

Deciphering the intricate role of mTOR signaling and autophagy in Parkinson's disease and therapeutic prospects

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ABSTRACT

The mammalian/mechanistic target of rapamycin (mTOR) has a prominent role in many neurodegenerative diseases such as Alzheimer's disease and Parkinson's disease (PD). mTOR and autophagy have a definitive role in the pathogenesis of PD. mTOR is a protein kinase that regulates various cellular processes, including cell survival and protein synthesis. mTOR also has other beneficial impacts, such as maintaining glucose homeostasis, regulating muscle mass, and increasing mitochondrial functions. Many evidence suggests that in animals which are induced by Parkinson's, overexpression of mTOR and its components are observed. mTOR complex 1 controls protein synthesis, whereas mTOR complex 2 controls cell survival and cytoskeleton organization regardless, it plays a crucial function in autophagy inhibition. Inhibition of autophagy is one of the reasons for the accumulation of α -synuclein, which pave the way for the development of PD. The role of mTOR is controversial as mTOR can produce either neuroprotection or neurotoxic effects depending upon the target in which it is acting. In this review, we shall define mTOR, its role, its involvement in autophagy, and potential PD treatment by targeting mTOR and its signaling components such as Unc-51-like kinase 1 and adenosine monophosphate-activated protein kinase. Furthermore, we also summarize the dual role of mTOR.

INTRODUCTION

Parkinson's disease (PD) is a neurodegenerative disorder characterized by losing midbrain dopaminergic neurons in the substantia nigra [1]. It is estimated that PD affects 1%–2% of the geriatric population. The threat of the economic burden of PD in the national healthcare system continues to rise [2]. However, treatment for Parkinson's is available, but it provides only symptomatic relief and does not stop progression because the exact mechanism is complex and unknown. Drugs were used to slow down the passage of the degeneration of neurons [3]. Understanding the mechanism of disease progression can help us target and get the right treatment choice. Many recent pieces of evidence indicate that

mechanistic target of rapamycin (mTOR) has a critical role in PD pathogenesis.

The mammalian/mTOR is a serine–threonine kinase. It controls many important functions of mammalian cells, such as cell survival and protein synthesis [4]. During the early 2000s, neuroscientists began their interest in mTOR targets. 4E-binding proteins and p70 ribosomal S6 protein kinase 1 are initially studied [5]. Soon after identifying mTOR's role in neuronal morphogenesis, survival, and differentiation, the target became popular, and many scientists observed its role in different diseases such as PD and Alzheimer's disease (AD). The list of physiological conditions and neuropathologies associated with mTOR rapidly increased, but a thorough knowledge of mTOR modulation and its cellular effectors in neurons remained elusive. Autophagy, translation, cell signaling, transcription, and cytoskeleton dynamics are all influenced by changes in the mTOR activity [6]. According to new research, the overexpression of mTOR is linked to the pathogenesis of PD [7,8]. As a result, mTOR might be one of the possible treatment targets for PD [9]. mTOR activity is quite controversial. It has

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different actions in different pathological conditions [10]. The mTOR signaling system, its positive and negative functions in pathological conditions, its regulation, and its participation in autophagy are all discussed in this article. We also discuss possible mTOR modulators that can be used in treating PD.

We conducted a review of the literature in PubMed. Only articles in the English language are selected. The papers are reviewed until December 2021. The following terms are used to search articles: PD and each of the following terms or a combination of the following terms: mTOR, PD, akt, α -synuclein accumulation, Unc-51-like kinase 1 (ULK1), energy deficit mTOR, TSC1/2, autophagy, protein synthesis, cell cycle, inflammation, neuroprotective, toxicities mTOR, and small molecules (Fig. 1).

Components and structure of mTOR

mTOR or mammalian/mTOR is a serine/threonine protein kinase. It involves various cell functions such as gene transcription, protein synthesis, cell metabolism, survival, and proliferation [11]. The multidomain protein mTOR is found in two multiprotein complexes: mTOR complex 1 (mTORC1) and mTOR complex 2 (mTORC2) [12]. mTORC1 controls protein synthesis, whereas mTORC2 controls cell survival and cytoskeleton organization [13].

