



# Deciphering the intricate role of mTOR signaling and autophagy in Parkinson's disease and therapeutic prospects

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## ABSTRACT

The mammalian/mechanistic target of rapamycin (mTOR) has a prominent role in many neurodegenerative diseases such as Alzheimer's disease and Parkinson's disease (PD). mTOR and autophagy have a definitive role in the pathogenesis of PD. mTOR is a protein kinase that regulates various cellular processes, including cell survival and protein synthesis. mTOR also has other beneficial impacts, such as maintaining glucose homeostasis, regulating muscle mass, and increasing mitochondrial functions. Many evidence suggests that in animals which are induced by Parkinson's, overexpression of mTOR and its components are observed. mTOR complex 1 controls protein synthesis, whereas mTOR complex 2 controls cell survival and cytoskeleton organization regardless, it plays a crucial function in autophagy inhibition. Inhibition of autophagy is one of the reasons for the accumulation of  $\alpha$ -synuclein, which pave the way for the development of PD. The role of mTOR is controversial as mTOR can produce either neuroprotection or neurotoxic effects depending upon the target in which it is acting. In this review, we shall define mTOR, its role, its involvement in autophagy, and potential PD treatment by targeting mTOR and its signaling components such as Unc-51-like kinase 1 and adenosine monophosphate-activated protein kinase. Furthermore, we also summarize the dual role of mTOR.

## INTRODUCTION

Parkinson's disease (PD) is a neurodegenerative disorder characterized by losing midbrain dopaminergic neurons in the substantia nigra [1]. It is estimated that PD affects 1%–2% of the geriatric population. The threat of the economic burden of PD in the national healthcare system continues to rise [2]. However, treatment for Parkinson's is available, but it provides only symptomatic relief and does not stop progression because the exact mechanism is complex and unknown. Drugs were used to slow down the passage of the degeneration of neurons [3]. Understanding the mechanism of disease progression can help us target and get the right treatment choice. Many recent pieces of evidence indicate that

mechanistic target of rapamycin (mTOR) has a critical role in PD pathogenesis.

The mammalian/mTOR is a serine–threonine kinase. It controls many important functions of mammalian cells, such as cell survival and protein synthesis [4]. During the early 2000s, neuroscientists began their interest in mTOR targets. 4E-binding proteins and p70 ribosomal S6 protein kinase 1 are initially studied [5]. Soon after identifying mTOR's role in neuronal morphogenesis, survival, and differentiation, the target became popular, and many scientists observed its role in different diseases such as PD and Alzheimer's disease (AD). The list of physiological conditions and neuropathologies associated with mTOR rapidly increased, but a thorough knowledge of mTOR modulation and its cellular effectors in neurons remained elusive. Autophagy, translation, cell signaling, transcription, and cytoskeleton dynamics are all influenced by changes in the mTOR activity [6]. According to new research, the overexpression of mTOR is linked to the pathogenesis of PD [7,8]. As a result, mTOR might be one of the possible treatment targets for PD [9]. mTOR activity is quite controversial. It has

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different actions in different pathological conditions [10]. The mTOR signaling system, its positive and negative functions in pathological conditions, its regulation, and its participation in autophagy are all discussed in this article. We also discuss possible mTOR modulators that can be used in treating PD.

We conducted a review of the literature in PubMed. Only articles in the English language are selected. The papers are reviewed until December 2021. The following terms are used to search articles: PD and each of the following terms or a combination of the following terms: mTOR, PD, akt,  $\alpha$ -synuclein accumulation, Unc-51-like kinase 1 (ULK1), energy deficit mTOR, TSC1/2, autophagy, protein synthesis, cell cycle, inflammation, neuroprotective, toxicities mTOR, and small molecules (Fig. 1).

### Components and structure of mTOR

mTOR or mammalian/mTOR is a serine/threonine protein kinase. It involves various cell functions such as gene transcription, protein synthesis, cell metabolism, survival, and proliferation [11]. The multidomain protein mTOR is found in two multiprotein complexes: mTOR complex 1 (mTORC1) and mTOR complex 2 (mTORC2) [12]. mTORC1 controls protein synthesis, whereas mTORC2 controls cell survival and cytoskeleton organization [13].

Each complex contains mTOR protein, mammalian lethal with SEC13 protein 8 (mLST8), DEP domain-containing mTOR interacting protein (DEPTOR), Tel two-interacting protein1, and telomere maintenance 2 (Tel2) [14,15]. Two proteins are specific to mTORC1, the regulatory-associated protein of mTOR (Raptor) and proline-rich AKT1 substrate 40 kDa (PRAS40) [16,17]. In these complexes, DEPTOR and PRAS40 can negatively regulate mTORC1 activity [18]. mTORC1 plays a major role in regulating cell growth and proliferation and senses stimuli such as insulin level and amino acid-like Leucine [19,20]. In addition, it is involved in protein synthesis, lipid metabolism, and autophagy [21,22].

mTORC2 contains a Rapamycin-insensitive companion of mTOR, mammalian stress-activated protein kinase-interacting protein1 (mSIN1), and protein observed with Rictor [23], mLST8, and Ttil/Tel2. Akt is an important substrate of mTORC2. Phosphorylation of Akt induces phosphorylation of mSIN1, which increases mTORC2 activity [24]. mTORC2 controls the remodeling of cytoskeleton and electrolyte homeostasis [25].

### mTOR signaling pathways

#### *mTORC1 signaling pathway*

Insulin-like growth factor 1 (IGF-1) activates mTOR. IGF starts the conversion of Ras guanosine 5'-triphosphate (GTP) to Ras guanosine diphosphate (GDP). This conversion leads to the cascade of small molecule activation, which includes Ras, Mek, Erk, and Rsk. Erk and Rsk inhibit the activation of TSC1/2. TSC1/2 complex is an essential regulator in this process [26]. TSC1/2 activates the conversion of GTPase Rheb to GDPase Rheb, an inactive form. This GDP-bound state negatively regulates the mTOR pathway [27]. Other factors, such as inflammation and hypoxia, can also lead to mTOR activation.

