that of the standard drug 5-fluorouracil (5-FU). In addition, the CSD-loaded floating tablets could complement the drug 5-FU efficacy in treating stomach/lung cancers, with a cancerous tumor reduction rate of ~65%. Finally, the research confirmed that using the tablet in the early stages would reduce the cancer rate better than using it in the late stages. In conclusion, the CSD-loaded floating tablets showed much potential as an oral dosage form for stomach and lung cancer treatments.

INTRODUCTION

Lung cancer and stomach cancer are two of the most common cancers in the world with the first [1] and the fourth [2] leading cause of cancer death, respectively. Recently, the applications of surgery, chemotherapy, and radiation therapy in the treatment of stomach/lung cancer are necessary. However, synthetic chemicals with high toxicity attack nonspecifically both cancer cells and normal ones, thus possessing numerous side effects and significantly affecting patients' health [3,4]. Therefore, a novel therapeutic approach is necessary to circumvent the mentioned issues.

Curcumin, the main component of the rhizome of turmeric (Curcuma longa L., Zingiberaceae), has been well studied and considered as an anti-cancer agent for various kinds of cancers such as stomach, liver, lung, colon, breast, and skin cancer, as well as an active ingredient for anti-inflammation, antioxidation, antibacterial, antiviral, and antifungal [1,2,5-8]. Moreover, biological and clinical trials have proven that curcumin could safely and effectively treat cancers, especially stomach cancer [1,9,10]. Nevertheless, curcumin has not been widely used in oral drugs due to its low solubility in water and acidic pH, and its instability in alkaline environments [2,8,11-13]. Therefore, in our previous report, curcumin was employed in the curcumin-solid-dispersion (CSD) loaded floating tablets, which demonstrated perfect physicochemical properties such as short floating potential of 35 ± 1 seconds, long floating retention time of more than 8 hours, enhanced curcumin solubility to more than 200 times, outstanding stability for at least 2 years at room temperature, and reserved curcumin cytotoxicity on the

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N87 stomach cell lines with a better efficacy than the standard drug 5-fluorouracil (5-FU) [14].

Herein, this present study further evaluated the anticancer effects of the CSD-loaded floating tablets in an *in-vivo* setting in stomach and lung cancer mice models. First, the acute toxicity of the tablets was tested in healthy mice. Then, the cytotoxicity effects of the tablets were determined in tumorbearing mice, induced by the 7,12-dimethyl benz[a]anthracene (DMBA) chemical. The mice were monitored for their vital signs, weights, and tumor characteristics (i.e., size, positions, macroscopic and microscopic images).

MATERIALS AND METHODS

Materials

Healthy Swiss albino mice (weight 30 to 60 g), 7–8 weeks old, were obtained from the Pasteur Institute in Ho Chi Minh City, Vietnam, and settled in appropriate laboratory conditions for a week before experimentations. Mice from different groups were housed in separate cages (max. 10 mice/ cage) under room temperature and natural lighting conditions. Mice were fed a standard diet of 2.5/10 g body weight per day with free water intake. The cages were regularly cleaned throughout the experiment period.

Curcumin (purity of 95.5%) was imported from India. Xanthan gum, hydroxypropyl methylcellulose (HPMC) K15M, and poly(vinyl pyrrolidone) (PVP) K30 were purchased from the USA. Aerosil, magnesium stearate, NaHCO₃, and citric acid were bought from China. Corn oil imported from Argentina. DMBA was purchased from the USA, and 5-FU (Ebewe[®] 500 mg/10 ml) was bought from Austria.

Preparations of CSD-loaded floating tablets

The CSD-loaded floating tablets were formulated following our previous report [14], with the optimal formula presented in Table 1. Briefly, the CSD was first fabricated by

incorporating curcumin in a mixture of PVP K30 and tween 80, with a ratio of curcumin: PVP K30: Tween 80 of 1:4:0.22 w/w/w, by the solvent method [15,16]. Then, the CSD floating tablets were formulated using the simple wet granulation method, with slight modifications [17]. The tablets are composed of the CSD (active ingredient), xanthan gum and HPMC K15M (matrix generators), PVP K30 (binder), NaHCO₂ and citric acid (gas-generating excipients), aerosil and magnesium stearate (glidant/lubricant), and lactose (diluent) (Table 1). The process began by homogeneous mixing the ingredients, followed by moistening and wetgranulating with PVP K30 ethanolic solution through a 2-mm sieve, drying at 50°C–60°C to moisture of \leq 7%, and mixing with gas-generating excipients and glidant/lubricant. Finally, the granules were compressed on a rotary tablet press (Rimek, India) with a tablet weight of ~1,000 mg. The blank floating tablets were similarly prepared, without the addition of CSD.