Each complex contains mTOR protein, mammalian lethal with SEC13 protein 8 (mLST8), DEP domain-containing mTOR interacting protein (DEPTOR), Tel two-interacting protein1, and telomere maintenance 2 (Tel2) [14,15]. Two proteins are specific to mTORC1, the regulatory-associated protein of mTOR (Raptor) and proline-rich AKT1 substrate 40 kDa (PRAS40) [16,17]. In these complexes, DEPTOR and PRAS40 can negatively regulate mTORC1 activity [18]. mTORC1 plays a major role in regulating cell growth and proliferation and senses stimuli such as insulin level and amino acid-like Leucine [19,20]. In addition, it is involved in protein synthesis, lipid metabolism, and autophagy [21,22].

mTORC2 contains a Rapamycin-insensitive companion of mTOR, mammalian stress-activated protein kinase-interacting protein1 (mSIN1), and protein observed with Rictor [23], mLST8, and Tti1/Tel2. Akt is an important substrate of mTORC2. Phosphorylation of Akt induces phosphorylation of mSIN1, which increases mTORC2 activity [24]. mTORC2 controls the remodeling of cytoskeleton and electrolyte homeostasis [25].

mTOR signaling pathways

mTORC1 signaling pathway

Insulin-like growth factor 1 (IGF-1) activates mTOR. IGF starts the conversion of Ras guanosine 5'-triphosphate (GTP) to Ras guanosine diphosphate (GDP). This conversion leads to the cascade of small molecule activation, which includes Ras, Mek, Erk, and Rsk. Erk and Rsk inhibit the activation of TSC1/2. TSC1/2 complex is an essential regulator in this process [26]. TSC1/2 activates the conversion of GTPase Rheb to GDPase Rheb, an inactive form. This GDP-bound state negatively regulates the mTOR pathway [27]. Other factors, such as inflammation and hypoxia, can also lead to mTOR activation.

In response to the cellular energy deficiency, LKB1 is activated. This activated LKB1 increases the adenosine monophosphate (AMP)/adenosine triphosphate (ATP) ratio. This elevated level of AMP leads to phosphorylation of AMP-activated protein kinase (AMPK), which consecutively activates TSC1/2. The activation of TSC1/2 leads to an increase in the GAP activity of the TSC1/2 complex and inhibits mTORC1 activity [28]. It is further noted that hypoxia can also inhibit mTORC1 activity by the increase in regulated in development and DNA damage responses (REDD1) expression [29]. Inflammation can also lead to the inhibition of TSC1/2 by activating Ikk β .

The activated mTORC1 plays its role in protein regulation by two possible mechanisms. One by phosphorylation of ribosomal protein S6 kinase beta-1 (S6K1) and eukaryotic initiation factor 4E-binding protein 1 (4E-BP1) [30]. S6K1 is phosphorylated and activated by mTORC1, which again phosphorylates and activates S6 (a component of the S40 ribosomal subunit). S6K1 also activates several small molecules such as CBP80, Tripartite motif-containing protein-24 (TIF1A), SKAR, and eIF4B, and inhibits eukaryotic elongation factor 2 kinase (eEF2K) and PDCD4. S6K1 inhibits eEF2K by phosphorylation, thus inhibiting the translation and elongation of proteins. Phosphorylation of PDCD4 leads to the inhibition of eukaryotic initiation factor-4A, which is responsible for translation initiation [31]. S6K1 phosphorylates and activates SKAR and CBP80. This activation leads to mRNA biogenesis [32]. S6K1 also activates TIF1A, elevating rRNA expression through interaction through POL1.