In response to the cellular energy deficiency, LKB1 is activated. This activated LKB1 increases the adenosine monophosphate (AMP)/adenosine triphosphate (ATP) ratio. This elevated level of AMP leads to phosphorylation of AMP-activated protein kinase (AMPK), which consecutively activates TSC1/2. The activation of TSC1/2 leads to an increase in the GAP activity of the TSC1/2 complex and inhibits mTORC1 activity [28]. It is further noted that hypoxia can also inhibit mTORC1 activity by the increase in regulated in development and DNA damage responses (REDD1) expression [29]. Inflammation can also lead to the inhibition of TSC1/2 by activating Ikk $\beta$ .

The activated mTORC1 plays its role in protein regulation by two possible mechanisms. One by phosphorylation of ribosomal protein S6 kinase beta-1 (S6K1) and eukaryotic initiation factor 4E-binding protein 1 (4E-BP1) [30]. S6K1 is phosphorylated and activated by mTORC1, which again phosphorylates and activates S6 (a component of the S40 ribosomal subunit). S6K1 also activates several small molecules such as CBP80, Tripartite motif-containing protein-24 (TIF1A), SKAR, and eIF4B, and inhibits eukaryotic elongation factor 2 kinase (eEF2K) and PDCD4. S6K1 inhibits eEF2K by phosphorylation, thus inhibiting the translation and elongation of proteins. Phosphorylation of PDCD4 leads to the inhibition of eukaryotic initiation factor-4A, which is responsible for translation initiation [31]. S6K1 phosphorylates and activates SKAR and CBP80. This activation leads to mRNA biogenesis [32]. S6K1 also activates TIF1A, elevating rRNA expression through interaction through POL1.

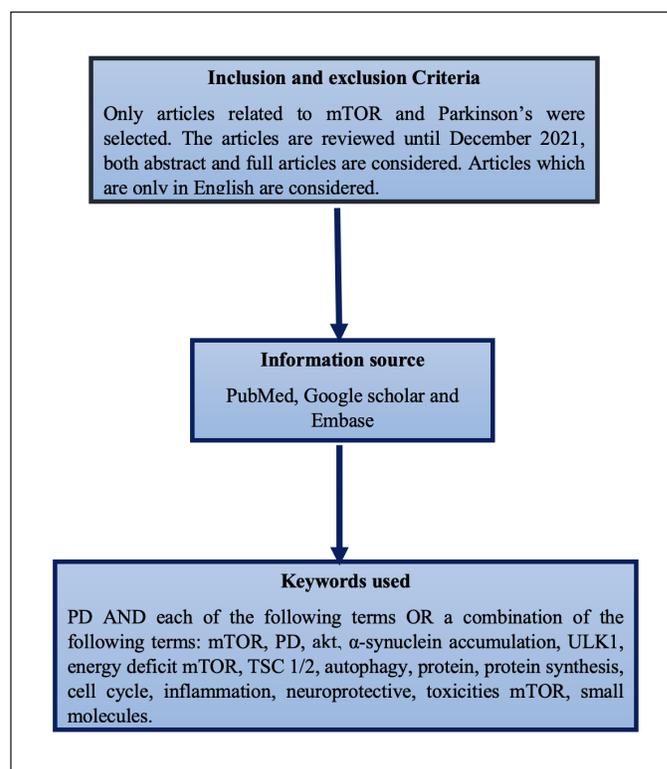


Figure 1. Methodology.





mechanism. In contrast, Chaperone-mediated autophagy exerts degradation in a non-specific manner, i.e., molecule-by-molecule fashion [72]. The major signaling pathway which controls autophagy is mTOR [73]. mTOR exerts its role in the various stages of autophagy, and its activity is tightly regulated by a complex interplay between stimulatory and inhibitory signals [74]. The various signaling pathways through which mTOR regulates autophagy are discussed below:

#### **ULK1 signaling pathway**

The downstream of mTORC1 contains the ULK1 complex, which consists of ATG101, ATG13, and focal adhesion kinase family interacting protein of 200 kD. mTORC1 activates and phosphorylates ULK1 [75]. This phosphorylation leads to the inhibition of interaction with AMPK and inhibits autophagy. On cellular stress, ULK1 is phosphorylated by AMPK, which further leads to the induction of autophagy. Hence, mTORC1 negatively regulates autophagy by the ULK1 complex [76].

#### **FEB signaling pathway**

Transcription factor EB (TFEB) is a part of the bHLH leucine-zipper transcription factor family that regulates autophagy and lysosomal gene expression [77]. Normally inhibition of mTORC1 results in the release of TFEB, which releases transcriptional factors. These transcriptional factors cause autophagy and lysosome biogenesis [78].

#### **Vacuolar protein sorting 34 (VPS34) complex signaling pathway**

Vacuolar protein sorting 34 is another protein through which autophagy induction occurs. VPS34 consists of two complexes: Complex 1 and Complex 2. Complex 1 consists of VPS34, VPS15, beclin 1, and ATG14. Complex 2 consists of VPS34, VPS15, beclin 1, and UVRAG [79]. mTORC1 directly inhibits VPS34 by phosphorylating ATG14 at various sites. Autophagy might be improved by mutating these sites, which are resistant to mTOR inhibition [80].

#### **Autophagic lysosome reformation (ALR)**

Lysosomes are repurposed from autolysosomes at the end of autophagy by a process known as ALR, which comprises proto-lysosome tubule formation, elongation, and termination [81]. In short-term food deprivation, mTOR is inhibited, and in nutrient-rich conditions, mTOR is activated. Also, during prolonged starvation, mTOR is reactivated by a negative feedback loop of autophagy. For this reactivation, autophagic lysosomes have to be degraded. Spinster, a lysosomal efflux protein, is a regulator of ALR. Prolonged starvation leads to defective spinsters. mTOR reactivation failed due to the defective spinster [82]. The general amino acid control pathway is activated by starvation, which leads to the overexpression of amino acid transporters in the plasma membrane, increasing amino acid absorption and contributing to mTOR reactivation [83].

#### **Lysosomal acidification and autophagy**

Lysosomes are crucial organelles involved in the degradation of cellular waste. Lysosomal acidification is

facilitated by proton pumps, predominantly the vacuolar H<sup>+</sup>-ATPase (V-ATPase), which actively pumps protons into the lysosomal lumen. The acidic pH inside lysosomes is crucial for the activation of hydrolytic enzymes, enabling efficient degradation of cellular components during autophagy [84].