Table 1. CSD loaded floating tablet formulation.

Ingredient	Function	Amount (mg)
CSD ^a	Active ingredient	500
Xanthan gum	Matrix generator	49.5
HPMC K15M	Matrix generator	198.1
PVP K30	Binder	40
NaHCO ₃	Gas-generating excipient	101.5
Citric acid	Gas-generating excipient	50.8
Aerosil	Glidant/Lubricant	5
Magnesium stearate	Glidant/Lubricant	15
Lactose	Diluent	ad. 1,000

(a) Curcumin solid dispersion was fabricated using the solvent method with a ratio of curcumin: PVP K30: tween 80 of 1:4:0.22 w/w/w.

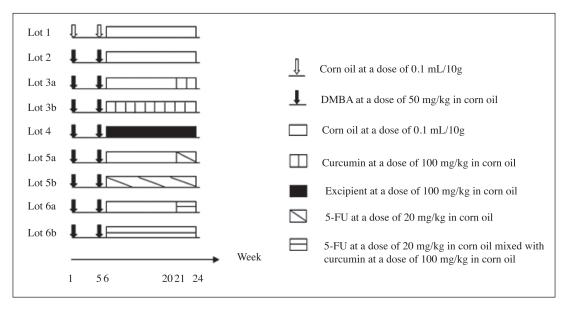


Figure 1. In-vivo cytotoxicity tests in lung/stomach cancer mice models timeline.

Blank floating tablets					CSD loaded floating tablets							
Test mice	1	2	3	4	5	6	1	2	3	4	5	6
Sex	8	8	8	Ŷ	Ŷ	Ŷ	3	8	8	Ŷ	Ŷ	Ŷ
Weight (g)	24.1	23.7	24.7	21.7	24.8	22.1	21.3	24.1	23.3	24.8	22.2	21.8
Volume (ml)	0.48	0.48	0.49	0.43	0.50	0.44	0.43	0.48	0.47	0.50	0.44	0.44
Total weight (g)		138.1					137.5					
Total volume (ml)		2.82					2.76					
Oral dose (mg/kg)	10,210						10,036					
Dead mice after 72 hours	0						0					
Dead mice after 14 days		0 0										

Table 2. Acute toxicity test of the blank and CSD loaded floating tablets on experimental mice.

Table 3. The number of dead mice in each experimental g	group	during
the anti-cancer testing period.		

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Stage	1	2	3a	3b	4	5a	5b	6a	6b	Total
Week 1-4	0	2	2	1	1	0	2	0	3	11
Week 5-24	0	1	1	4	4	2	5	2	5	24
Total	0	3	3	5	5	2	7	2	8	35

In-vivo acute toxicity test

Six Swiss albino mice (50% male, 50% female) were fasted for at least 12 hours before being given the maximum oral doses of the test samples (the blank floating tablets and the CSD-loaded floating tablets). The mice parameters of general movement, behavior, hair state, eating, urination, and the number of dead mice were monitored and recorded within 72 hours. After 72 hours, the mice with no abnormal signs were monitored for the next 14 days, followed by dissection to observe the macroscopic changes (if any) in vital organs, including the heart, lungs, intestines, liver, and kidneys.

After the test, the lethal dose (LD) was determined based on the number of dead mice. If 0% of mice died, the maximum dose (D_{max}) would be determined. If the mortality rate was 100% or lower, the dose would be repetitively reduced to half-level of the initial dose until an LD₅₀ was reached (i.e., a dose of which 50% of the mice died).