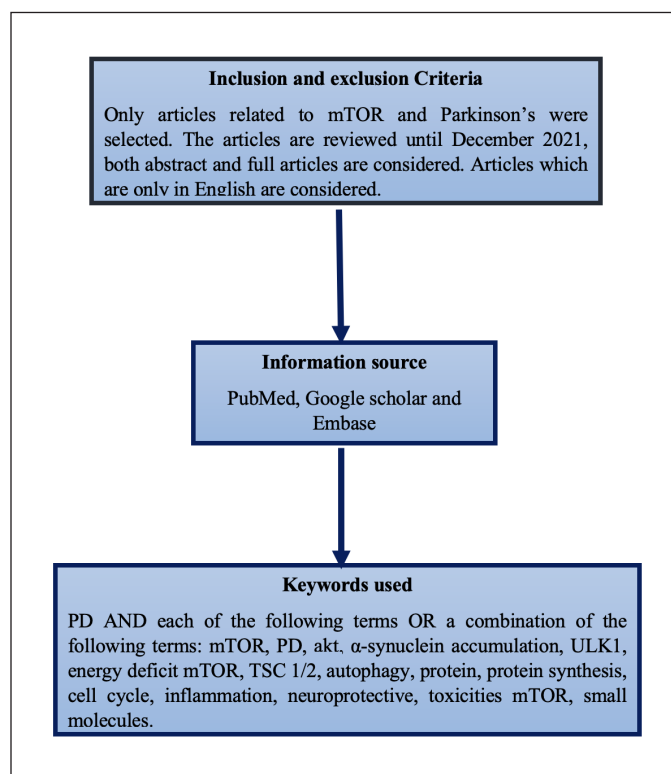


Figure 1. Methodology.

Another pathway by which mTOR regulates protein synthesis is by 4E-BP1. By blocking the link between 4E-BP1 and eukaryotic translation initiation factor 4E, active mTORC1 phosphorylates and inactivates 4E-BP1, a regulator of mRNA translation (eIF-4E). These signaling pathways lead to protein and lipid synthesis and inhibit autophagy and lysosome biogenesis [33]. Autophagy is induced by the ULK1 complex when mTOR is inhibited. Hence, mTOR negatively regulates autophagy [34].

mTORC2 signaling pathway

mTORC2 also responds to growth factors like mTORC1, through the PI3K pathway. However, in the case of mTORC2, TSC1/2 activates mTORC2 rather than suppressing it like mTORC1. Some *in-vitro* studies reported that removal of TSC1/2 leads to loss of mTORC2 activity [35]. mTORC2 exerts its role by activating protein kinases A, G, and C which consist of Akt, serum/glucocorticoid regulated kinase 1 (SGK1), and protein kinase C alpha (PKCα) [36]. mTORC2 regulates cytoskeleton remodeling by phosphorylation of PKCα and ion transport by phosphorylation of SGK1 [37]. Akt paves the way for cell survival and metabolism [38]. Downstream of the Akt pathway includes activation of murine double minute 2 (MDM2) and inhibition of Forkhead box O (FOXO) and PRAS40. MDM2 is responsible for cytoskeleton organization, and FOXO inhibits proliferation and cell cycle arrest. PRAS40 regulates mTORC1 by directly inhibiting mTORC1 [39]. Figure 2 represents the signaling pathway of mTORC1 and mTORC2.

mTOR-double-edged sword

mTOR is vital in many neurodegenerative diseases such as AD and PD. In recent years, the role of mTOR in Parkinson's has been studied widely. However, the role of mTOR in PD remains controversial [40]. Many studies have

shown evidence of both neuroprotective and neurotoxic effects. For example, overexpression of mTOR in toxin-induced PD can partially prevent neuronal loss [41]. In contrast, another study indicates that PD toxins suppress mTOR signaling, which leads to a decrease in cell viability [42]. Here, we discuss the neuroprotective and neurotoxic effects of mTOR.

Neuroprotective effects of mTOR

The stimulation of mTOR in dopaminergic neurons may result in axon regrowth [43]. Deletion of phosphatase and tensin homolog deleted on Chromosome 10 activates mTOR and protects dopaminergic neurons by promoting Akt signaling [44]. REDD1 (regulated in development and DNA damage responses) inhibition causes mTOR activation and provides neuroprotection in cellular models [45]. Memory is similarly affected by mTOR. Inhibition of mTOR by rapamycin leads to memory deficits in PD models [46]. mTOR interacts with Raptor and produces signals for cell cycle regulation. The activation of mTOR significantly influences neurogenesis, particularly dendritic formation. Cell development needs average cell growth and advancement, governed by the mTOR pathway [47]. mTOR induces translation through direct or indirect phosphorylation in the presence of signals such as growth hormones and energy deprivation. mTOR also has other beneficial impacts, such as maintaining glucose homeostasis [48], regulating muscle mass [49], and increasing mitochondrial functions [50] (Fig. 3).