#### **The interplay between mTOR and lysosomal acidification**

An intriguing feedback loop exists between mTOR and lysosomal acidification. Amino acids, critical activators of mTORC1, are transported into lysosomes, where they play a pivotal role in recruiting and activating mTORC1 at the lysosomal surface. The V-ATPase, responsible for lysosomal acidification, is central to this process [85]. Understanding the interplay between mTOR and lysosomal acidification holds significant implications for cellular health and disease. Dysregulation of either process can lead to disruptions in autophagy, compromising cellular homeostasis and contributing to the pathogenesis of various diseases, such as cancer, neurodegenerative disorders, and metabolic conditions [86]. In PD, proper lysosomal acidification is necessary for the optimal function of lysosomal enzymes [87]. Genetic investigations have unveiled a significant association between lysosomal function and PD. Several autosomal dominant and recessive genes linked to PD, as well as various genetic risk factors, encode proteins involved in lysosomal, autophagic, and endosomal pathways. Mutations in these PD-associated genes can lead to lysosomal dysfunction, and considering that  $\alpha$ -synuclein degradation is primarily reliant on lysosomal processes, this impairment can hinder  $\alpha$ -synuclein turnover. Consequently, this disruption contributes to elevated intracellular levels of  $\alpha$ -synuclein, facilitating its accumulation and subsequent aggregation, among other consequences.

#### **Feedback loop-AMPK, ULK1, and mTOR**

The main reasons for many neurodegenerative diseases are the overproduction of abnormal proteins and autophagy dysregulation [88]. Much evidence suggests that mTOR signaling is overactivated in PD patients. Overexpression of this mTOR will lead to autophagy inhibition [89]. The activity of mTOR is reduced in animals that cause autophagy induction. Hence, the direct correlation between mTOR and autophagy inducer ULK1 is studied. On the other hand, during energy deficit conditions, AMPK is activated, which inhibits mTORC1, thereby activating ULK1 [90]. This AMPK is further inactivated by mTOR. These shreds of evidence suggest that mTOR, ULK1, and AMPK have a direct relationship. A feedback loop is a mechanism that restores the body to its original state. All stable system has a feedback mechanism to retain control internally. Protein regulator mTOR and autophagy regulator ULK1 and AMPK have a direct feedback link. Two possible cycles exist between them. AMPK activates ULK1 as a process of autophagy induction. This activation leads to direct inhibition of AMPK by ULK1 as a feedback loop. This feedback loop leads to delayed inhibition of ULK1, which results in oscillation of autophagy induction [57]. The double negative-feedback loop in which AMPK directly phosphorylates the mTOR complex and inhibits it. This inhibition results in the inhibition of ULK1, followed by AMPK inhibition [91].

We know that mTOR is activated in nutrient-rich conditions and inhibits autophagy by phosphorylating ULK1. On starvation, phosphorylated AMPK inhibits mTOR, thereby activating autophagy. The factors that activate AMPK are LKB1, CaMKK $\beta$ , TAK1, DNA damage, and a decrease in ATP/AMP ratio [76]. On the other hand, ULK1 is activated by AMPK and inhibited by mTOR. ULK1 also counteracts and inhibits mTOR [92]. mTOR inhibition cannot be permanent. Prolonged inhibition of mTOR leads to reactivation [93]. A recent study suggests that AMPK alone cannot induce autophagy when both mTOR and ULK1 are inhibited [94]. AMPK remains dephosphorylated until mTOR is activated [95]. Therefore, the balance between autophagy and mTOR is challenging to achieve. Currently, mTOR, ULK1, and AMPK gained an attractive target for the treatment of PD. These feedback loops help maintain homeostasis and regulate the balance between protein synthesis and clearance. Hence, the complex interplay of the feedback loops has to be explored, and a better understanding of their autophagy modulation will help us treat PD.

#### Potential PD treatment by targeting mTOR and autophagy

Targeting mTOR and autophagy has emerged as a potential therapeutic strategy for PD due to their critical roles in cellular homeostasis and protein degradation. Dysregulation of mTOR and autophagy pathways has been implicated in the accumulation of toxic protein aggregates, like alpha-synuclein, and the degeneration of dopaminergic neurons, hallmark features of PD [96]. Here are some approaches for potential PD treatment by targeting mTOR and autophagy:

##### mTOR inhibitors

Rapamycin and its analogs (rapalog) are mTOR inhibitors that have shown promise in preclinical studies for their neuroprotective effects in PD models. These inhibitors promote autophagy and reduce the accumulation of toxic protein aggregates [97]. However, further research is needed to optimize their dosing and delivery to the brain while minimizing side effects. Small molecules that control autophagy and mTOR can be targeted, which assists in mTOR and autophagy balance (Table 1).

##### Activators of autophagy

Enhancing autophagy through pharmacological interventions has been explored as a therapeutic strategy for PD. Certain compounds, such as trehalose [98] and lithium [99,100], have been shown to induce autophagy and promote the clearance of toxic proteins. Clinical trials to evaluate their efficacy and safety in PD patients are ongoing.

##### Lysosomal enzyme modulators

Enhancing lysosomal function and promoting efficient degradation of toxic protein aggregates is another potential approach. Small molecules that modulate lysosomal enzymes, such as glucocerebrosidase (GBA), have shown promise in preclinical studies and may hold therapeutic potential for PD with GBA mutations [101].

**Table 1.** The action of small molecules and drugs on various phases of autophagy.

Autophagy phase	Small molecules	Drugs that modulate autophagy
Initiation	↑AMPK	Metformin, [102]
		Resveratrol, [103]
	↓mTOR	Rosuvastatin, [104]
		Temozolomide [105]
		Rapamycin, [106]
Nucleation	↑ULK1	Corynoxine [107]
	↑Beclin1	BL-918 [108]
		Isorhynchophylline, [109]
	↓IMP ase	Glycyrrhizic acid [110]
		Valproate, [111]
Elongation	↑GC case	Carbamazepine [112]
		Curcumin, [113]
	↑ERR- $\alpha$	Trehalose [114]
Degradation	↓c-ABL	Ambroxol [115]
		XCT 790 [116]
		Synphilin-1 [117]

## CONCLUSION

Due to its dual function of neuroprotective and neurotoxic effects, the role of mTOR in neurodegenerative diseases is still debated. Because mTOR is critical for protein synthesis and cell survival processes, it is worth noting that mTOR inhibition causes autophagy to activate, which may play a role in neuroprotection. Long-term inhibition, however, can trigger the feedback loop and mTORC1, preventing autophagy. As a result, mTOR is a double-edged sword. It can play both neuroprotective and neurotoxic. Hence, the signaling between mTOR and autophagy has to be balanced. If the balance between mTOR and autophagy can be achieved, mTOR might be a possible target for the therapy of PD.