In-vivo cytotoxicity tests in lung/stomach cancer mice models

The experiments were carried out on 85 Swiss albino mice, which were randomly divided into 9 groups. Each group included 50% male and 50% female mice. The experimental process was conducted for 24 weeks following the timeline described in Figure 1 [18–20]. Briefly, mice in group 1 (physiologic control group) were taken corn oil, a medium that helps increase the DMBA absorption in the intestine, throughout the experimental process. Group 2 (disease control group) was fed with DMBA for the initial 5 weeks to induce cancerous tumors in the mice's stomach and lungs. Groups 3a and b were designed to test the effects of CSD-loaded floating tablets from week 21 to week 6 of this study, respectively, which evaluated the products' efficacy on these cancers at the early and late stages. The blank floating tablets were used in group 4 to assess whether excipients had any anti-cancer activity. Groups 5a and b (positive/reference control groups) were mice treated with 5-FU, a clinically used chemotherapy to treat stomach and lung cancers, from week 21 to 6, respectively. Finally, a combination of the experimental floating tablets and 5-FU was used in groups 6a and b from week 21 to 6, respectively, to evaluate the complementary anti-cancer effects of curcumin in the disease's early and late stages.

At each predetermined time interval of the testing duration, mice were monitored for their vital signs, weights, and tumor characteristics (i.e., size, positions, macroscopic and microscopic images).

Weights

Mice were weighed at a fixed hour each week from the beginning to week 24. The mean weight of different groups was calculated and compared.

Tumor macroscopic evaluation

After the experimental duration (24 weeks), mice were killed after being anesthetized with iced CO_2 , and their abdominal cavities were observed macroscopically. The mice's stomachs and lungs were separated and washed with 0.9% NaCl. The characteristics of color, surface states, and lesions/tumors appearance/numbers were noted. The newly formed tumors, if any, were fixed in a 10% formalin solution. After 48 hours, the number and volume of tumors in the lungs and stomachs were calculated in the following equations (1) to (4) [17,20–22].

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The percentage of
mice with tumors (%) = \frac{\text{The number of mice with tumors}}{\text{The total number of mice with}} \times 100\%
a full dose of DMBA
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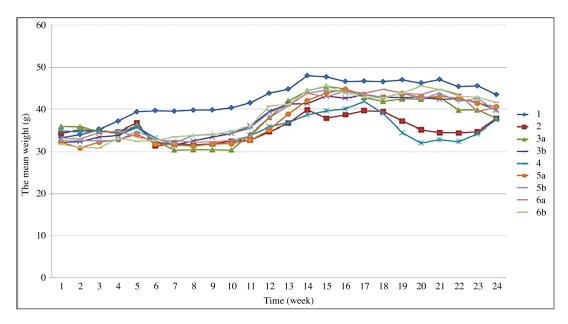


Figure 2. Mice weight changes of different experimental groups during the anti-cancer testing period.

Table 4. Fat index of different groups of mice from week 5 to 24.Results are expressed as mean \pm SD.

Group	Fat index (%)	Group	Fat index (%)	Group	Fat index (%)
1	0.26 ± 0.26	3b	$0.45\pm0.13^{\scriptscriptstyle\#}$	5b	$0.37\pm0.13^{\#}$
2	$2.06\pm0.53^{\ast}$	4	$2.09\pm0.46^{\ast}$	6a	$0.47\pm0.15^{\scriptscriptstyle\#}$
3a	$0.50\pm0.15^{\scriptscriptstyle\#}$	5a	$0.48\pm0.26^{\scriptscriptstyle\#}$	6b	$0.29\pm0.10^{\#}$

*p < 0.05 when compared to group 1; #p < 0.05 when compared to group 2.

The mean number of tumors =
$$\frac{\text{The mean number of tumors}}{\text{The total number of mice}}$$

with tumors

(2)

(3)

The tumor volume (mm³) = $1/2 \times$ The largest tumor diameter \times The smallest tumor diameter²

The mean tumor volume =
$$\frac{\text{The total volume of tumors}}{\text{The number of tumors in}}.$$
(4)

Tumor microscopic evaluation

For the microscopic analysis, the mice's stomachs and lungs were soaked in the 10% formalin solution, followed by staining with hematoxylin-eosin. Microscopic tumors were observed and assessed with optical microscopes in terms of cell morphology, dysplasia, and carcinoma [17,20–22].