Neurotoxicity of mTOR

The neurotoxic effects of mTOR are mostly related to the overexpression of mTOR signaling. An elevated amount of mTOR is seen in the post-mortem brains of people with PD. Maneb and paraquat are the major environmental risk factors of PD. Among them, paraquat has been shown to drastically elevate

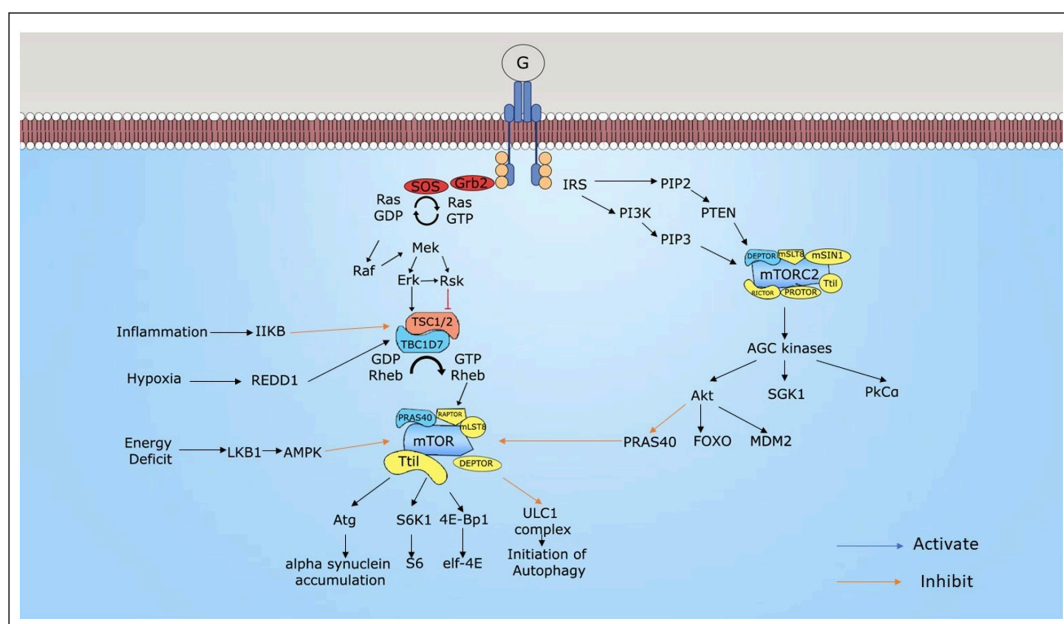


Figure 2. Signaling pathway of mTORC1 and mTORC2.

Role of mTOR in neurodegenerative diseases

[illegible]

Figure 3. Neuroprotective and neurotoxic effects of mTOR.

In a study, rapamycin which is a potent mTOR inhibitor, promotes autophagy in both *in vivo* and *in vitro* [66]. This exhibits the relationship between autophagy and mTOR inhibition. The inhibitors of mTOR are widely searched since they help in the inhibition of protein synthesis and induction of autophagy. Rapamycin (Sirolimus) is the first rapalog of this class. Everolimus and tacrolimus came into existence after that [67]. Despite blocking protein translation and autophagy induction, long-time exposure to these drugs causes tissue damage and impairment of metabolism. Small compounds that specifically alter the function of proteins governing autophagy downstream of mTORC1 signaling might be developed to trigger this process in a more targeted manner (Fig. 4).

Autophagy is the highly conserved mechanism through which the misfolded proteins, and damaged organelles are removed from the body to maintain homeostasis [68]. Autophagy exerts its action through the lysosomal degradation pathway, thereby degrading the catabolic contents [69]. Autophagy is mediated through microautophagy, macroautophagy, and chaperone-mediated autophagy. In macroautophagy, the cytoplasmic portions are wrapped within the autophagosome and carried to lysosomes for bulk degradation [70]. Microautophagy includes autophagic tubes directly engulfing the cytoplasmic components [71]. Both micro and macroautophagy engulf cytoplasmic components through a selective and non-selective