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## LIST OF ABBREVIATIONS

AD, Alzheimer's disease; ALR, Autophagic lysosome reformation; AMPK, Adenosine monophosphate-activated protein kinase; DEPTOR, DEP domain-containing mTOR interacting protein; eEF2K, Eukaryotic elongation factor 2 kinase; IGF-1, Insulin-like growth factor 1; mSIN1, Mammalian stress-activated protein kinase-Interacting protein 1; mTOR, Mammalian/mTOR; mTORC1 mTOR complex 1; mTORC2, mTOR complex 2; PD, Parkinson's disease; PKC $\alpha$ , Protein kinase C alpha; Raptor, Regulatory-associated protein of mTOR; REDD1, Regulated in development and DNA damage responses; SGK1, Serum/glucocorticoid regulated

kinase 1; Tel2, Telomere maintenance 2; TIF1A, Tripartite motif-containing protein-24; ULK1, Unc-51-like kinase 1.

#### AUTHOR CONTRIBUTION

TLN, SKM, DT, AB, YMT, SK, and SNM made a significant contribution to the work reported, whether that is in the conception, execution, or the acquisition, analysis, or interpretation of data, or all the areas; took part in drafting, revising, or critically reviewing the article; and gave final approval of the version to be published. All have read and agreed to the published version of the manuscript.

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The authors report no financial or any other conflicts of interest in this work.

#### CONSENT TO PARTICIPATE

It is a review article, thus it is not applicable

#### ETHICAL APPROVALS

This study does not involve experiments on animals or human subjects.

#### DATA AVAILABILITY

The data that support the findings of this study are available in standard research databases such as PubMed, Science Direct, or Google Scholar, and/or on public domains that can be searched with either key words or DOI numbers.

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#### REFERENCES

- Ping LA, Chen J, Zhao Y, Chai Z, Hu Y. mTOR signaling in Parkinson's disease. *Neuromolecular Med.* 2017;19(1):1–10.
- Yang W, Hamilton JL, Kopil C, Beck JC, Tanner CM, Albin RL, *et al.* Current and projected future economic burden of Parkinson's disease in the U.S. *NPJ Parkinsons Dis* [Internet]. 2020 Dec 1 [cited 2023 Aug 21];6(1):15. Available from: [/pmc/articles/PMC7347582/](https://pubmed.ncbi.nlm.nih.gov/35740927/)
- Darden L. Mechanisms and models. *Camb Companion Philos Biol.* 2007;39:139–59.
- Laplanche M, Sabatini DM. MTOR signaling in growth control and disease. *Cell.* 2012;149(2):274–93.
- Burnett PE, Barrow RK, Cohen NA, Snyder SH, Sabatini DM. RAFT1 phosphorylation of the translational regulators p70 S6 kinase and 4E-BP1. *Proc Natl Acad Sci U S A.* 1998;95(4):1432–7.
- Deleyto-Seldas N, Efeyan A. The mTOR–autophagy axis and the control of metabolism. *Front Cell Dev Biol.* 2021 Jul 1;9:655731.
- Crews L, Spencer B, Desplats P, Patrick C, Paulino A, Rockenstein E, *et al.* Selective molecular alterations in the autophagy pathway in patients with lewy body disease and in models of  $\alpha$ -synucleinopathy. *PLoS One.* 2010;5(2):e9313.
- Tan C, Ai J, Zhu Y. mTORC1-dependent protein and Parkinson's disease: a mendelian randomization study. *Brain Sci* [Internet]. 2023 Apr 1 [cited 2023 Aug 21];13(4):536. Available from: [/pmc/articles/PMC10137243/](https://pubmed.ncbi.nlm.nih.gov/41137243/)
- Zhou Q, Liu C, Liu W, Zhang H, Zhang R, Liu J, *et al.* Rotenone induction of hydrogen peroxide inhibits mTOR-mediated S6K1 and 4E-BP1/eIF4E pathways, leading to neuronal apoptosis. *Toxicol Sci.* 2015;143(1):81–96.
- Tuo Y, Xiang M. mTOR: a double-edged sword for diabetes. *J Leukoc Biol.* 2019;106(2):385–95.
- Yang M, Lu Y, Piao W, Jin H. The translational regulation in mTOR pathway. *Biomolecules* [Internet]. 2022 Jun 1 [cited 2023 Aug 2];12(6):802. Available from: <https://pubmed.ncbi.nlm.nih.gov/35740927/>
- Yu L, Wei J, Liu P. Attacking the PI3K/Akt/mTOR signaling pathway for targeted therapeutic treatment in human cancer. *Semin Cancer Biol* [Internet]. 2022 Oct 1 [cited 2023 Aug 21];85:69–94. Available from: <https://pubmed.ncbi.nlm.nih.gov/34175443/>
- Sun Y, Wang H, Qu T, Luo J, An P, Ren F, *et al.* mTORC2: a multifaceted regulator of autophagy. *Cell Commun Signal* [Internet]. 2023 Dec 1 [cited 2023 Aug 21];21(1):4. Available from: <https://pubmed.ncbi.nlm.nih.gov/36604720/>
- Kaizuka T, Hara T, Oshiro N, Kikkawa U, Yonezawa K, Takehana K, *et al.* Tti1 and Tel2 are critical factors in mammalian target of rapamycin complex assembly. *J Biol Chem.* 2010;285(26):20109–16.
- Feng SW, Wu ZS, Chiu YL, Huang SM. Exploring the functional roles of telomere maintenance 2 in the tumorigenesis of glioblastoma multiforme and drug responsiveness to temozolomide. *Int J Mol Sci.* 2023;24:9256. Available from: <https://www.mdpi.com/1422-0067/24/11/9256/htm>
- Sancak Y, Thoreen CC, Peterson TR, Lindquist RA, Kang SA, Spooner E, *et al.* PRAS40 is an insulin-regulated inhibitor of the mTORC1 protein kinase. *Mol Cell.* 2007;25(6):903–15.
- Eum WS, Kim DW, Yeo EJ, Yeo HJ, Choi YJ, Cha HJ, *et al.* Transduced Tat-PRAS40 prevents dopaminergic neuronal cell death through ROS inhibition and interaction with 14-3-3 $\sigma$  protein. *Free Radic Biol Med* [Internet]. 2021 Aug 20 [cited 2023 Aug 21];172:418–29. Available from: <https://pubmed.ncbi.nlm.nih.gov/34175438/>
- de la Cruz López KG, Toledo Guzmán ME, Sánchez EO, García Carrancá A. mTORC1 as a regulator of mitochondrial functions and a therapeutic target in cancer. *Front Oncol.* 2019 Dec 13;9:492202.
- Di Malta C, Siciliano D, Calcagni A, Monfregola J, Punzi S, Pastore N, *et al.* Transcriptional activation of RagD GTPase controls mTORC1 and promotes cancer growth. *Science (1979).* 2017;356(6343):1188–93.
- Beirowski B, Wong KM, Babetto E, Milbrandt J. MTORC1 promotes proliferation of immature Schwann cells and myelin growth of differentiated Schwann cells. *Proc Natl Acad Sci U S A.* 2017;114(21):E4261–70.
- Chiarini F, Evangelisti C, McCubrey JA, Martelli AM. Current treatment strategies for inhibiting mTOR in cancer. *Trends Pharmacol Sci.* 2015;36(2):124–35.
- Caron A, Richard D, Laplanche M. The roles of mTOR complexes in lipid metabolism. *Annu Rev Nutr.* 2015;35(1):321–48.
- Wang B, Jie Z, Joo D, Ordureau A, Liu P, Gan W, *et al.* TRAF2 and OTUD7B govern a ubiquitin-dependent switch that regulates mTORC2 signalling. *Nature.* 2017;545(7654):365–9.
- Zhang Z, Sun X, Wang K, Yu Y, Zhang L, Zhang K, *et al.* Hydrogen-saturated saline mediated neuroprotection through autophagy via PI3K/AKT/mTOR pathway in early and medium stages of rotenone-induced Parkinson's disease rats. *Brain Res Bull.* 2021;172(January):1–13.
- Querfurth H, Lee HK. Mammalian/mechanistic target of rapamycin (mTOR) complexes in neurodegeneration. *Mol Neurodegener.* 2021;16(1):1–25.