Statistical analysis

The results were processed by Excel software, presented as mean \pm SD, and evaluated for statistical significance using SPSS version 22.

RESULTS AND DISCUSSIONS

In-vivo acute toxicity test

To check the toxicity of the formulas and to determine the test dose, the acute toxicity test of the blank and CSDloaded floating tablets was conducted on six mice (Table 2). The maximum dose levels of the blank and CSD-loaded floating tablets were 10.21 and 10.04 g/kg, respectively. At these doses, no dead mice were observed, and all mice were healthy with normal eating, drinking, urinating, and movement patterns, and had no abnormal signs within 72 hours after the test. Furthermore, 14 days after the test, mice dissections demonstrated no abnormalities in the heart, lungs, intestines, livers, and kidneys. Conclusively, the tablets, at a dose of as high as 10 g/kg, equivalent to curcumin amount, did not possess acute toxicity in the experimental mice.

In-vivo cytotoxicity tests in lung/stomach cancer mice models

Dead mice evaluation

First, the number of dead mice during the experiment was assessed and is shown in Table 3. For this, the initial number of mice was 85 and the total number of dead mice was 35, accounting for 41.18%. Regarding the timeline, the number of dead mice from week 1 to week 4 was 11 (12.94%), which was because some mice had not adapted to environmental conditions and were immunocompromised due to DMBA, and from week 5 to week 24 was 24 (28.24%), which was because of the suffocation when being taken the drug and impaired immunity with reduced weight.

Mice weight investigation

Second, the mice weight changes during the testing period were recorded (Fig. 2). In the beginning, the mean weights of these groups were not significantly different from each other (p > 0.05). After the testing period, the overall weights of all groups increased. From week 6 to week 8, the mice's weight of

		Stomach canc	er	Lung cáncer				
Group	Mice with tumors (%)	Tumor number	Tumor volume (mm ³)	Mice with tumors (%)	Tumor number	Tumor volume (mm ³)		
1	0	0	0	0	0	0		
2	100*	$3.50 \pm 0.63^{**}$	$6.38 \pm 0.94^{**}$	90.00*	$4.22\pm0.22^*$	$5.60\pm0.27^{\ast}$		
3a	87.50	$1.71\pm0.29^{\scriptscriptstyle\#}$	$1.43 \pm 0.17^{\text{###}}$	87.50	$1.86\pm0.34^{\scriptscriptstyle\#}$	$1.55\pm0.20^{\scriptscriptstyle\#}$		
3b	77.80	$1.57\pm0.30^{\#}$	$1.21\pm 0.11^{\#\!\#\!}$	66.67	$1.67\pm0.21^{\scriptscriptstyle\#}$	$1.31\pm0.93^{\scriptscriptstyle\#}$		
4	88.90**	$3.38 \pm 0.53^{**}$	$6.13 \pm 0.59^{**}$	88.89*	$4.00\pm0.27*$	$5.55\pm0.29^{\ast}$		
5a	70.00	$1.43\pm0.20^{\scriptscriptstyle\#}$	$1.18\pm 0.16^{\text{\#}}$	80.00	$1.50\pm0.27^{\scriptscriptstyle\#}$	$1.24\pm0.19^{\scriptscriptstyle\#}$		
5b	62.50	$1.40\pm0.25^{\scriptscriptstyle\#}$	$1.06 \pm 0.07^{\text{###}}$	50.00	$1.25\pm0.25^{\scriptscriptstyle\#}$	$1.11\pm0.05^{\scriptscriptstyle\#}$		
6a	80.00	$1.38\pm 0.18^{\text{\#}}$	$1.08\pm 0.08^{\text{###}}$	90.00	$1.44\pm0.18^{\scriptscriptstyle\#}$	$1.17\pm0.08^{\scriptscriptstyle\#}$		
6b	57.10#	$1.25\pm0.25^{\scriptscriptstyle\#}$	$0.98 \pm 0.13^{\text{\#}}$	37.50###	1.00#	$0.94\pm0.05^{\scriptscriptstyle\#}$		

Table 5. Macroscopic results of the mice's stomach and lung from week 5 to 24. Results are
expressed as mean \pm SD.