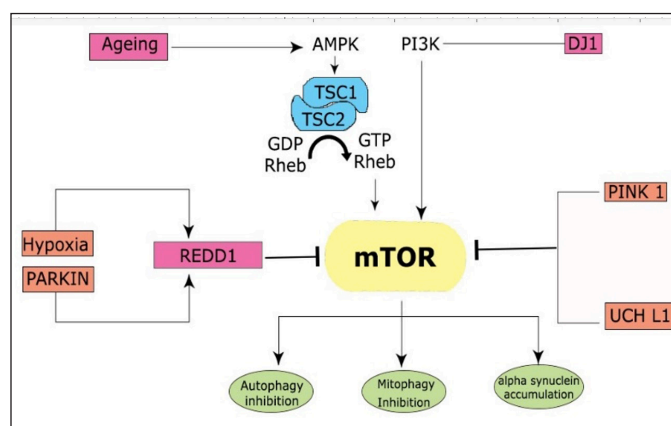


Figure 4. The various factors that modulate mTOR signaling include aging, REDD1, PARKIN, PINK, UCHL1, and DJ1, which lead to the onset of PD. PINK and PARKIN cause mitophagy through mTOR. REDD1, UCHL1, and PINK inhibit mTOR, thereby reducing protein translation and leading to neuronal death. Aging inhibits AMPK, which activates mTOR and inhibits autophagy. DJ1 activates mTOR through the PI3K pathway resulting in alpha-synuclein accumulation and inhibition of autophagy.

mechanism. In contrast, Chaperone-mediated autophagy exerts degradation in a non-specific manner, i.e., molecule-by-molecule fashion [72]. The major signaling pathway which controls autophagy is mTOR [73]. mTOR exerts its role in the various stages of autophagy, and its activity is tightly regulated by a complex interplay between stimulatory and inhibitory signals [74]. The various signaling pathways through which mTOR regulates autophagy are discussed below:

ULK1 signaling pathway

The downstream of mTORC1 contains the ULK1 complex, which consists of ATG101, ATG13, and focal adhesion kinase family interacting protein of 200 kD. mTORC1 activates and phosphorylates ULK1 [75]. This phosphorylation leads to the inhibition of interaction with AMPK and inhibits autophagy. On cellular stress, ULK1 is phosphorylated by AMPK, which further leads to the induction of autophagy. Hence, mTORC1 negatively regulates autophagy by the ULK1 complex [76].

TFEB signaling pathway

Transcription factor EB (TFEB) is a part of the bHLH leucine-zipper transcription factor family that regulates autophagy and lysosomal gene expression [77]. Normally inhibition of mTORC1 results in the release of TFEB, which releases transcriptional factors. These transcriptional factors cause autophagy and lysosome biogenesis [78].

Vacuolar protein sorting 34 (VPS34) complex signaling pathway

Vacuolar protein sorting 34 is another protein through which autophagy induction occurs. VPS34 consists of two complexes: Complex 1 and Complex 2. Complex 1 consists of VPS34, VPS15, beclin 1, and ATG14. Complex 2 consists of VPS34, VPS15, beclin 1, and UVRAG [79]. mTORC1 directly inhibits VPS34 by phosphorylating ATG14 at various sites. Autophagy might be improved by mutating these sites, which are resistant to mTOR inhibition [80].

Autophagic lysosome reformation (ALR)

Lysosomes are repurposed from autolysosomes at the end of autophagy by a process known as ALR, which comprises proto-lysosome tubule formation, elongation, and termination [81]. In short-term food deprivation, mTOR is inhibited, and in nutrient-rich conditions, mTOR is activated. Also, during prolonged starvation, mTOR is reactivated by a negative feedback loop of autophagy. For this reactivation, autophagic lysosomes have to be degraded. Spinster, a lysosomal efflux protein, is a regulator of ALR. Prolonged starvation leads to defective spinsters. mTOR reactivation failed due to the defective spinster [82]. The general amino acid control pathway is activated by starvation, which leads to the overexpression of amino acid transporters in the plasma membrane, increasing amino acid absorption and contributing to mTOR reactivation [83].

Lysosomal acidification and autophagy

Lysosomes are crucial organelles involved in the degradation of cellular waste. Lysosomal acidification is

facilitated by proton pumps, predominantly the vacuolar H⁺-ATPase (V-ATPase), which actively pumps protons into the lysosomal lumen. The acidic pH inside lysosomes is crucial for the activation of hydrolytic enzymes, enabling efficient degradation of cellular components during autophagy [84].