26. Fitzian K, Brückner A, Brohée L, Zech R, Antoni C, Kiontke S, *et al.* TSC1 binding to lysosomal PIPs is required for TSC complex translocation and mTORC1 regulation. *Mol Cell*. 2021 Jul 1;81(13):2705–21.e8.
27. Zhong Y, Zhou X, Guan KL, Zhang J. Rheb regulates nuclear mTORC1 activity independent of farnesylation. *Cell Chem Biol*. 2022 Jun 16;29(6):1037–45.e4.
28. Kahn BB, Alquier T, Carling D, Hardie DG. AMP-activated protein kinase: ancient energy gauge provides clues to modern understanding of metabolism. *Cell Metab*. 2005;1(1):15–25.
29. Murakami T. Isometric contraction induces transient increase of REDD1 expression in non-contracted muscles partly through glucocorticoids. *Physiol Rep* [Internet]. 2023 Jun 1 [cited 2023 Aug 22];11(11):e15745. Available from: <https://onlinelibrary.wiley.com/doi/full/10.14814/phy2.15745>
30. Cornu M, Albert V, Hall MN. mTOR in aging, metabolism, and cancer. *Curr Opin Genet Dev*. 2013;23(1):53–62.
31. Andreou AZ, Klostermeier D. The DEAD-box helicase eIF4A. *RNA Biol*. 2013;10(1):19–32.
32. Kim S, Yang JY, Xu J, Jang IC, Prigge MJ, Chua NH. Two cap-binding proteins CBP20 and CBP80 are involved in processing primary microRNAs. *Plant Cell Physiol*. 2008;49(11):1634–44.
33. Ma XM, Blenis J. Molecular mechanisms of mTOR-mediated translational control. *Nat Rev Mol Cell Biol*. 2009;10(5):307–18.
34. Maiti P, Manna J, Dunbar GL, Maiti P, Dunbar GL. Current understanding of the molecular mechanisms in Parkinson's disease: targets for potential treatments. *Transl Neurodegener*. 2017;6(1):1–35.
35. Huang J, Wu S, Wu CL, Manning BD. Signaling events downstream of mammalian target of rapamycin complex 2 are attenuated in cells and tumors deficient for the tuberous sclerosis complex tumor suppressors. *Cancer Res*. 2009;69(15):6107–14.
36. Rosner M, Hengstschläger M. Cytoplasmic and nuclear distribution of the protein complexes mTORC1 and mTORC2: rapamycin triggers dephosphorylation and delocalization of the mTORC2 components rictor and sin1. *Hum Mol Genet*. 2008;17(19):2934–48.
37. Fu W, Wu G. Targeting mTOR for anti-aging and anti-cancer therapy. *Molecules*. 2023;28:3157. Available from: <https://www.mdpi.com/1420-3049/28/7/3157/htm>
38. Tsai PJ, Lai YH, Manne RK, Tsai YS, Sarbassov D, Lin HK. Akt: a key transducer in cancer. *J Biomed Sci*. 2022;29:1–17. Available from: <https://jbiomedsci.biomedcentral.com/articles/10.1186/s12929-022-00860-9>
39. Knight J, Caseldine C, Boykoff MT. Forum review. *Geogr J*. 2010;176(3):267–9.
40. Zhu Z, Yang C, Iyaswamy A, Krishnamoorthi S, Sreenivasurthy SG, Liu J, *et al.* Balancing mTOR signaling and autophagy in the treatment of Parkinson's disease. *Int J Mol Sci*. 2019;20(3):1–15.
41. Malagelada C, Jin ZH, Jackson-Lewis V, Przedborski S, Greene LA. Rapamycin protects against neuron death in *in vitro* and *in vivo* models of Parkinson's disease. *J Neurosci*. 2010;30(3):1166–75.
42. Chen L, Xu B, Liu L, Luo Y, Yin J, Zhou H, *et al.* Hydrogen peroxide inhibits mTOR signaling by activation of AMPK $\alpha$  leading to apoptosis of neuronal cells. *Lab Invest*. 2010;90(5):762–73.
43. Kim SR, Chen X, Oo TF, Kareva T, Yarygina O, Wang C, *et al.* Dopaminergic pathway reconstruction by Akt/Rheb-induced axon regeneration. *Ann Neurol*. 2011;70(1):110–20.
44. Domanskyi A, Geißler C, Vinnikov IA, Alter H, Schober A, Vogt MA, *et al.* Pten ablation in adult dopaminergic neurons is neuroprotective in Parkinson's disease models. *FASEB J*. 2011;25(9):2898–910.
45. Deyoung MP, Horak P, Sofer A, Sgroi D, Ellisen LW. Hypoxia regulates TSC1/2-mTOR signaling and tumor suppression through REDD1-mediated 14-3-3 shuttling. *Genes Dev*. 2008;22(2):239–51.
46. Ainslie GR, Gibson KM, Vogel KR. mTOR, autophagy, aminoacidopathies, and human genetic disorders. *Molecules to medicine with mTOR: translating critical pathways into novel therapeutic strategies*. Amsterdam, The Netherlands: Elsevier Inc.; 2016. 143–66 pp.
47. Licausi F, Hartman NW. Role of mTOR complexes in neurogenesis. *Int J Mol Sci*. 2018;19(5):1544.
48. Mao Z, Zhang W. Role of mTOR in glucose and lipid metabolism. *Int J Mol Sci*. 2018;19(7):1–14.
49. Yoon MS. mTOR as a key regulator in maintaining skeletal muscle mass. *Front Physiol*. 2017;8(OCT):1–9.
50. Morita M, Gravel SP, Hulea L, Larsson O, Pollak M, St-Pierre J, *et al.* mTOR coordinates protein synthesis, mitochondrial activity. *Cell Cycle*. 2015;14(4):473–80.
51. Wills J, Credle J, Oaks AW, Duka V, Lee JH, Jones J, *et al.* Paraquat, but not maneb, induces synucleinopathy and tauopathy in striata of mice through inhibition of proteasomal and autophagic pathways. *PLoS One*. 2012;7(1):1–12.
52. Mao B, Zhang Q, Ma L, Zhao DS, Zhao P, Yan P, *et al.* Overview of research into mTOR inhibitors. *Molecules*. 2022;27:5295. Available from: <https://www.mdpi.com/1420-3049/27/16/5295/htm>
53. Xu Y, Liu C, Chen S, Ye Y, Guo M, Ren Q, *et al.* Activation of AMPK and inactivation of Akt result in suppression of mTOR-mediated S6K1 and 4E-BP1 pathways leading to neuronal cell death in *in vitro* models of Parkinson's disease. *Cell Signal*. 2014;26(8):1680–9.
54. Watson CJE, Clatworthy MR. mTOR inhibitors: sirolimus and everolimus. *Kidney transplantation—principles and practice*. 8th ed. Amsterdam, The Netherlands: Elsevier; 2019. pp 261–82.
55. Van Duijnhoven EM, Boots JMM, Christiaans MHL, Wolffenbuttel BHR, Van Hooff JP. Influence of tacrolimus on glucose metabolism before and after renal transplantation: a prospective study. *J Am Soc Nephrol*. 2001;12(3):583–8.
56. Guo Z, Chen X, Feng P, Yu Q. Short-term rapamycin administration elevated testosterone levels and exacerbated reproductive disorder in dehydroepiandrosterone-induced polycystic ovary syndrome mice. *J Ovarian Res*. 2021;14(1):1–10.
57. Kim J, Kundu M, Viollet B, Guan KL. AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat Cell Biol*. 2011;13(2):132–41.
58. Jung S, Jeong H, Yu SW. Autophagy as a decisive process for cell death. *Exp Mol Med*. 2020;52(6):921–30.
59. Denton D, Kumar S. Autophagy-dependent cell death. *Cell Death Differ*. 2019;26(4):605–16.
60. Yonekawa T, Thorburn A. Autophagy and cell death. *Essays Biochem*. 2013;55(1):105–17.
61. Pandey S, Srivanthapoom P. Levodopa-induced Dyskinesia: clinical features, pathophysiology, and medical management. *Ann Indian Acad Neurol* [Internet]. 2017 Jul 1 [cited 2023 Aug 2];20(3):190. Available from: [/pmc/articles/PMC5586110/](https://pubmed.ncbi.nlm.nih.gov/36690274/)
62. Jankovic J. Motor fluctuations and dyskinesias in Parkinson's disease: clinical manifestations. *Mov Disord*. 2005;20(SUPPL. 11):S11–6.
63. Gorman AM. Neuronal cell death in neurodegenerative diseases: recurring themes around protein handling: apoptosis review series. *J Cell Mol Med*. 2008;12(6A):2263–80.
64. Guo F, Liu X, Cai H, Le W. Autophagy in neurodegenerative diseases: pathogenesis and therapy. *Brain Pathol*. 2018;28(1):3–13. doi: <https://doi.org/10.1111/bpa.12545>
65. Mushtaq A, Ashraf NU, Altaf M. The mTORC1-G9a-H3K9me2 axis negatively regulates autophagy in fatty acid-induced hepatocellular lipotoxicity. *J Biol Chem* [Internet]. 2023 Mar 1 [cited 2023 Aug 22];299(3):102937. Available from: <https://pubmed.ncbi.nlm.nih.gov/36690274/>
66. Tanemura M, Ohmura Y, Deguchi T, MacHida T, Tsukamoto R, Wada H, *et al.* Rapamycin causes upregulation of autophagy and impairs islets function both *in vitro* and *in vivo*. *Am J Transplant*. 2012;12(1):102–14.