 $p^{*} < 0.001$, $p^{**} < 0.01$ when compared to group 1; $p^{*} < 0.001$, $p^{**} < 0.01$, $p^{**} < 0.05$ when compared to group 2

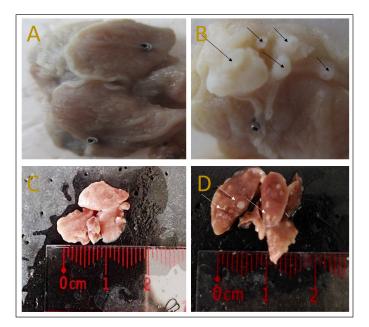


Figure 3. Macroscopic analyses of (A) normal stomach, (B) cancerous stomach, (C) normal lung, and (D) cancerous lung.

groups 2, 3a, 4, 5a, 5b, and 6a decreased and was significantly less than that of group 1 (p < 0.05) because mice in these groups got immunocompromised by DMBA. In addition, although mice in groups 3b, 5b, and 6b were taking CSD-loaded floating tablets and 5-FU from week 6, the experimental time was short, thus, there were no significant differences in mice's weight between these groups, similar to the report of Da *et al.* [23]. From week 12 to week 24, the weight gain in groups 3a, 3b, 5a, 5b, 6a, and 6b was not significantly different from that in group 1 because mice adapted to the high-fat diet in corn oil, and the immune system gradually recovered. Moreover, the mice's weights increased nonlinearly between weeks, which could be due to the biological fluctuations of the mice. Lastly, a weak/inadequate correlation between the mice weight changes and tumor formation was noted (0 < r < 0.2), suggesting that the tumor/cancer formation and progression were not well connected to the mice weights.

Fat accumulation determination

Third, the fat accumulation of mice is expressed through the fat index (Table 4). Fat accumulation is the phenomenon of increased triglycerides and lipids because tumor cells synthesize these molecules by themselves and partly obtain them from their host [24]. Compared with group 1, mice in group 2 and group 4 possessed significantly higher fat accumulation (p < 0.05), indicating the tumor's hyperactivity after using DMBA, in agreement with a previous study [24]. There was no statistical difference between group 2 and 4 (p > 0.05), suggesting that the floating tablet excipients did not affect the mice fat accumulation. Interestingly, when being treated with CSD-loaded floating tablets and/or 5-FU, the mice's lipid levels reduced significantly (groups 3a, 3b, 5a, 5b, 6a, and 6b) compared with the disease control group (group 2) (p < 0.05), indicating the treatment efficacy. Curcumin has an anti-inflammatory effect on mice tumors, which helps reduce fat accumulation.

Macroscopic analyses

Fourthly, the macroscopic analyses were conducted and reported (Table 5 and Fig. 3). For both cancer types of stomach and lung, none of the mice in group 1 had tumors, and the percentages of tumors in group 2 and 4 were >85%. The results were similar to the study of Azuine and Bhide [18] and Huang et al. [20], which demonstrated that corn oil did not cause tumors and DMBA initiates tumor formations in mice with a percentage of approximately 100%. The difference between group 4 and 2 was not statistically significant (p >0.05), showing that the floating tablet excipients had no effects on tumor formation and progression. Compared to group 2, in terms of the tumor percentage, the mean number of tumors, and the mean tumor volume, the treatment groups (3a, 3b, 5a, 5b, 6a, and 6b) possessed significantly lower values, indicating that the CSD loaded floating tablets and/or 5-FU could treat the stomach/lung cancer in mice, in agreement with previous studies

Table 6. Microscopic results of the mice's stomach and lung fromweek 5 to 24. Results are expressed as mean \pm SD.