The interplay between mTOR and lysosomal acidification

An intriguing feedback loop exists between mTOR and lysosomal acidification. Amino acids, critical activators of mTORC1, are transported into lysosomes, where they play a pivotal role in recruiting and activating mTORC1 at the lysosomal surface. The V-ATPase, responsible for lysosomal acidification, is central to this process [85]. Understanding the interplay between mTOR and lysosomal acidification holds significant implications for cellular health and disease. Dysregulation of either process can lead to disruptions in autophagy, compromising cellular homeostasis and contributing to the pathogenesis of various diseases, such as cancer, neurodegenerative disorders, and metabolic conditions [86]. In PD, proper lysosomal acidification is necessary for the optimal function of lysosomal enzymes [87]. Genetic investigations have unveiled a significant association between lysosomal function and PD. Several autosomal dominant and recessive genes linked to PD, as well as various genetic risk factors, encode proteins involved in lysosomal, autophagic, and endosomal pathways. Mutations in these PD-associated genes can lead to lysosomal dysfunction, and considering that α -synuclein degradation is primarily reliant on lysosomal processes, this impairment can hinder α -synuclein turnover. Consequently, this disruption contributes to elevated intracellular levels of α -synuclein, facilitating its accumulation and subsequent aggregation, among other consequences.

Feedback loop-AMPK, ULK1, and mTOR

The main reasons for many neurodegenerative diseases are the overproduction of abnormal proteins and autophagy dysregulation [88]. Much evidence suggests that mTOR signaling is overactivated in PD patients. Overexpression of this mTOR will lead to autophagy inhibition [89]. The activity of mTOR is reduced in animals that cause autophagy induction. Hence, the direct correlation between mTOR and autophagy inducer ULK1 is studied. On the other hand, during energy deficit conditions, AMPK is activated, which inhibits mTORC1, thereby activating ULK1 [90]. This AMPK is further inactivated by mTOR. These shreds of evidence suggest that mTOR, ULK1, and AMPK have a direct relationship. A feedback loop is a mechanism that restores the body to its original state. All stable system has a feedback mechanism to retain control internally. Protein regulator mTOR and autophagy regulator ULK1 and AMPK have a direct feedback link. Two possible cycles exist between them. AMPK activates ULK1 as a process of autophagy induction. This activation leads to direct inhibition of AMPK by ULK1 as a feedback loop. This feedback loop leads to delayed inhibition of ULK1, which results in oscillation of autophagy induction [57]. The double negative-feedback loop in which AMPK directly phosphorylates the mTOR complex and inhibits it. This inhibition results in the inhibition of ULK1, followed by AMPK inhibition [91].

We know that mTOR is activated in nutrient-rich conditions and inhibits autophagy by phosphorylating ULK1. On starvation, phosphorylated AMPK inhibits mTOR, thereby activating autophagy. The factors that activate AMPK are LKB1, CaMKK β , TAK1, DNA damage, and a decrease in ATP/AMP ratio [76]. On the other hand, ULK1 is activated by AMPK and inhibited by mTOR. ULK1 also counteracts and inhibits mTOR [92]. mTOR inhibition cannot be permanent. Prolonged inhibition of mTOR leads to reactivation [93]. A recent study suggests that AMPK alone cannot induce autophagy when both mTOR and ULK1 are inhibited [94]. AMPK remains dephosphorylated until mTOR is activated [95]. Therefore, the balance between autophagy and mTOR is challenging to achieve. Currently, mTOR, ULK1, and AMPK gained an attractive target for the treatment of PD. These feedback loops help maintain homeostasis and regulate the balance between protein synthesis and clearance. Hence, the complex interplay of the feedback loops has to be explored, and a better understanding of their autophagy modulation will help us treat PD.

Potential PD treatment by targeting mTOR and autophagy

Targeting mTOR and autophagy has emerged as a potential therapeutic strategy for PD due to their critical roles in cellular homeostasis and protein degradation. Dysregulation of mTOR and autophagy pathways has been implicated in the accumulation of toxic protein aggregates, like alpha-synuclein, and the degeneration of dopaminergic neurons, hallmark features of PD [96]. Here are some approaches for potential PD treatment by targeting mTOR and autophagy:

mTOR inhibitors

Rapamycin and its analogs (rapalog) are mTOR inhibitors that have shown promise in preclinical studies for their neuroprotective effects in PD models. These inhibitors promote autophagy and reduce the accumulation of toxic protein aggregates [97]. However, further research is needed to optimize their dosing and delivery to the brain while minimizing side effects. Small molecules that control autophagy and mTOR can be targeted, which assists in mTOR and autophagy balance (Table 1).