67. Meng LH, Zheng XF. Toward rapamycin analog (rapalog)-based precision cancer therapy. *Acta Pharmacol Sin.* 2015;36(10):1163–9. doi: <https://doi.org/10.1038/aps.2015.68>
68. Yang Z, Klionsky DJ. An overview of the molecular mechanism of autophagy. 2009;335:3112–23.
69. Eskelinen EL, Saftig P. Autophagy: a lysosomal degradation pathway with a central role in health and disease. *Biochim Biophys Acta Mol Cell Res.* 2009;1793(4):664–73.
70. Levine B, Kroemer G. SnapShot: macroautophagy. *Cell.* 2008;132(1):162.e1–162.e3.
71. Li WW, Li J, Bao JK. Microautophagy: lesser-known self-eating. *Cell Mol Life Sci.* 2012;69(7):1125–36.
72. Dice JF. Chaperone-mediated autophagy. *Autophagy.* 2007;3(4):295–9.
73. Dunlop EA, Tee AR. MTOR and autophagy: a dynamic relationship governed by nutrients and energy. *Semin Cell Dev Biol.* 2014;36:121–9.
74. Kim YC, Guan KL. MTOR: a pharmacologic target for autophagy regulation. *J Clin Invest.* 2015;125(1):25–32.
75. Glick D, Barth S, Macleod KF. Autophagy: cellular and molecular mechanisms. *J Pathol.* 2010;221(1):3–12.
76. Alers S, Löffler AS, Wesselborg S, Stork B. Role of AMPK-mTOR-Ulk1/2 in the regulation of autophagy: cross talk, shortcuts, and feedbacks. *Mol Cell Biol.* 2012;32(1):2–11.
77. Settembre C, Di Malta C, Polito VA, Arencibia MG, Vetrini F, Erdin S, *et al.* TFEB links autophagy to lysosomal biogenesis. *Science* (1979). 2011;332(6036):1429–33.
78. Chen Y, Gucek M, Puertollano R, Martina JA. MTORC1 functions as a transcriptional regulator of autophagy by preventing nuclear transport of TFEB. *Autophagy.* 2012;8(6):877–6.
79. Jaber N, Zong WX. Class III PI3K Vps34: essential roles in autophagy, endocytosis, and heart and liver function. *Ann N Y Acad Sci.* 2013;1280(1):48–51.
80. Jaber N, Zong WX. Class III PI3K Vps34: essential roles in autophagy, endocytosis, and heart and liver function. *Ann N Y Acad Sci.* 2013 Mar;1280:48–51. doi: <https://doi.org/10.1111/nyas.12026>
81. Chen Y, Yu L. Autophagic lysosome reformation. *Exp Cell Res.* 2013;319(2):142–6.
82. Rong Y, McPhee C, Denga S, Huang L, Chen L, Liu M, *et al.* Spinster is required for autophagic lysosome reformation and mTOR reactivation following starvation. *Proc Natl Acad Sci U S A.* 2011;108(19):7826–31.
83. Chen Y, Yu L. Development of research into autophagic lysosome reformation. *Mol Cells.* 2018;41(1):45–9.
84. Navarro-Romero A, Montpeyó M, Martínez-Vicente M. The emerging role of the lysosome in Parkinson's disease. *Cells* [Internet]. 2020 Nov 2 [cited 2023 Aug 2];9(11):2399. Available from: <https://pubmed.ncbi.nlm.nih.gov/36996994/>
85. Abu-Remaileh M, Wyant GA, Kim C, Laqtm NN, Abbasi M, Chan SH, *et al.* Lysosomal metabolomics reveals V-ATPase-and mTOR-dependent regulation of amino acid efflux from lysosomes. *Science* [Internet]. 2017 Nov 10 [cited 2023 Aug 2];358(6364):807–13. Available from: <https://pubmed.ncbi.nlm.nih.gov/29074583/>
86. Arotcarena ML, Soria FN, Cunha A, Doudnikoff E, Prévot G, Daniel J, *et al.* Acidic nanoparticles protect against  $\alpha$ -synuclein-induced neurodegeneration through the restoration of lysosomal function. *Aging Cell* [Internet]. 2022 Apr 1 [cited 2023 Aug 2];21(4):e13584. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1111/ace1.13584>
87. Fujii T, Nagamori S, Wiriyasermkul P, Zheng S, Yago A, Shimizu T, *et al.* Parkinson's disease-associated ATP13A2/PARK9 functions as a lysosomal H<sup>+</sup>,K<sup>+</sup>-ATPase. *Nat Commun.* 2023;14:1–11. Available from: <https://www.nature.com/articles/s41467-023-37815-z>
88. Deneubourg C, Ramm M, Smith LJ, Baron O, Singh K, Byrne SC, *et al.* The spectrum of neurodevelopmental, neuromuscular and neurodegenerative disorders due to defective autophagy. *Autophagy* [Internet]. 2022 [cited 2023 Aug 2];18(3):496–517. Available from: <https://www.tandfonline.com/doi/abs/10.1080/15548627.2021.1943177>