	Stoma	ach	Lung			
Group	Noncancerous lesions (%)	Cancerous tumors (%)	Noncancerous lesions (%)	Cancerous tumors (%)		
1	0	0	0	0		
2	25.00	75.00*	10.00	80.00^{*}		
3a	12.50	37.50	25.00	37.50		
3b	11.11	33.33###	22.22	33.33###		
4	11.11	77.78**	11.11	77.78**		
5a	30.00	30.00###	30.00	30.00###		
5b	37.50	12.50##	25.00	12.50##		
6a	20.00	30.00###	30.00	30.00###		
6b	14.29	14.29##	25.00	12.50##		

*p < 0.001, **p < 0.01 when compared to group 1; ##p < 0.01, ###p < 0.05 when compared to group 2

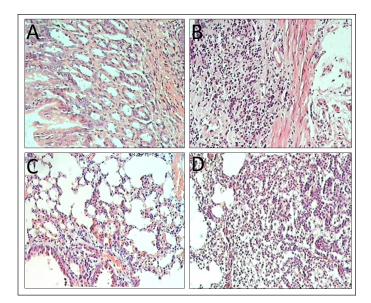


Figure 4. Microscopic analyses of (A) normal stomach, (B) cancerous stomach, (C) normal lung, and (D) cancerous lung.

[17,21,23,25]. Interestingly, as compared with groups 3 (CSDloaded floating tablet treatment only) and 5 (5-FU treatment only), the combination treatment (group 6) yielded a better outcome, suggesting the complementary action of curcumin with a synthetic drug (5-FU) in stomach/lung cancer treatment. Finally, concerning two groups that were being treated from week 6 to week 21 (subgroups a and b), the tumor percentage, the mean number of tumors, and the mean tumor volume in the group being treated from week 21 were higher than that in the group being treated from week 6 (3a > 3b, 5a > 5b, and 6a > 6b). Conclusively, using curcumin and/or 5-FU at the early stage would bring better anti-tumor effects than treating at a late stage.

Microscopic analyses

Finally, the microscopic analyses were reported in Table 6 and Figure 4. For both stomach and lung cancers,

all mice in group 1 did not have any lesions. On the other hand, group 2 possessed lesions, of which more than 70% of the lesions were cancerous, suggesting the DMBA could trigger cancers in mice. The noncancerous lesions were due to acute inflammation and congestion. The cancer lesions in group 4 were not significantly different compared to those in group 2 (p > 0.05), indicating the blank floating tablets had no cytotoxicity on the growth of cancer cells. Notably, the cancerous tumor numbers in group 3a were not different from that in group 2 (p > 0.05), whereas these numbers were significantly decreased in groups 3b, 5a, 5b, 6a, and 6b. This confirms the fact that curcumin could potentially inhibit the growth of cancer cells when used in the early stage, but might not in the late stage. Our results were similar to that of a previous report by Sintara *et al.* [26], which used a curcumin dose of 200 mg/kg in mice for 20 weeks in their study. The results determined that the cancer rate in the curcumin-treated group was reduced by 50% compared to that in the untreated group (100%). Furthermore, although the anti-cancer effects of curcumin (group 3) were lower than that of the standard drug 5-FU (group 5), it could well complement 5-FU to significantly enhance the drug efficacy (group 6).

CONCLUSION

This study investigated the stomach/lung anti-cancer effects of curcumin in a complete dosage form, the CSD-loaded floating tablets, in the *in-vivo* setting in mice. The results showed that the tablets could significantly inhibit the growth of stomach/lung cancer cells and reduce the cancer rate. In addition, the CSD-loaded floating tablets could potentiate the standard chemotherapeutic drug 5-FU efficacy in treating stomach/lung cancers, helping reduce the dose of 5-FU and limiting its toxicity in patients. Moreover, the study also proved that using curcumin in the early stages would help reduce the cancer rate better than using it in the late stages. In summary, the CSD-loaded floating tablets could be a novel dosage form for stomach/lung cancer treatments in the future.

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AUTHOR CONTRIBUTIONS

Concept: DTMH, DTP; design: DTP; literature search: DTMH, DTP; experimental studies: DTMH, VDT, SP, DTP; data analysis: DTMH, SP, DTP; statistical analysis: VDT; manuscript preparation, manuscript editing and manuscript review: DTMH, VDT, SP, DTP.

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CONFLICTS OF INTEREST

The authors report no financial or any other conflicts of interest in this work

ETHICAL APPROVALS

The research ethics were approved by the University of Medicine and Pharmacy at Ho Chi Minh City, Vietnam, code 1454/QD-DHYD-SDH.

DATA AVAILABILITY

All data generated and analyzed are included in this research article.

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