Activators of autophagy

Enhancing autophagy through pharmacological interventions has been explored as a therapeutic strategy for PD. Certain compounds, such as trehalose [98] and lithium [99,100], have been shown to induce autophagy and promote the clearance of toxic proteins. Clinical trials to evaluate their efficacy and safety in PD patients are ongoing.

Lysosomal enzyme modulators

Enhancing lysosomal function and promoting efficient degradation of toxic protein aggregates is another potential approach. Small molecules that modulate lysosomal enzymes, such as glucocerebrosidase (GBA), have shown promise in preclinical studies and may hold therapeutic potential for PD with GBA mutations [101].

Table 1. The action of small molecules and drugs on various phases of autophagy.

| Autophagy phase | Small molecules | Drugs that modulate autophagy |
|-----------------|--------------------------|-------------------------------|
| Initiation | \uparrow AMPK | Metformin, [102] |
| | | Resveratrol, [103] |
| | | Rosuvastatin, [104] |
| | | Temozolomide [105] |
| | \downarrow mTOR | Rapamycin, [106] |
| Nucleation | \uparrow ULK1 | Corynoxine [107] |
| | \uparrow Beclin1 | BL-918 [108] |
| | | Isorhynchophylline, [109] |
| | | Glycyrrhizic acid [110] |
| | \downarrow IMP ase | Valproate, [111] |
| Elongation | \uparrow TFEB | Carbamazepine [112] |
| | | Curcumin, [113] |
| | \uparrow GC case | Trehalose [114] |
| | | Ambroxol [115] |
| Degradation | \uparrow ERR- α | XCT 790 [116] |
| | \downarrow c-ABL | Synphilin-1 [117] |

CONCLUSION

Due to its dual function of neuroprotective and neurotoxic effects, the role of mTOR in neurodegenerative diseases is still debated. Because mTOR is critical for protein synthesis and cell survival processes, it is worth noting that mTOR inhibition causes autophagy to activate, which may play a role in neuroprotection. Long-term inhibition, however, can trigger the feedback loop and mTORC1, preventing autophagy. As a result, mTOR is a double-edged sword. It can play both neuroprotective and neurotoxic. Hence, the signaling between mTOR and autophagy has to be balanced. If the balance between mTOR and autophagy can be achieved, mTOR might be a possible target for the therapy of PD.

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LIST OF ABBREVIATIONS

AD, Alzheimer’s disease; ALR, Autophagic lysosome reformation; AMPK, Adenosine monophosphate-activated protein kinase; DEPTOR, DEP domain-containing mTOR interacting protein; eEF2K, Eukaryotic elongation factor 2 kinase; IGF-1, Insulin-like growth factor 1; mSIN1, Mammalian stress-activated protein kinase-Interacting protein 1; mTOR, Mammalian/mTOR; mTORC1 mTOR complex 1; mTORC2, mTOR complex 2; PD, Parkinson’s disease; PKC α , Protein kinase C alpha; Raptor, Regulatory-associated protein of mTOR; REDD1, Regulated in development and DNA damage responses; SGK1, Serum/glucocorticoid regulated

kinase 1; Tel2, Telomere maintenance 2; TIF1A, Tripartite motif-containing protein-24; ULK1, Unc-51-like kinase 1.

AUTHOR CONTRIBUTION

TLN, SKM, DT, AB, YMT, SK, and SNM made a significant contribution to the work reported, whether that is in the conception, execution, or the acquisition, analysis, or interpretation of data, or all the areas; took part in drafting, revising, or critically reviewing the article; and gave final approval of the version to be published. All have read and agreed to the published version of the manuscript.

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CONFLICTS OF INTEREST

The authors report no financial or any other conflicts of interest in this work.

CONSENT TO PARTICIPATE

It is a review article, thus it is not applicable

ETHICAL APPROVALS

This study does not involve experiments on animals or human subjects.

DATA AVAILABILITY

The data that support the findings of this study are available in standard research databases such as PubMed, Science Direct, or Google Scholar, and/or on public domains that can be searched with either key words or DOI numbers.

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