89. Wen Z, Zhang J, Tang P, Tu N, Wang K, Wu G. Overexpression of miR-185 inhibits autophagy and apoptosis of dopaminergic neurons by regulating the AMPK/mTOR signaling pathway in Parkinson's disease. *Mol Med Rep.* 2018;17(1):131–7.
90. Rey V, Tamargo-Gómez I. From kinases to diseases: investigating the role of AMPK in human pathologies. *Kinases Phosphatases.* 2023;1:181–205. Available from: <https://www.mdpi.com/2813-3757/1/3/12/htm>
91. Holczer M, Hajdú B, Lőrincz T, Szarka A, Bánhegyi G, Kapuy O. Fine-tuning of AMPK-ULK1-mTORC1 regulatory triangle is crucial for autophagy oscillation. *Sci Rep.* 2020;10(1):1–12.
92. Zou L, Liao M, Zhen Y, Zhu S, Chen X, Zhang J, *et al.* Autophagy and beyond: unraveling the complexity of UNC-51-like kinase 1 (ULK1) from biological functions to therapeutic implications. *Acta Pharm Sin B.* 2022 Oct 1;12(10):3743–82.
93. Hua R, Han S, Zhang N, Dai Q, Liu T, Li J. CPKC $\gamma$ -modulated sequential reactivation of mTOR inhibited autophagic flux in neurons exposed to oxygen glucose deprivation/reperfusion. *Int J Mol Sci.* 2018;19(5):1380.
94. Huang WQ, Wen JL, Lin RQ, Wei P, Huang F. Effects of mTOR/NF- $\kappa$ B signaling pathway and high thoracic epidural anesthesia on myocardial ischemia-reperfusion injury via autophagy in rats. *J Cell Physiol.* 2018;233(9):6669–78.
95. Holczer M, Hajdú B, Lőrincz T, Szarka A, Bánhegyi G, Kapuy O. A double negative feedback loop between MTORC1 and AMPK kinases guarantees precise autophagy induction upon cellular stress. *Int J Mol Sci.* 2019;20(22):5543.
96. Tewari D, Patni P, Bishayee A, Sah AN, Bishayee A. Natural products targeting the PI3K-Akt-mTOR signaling pathway in cancer: a novel therapeutic strategy. *Semin Cancer Biol* [Internet]. 2022 May 1 [cited 2023 Aug 2];80:1–17. Available from: <https://pubmed.ncbi.nlm.nih.gov/31866476/>
97. Wang D, Eisen HJ. Mechanistic target of rapamycin (mTOR) inhibitors. *Handb Exp Pharmacol* [Internet]. 2022 [cited 2023 Aug 2];272:53–72. Available from: <https://pubmed.ncbi.nlm.nih.gov/35091825/>
98. Darabi S, Noori-Zadeh A, Abbaszadeh HA, Rajaei F, Bakhtiyari S. Trehalose neuroprotective effects on the substantia nigra dopaminergic cells by activating autophagy and non-canonical Nrf2 pathways. *Iran J Pharm Res* [Internet]. 2019 Jun 1 [cited 2023 Aug 2];18(3):1419. Available from: <https://pubmed.ncbi.nlm.nih.gov/36996994/>
99. Lazzara CA, Kim YH. Potential application of lithium in Parkinson's and other neurodegenerative diseases. *Front Neurosci* [Internet]. 2015 [cited 2023 Aug 2];9(OCT):403. Available from: <https://pubmed.ncbi.nlm.nih.gov/25802027/>
100. Puglisi-Allegra S, Lazzeri G, Busceti CL, Giorgi FS, Biagioni F, Fornai F. Lithium engages autophagy for neuroprotection and neuroplasticity: translational evidence for therapy. *Neurosci Biobehav Rev* [Internet]. 2023 May 1 [cited 2023 Aug 2];148:105148. Available from: <https://pubmed.ncbi.nlm.nih.gov/36996994/>
101. Schapira AHV. Glucocerebrosidase and Parkinson disease: recent advances. *Mol Cell Neurosci* [Internet]. 2015 May 1 [cited 2023 Aug 2];66(Pt A):37–42. Available from: <https://pubmed.ncbi.nlm.nih.gov/25802027/>
102. Lu G, Wu Z, Shang J, Xie Z, Chen C, Zhang C. The effects of metformin on autophagy. *Biomed Pharmacother.* 2021;137(January):111286.
103. Tian Y, Song W, Li D, Cai L, Zhao Y. Resveratrol as a natural regulator of autophagy for prevention and treatment of cancer. *Oncotargets Ther.* 2019;12:8601–9.
104. Kang SY, Lee SB, Kim HJ, Kim HT, Yang HO, Jang W. Autophagic modulation by rosuvastatin prevents rotenone-induced neurotoxicity in an *in vitro* model of Parkinson's disease. *Neurosci Lett.* 2017;642:20–6.

105. Zou Y, Wang Q, Li B, Xie B, Wang W. Temozolomide induces autophagy via ATM-AMPK-ULK1 pathways in glioma. *Mol Med Rep.* 2014;10(1):411–6.
106. Lamming DW. Inhibition of the mechanistic target of rapamycin (mTOR)—rapamycin and beyond. *Cold Spring Harb Perspect Med.* 2016;6(5):a025924.
107. Chen LL, Song JX, Lu JH, Yuan ZW, Liu LF, Durairajan SSK, *et al.* Corynoxine, a natural autophagy enhancer, promotes the clearance of alpha-synuclein via Akt/mTOR pathway. *J Neuroimmune Pharmacol.* 2014;9(3):380–7.
108. Ouyang L, Zhang L, Zhang S, Yao D, Zhao Y, Wang G, *et al.* Small-molecule activator of UNC-51-like kinase 1 (ULK1) that induces cytoprotective autophagy for Parkinson's disease treatment. *J Med Chem.* 2018;61(7):2776–92.
109. Lu JH, Tan JQ, Durairajan SS, Liu LF, Zhang ZH, Ma L, *et al.* Isorhynchophylline, a natural alkaloid, promotes the degradation of alpha-synuclein in neuronal cells via inducing autophagy. *Autophagy.* 2012 Jan;8(1):98–108. doi: <https://doi.org/10.4161/auto.8.1.18313>. Epub 2012 Jan 1. Erratum in: *Autophagy.* 2012 May 1;8(5):864–6.
110. Laconi S, Madeddu MA, Pompei R. Autophagy activation and antiviral activity by a licorice triterpene. *Phytother Res.* 2014;28(12):1890–2.
111. Salsaa M, Aziz K, Lazcano P, Schmidtke MW, Tarsio M, Hüttemann M, *et al.* Valproate activates the Snf1 kinase in *Saccharomyces cerevisiae* by decreasing the cytosolic pH. *J Biol Chem.* 2021;297(4):101110.
112. Motoi Y, Shimada K, Ishiguro K, Hattori N. Lithium and autophagy. *ACS Chem Neurosci.* 2014;5(6):434–42.
113. Zhang J, Wang J, Xu J, Lu Y, Jiang J, Wang L, *et al.* Curcumin targets the TFEB-lysosome pathway for induction of autophagy. *Oncotarget.* 2016;7(46):75659–71.
114. Rusmini P, Cortese K, Crippa V, Cristofani R, Cicardi ME, Ferrari V, *et al.* Trehalose induces autophagy via lysosomal-mediated TFEB activation in models of motoneuron degeneration. *Autophagy.* 2019;15(4):631–51.
115. Kopytova AE, Rychkov GN, Nikolaev MA, Baydakova GV, Cheblov AA, Senkevich KA, *et al.* Ambroxol increases glucocerebrosidase (GCCase) activity and restores GCCase translocation in primary patient-derived macrophages in Gaucher disease and Parkinsonism. *Parkinsonism Relat Disord.* 2021;84(January):112–21.
116. Suresh SN, Jayaprakash Rao M, Manjithaya R. XCT 790 is a pharmacological aggregate inducer in a yeast model of proteotoxicity. *Cell Biol Int.* 2021;45(3):654–61.
117. Zhou ZH, Wu YF, Wang XM, Han YZ. The c-Abl inhibitor in Parkinson disease. *Neurol Sci.* 2017;38(4):547–52.

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