

Antiproliferative, proapoptotic, and antimigration activities of marine sponges against human colon cancer cell line (HCT116)

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ABSTRACT

Marine sponges' anticancer potential relies on various cellular and molecular pathways. *Agelas* sp., *Ircinia* sp., and *Petrosia weinbergi* were collected off the coast of Iligan Bay, Misamis Oriental, Philippines. The antiproliferation, proapoptotic, and antimigration properties of their hexane (H), dichloromethane (DCM), and methanol: water (MW) extracts against Human colorectal carcinoma cell line (HCT116) carcinoma cells were investigated using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), Caspase-Glo[®] 3/7, and *in vitro* scratch wound assay, respectively. The extracts exhibited cytotoxicity against HCT116. The viability of the treated cells was significantly lower ($p < 0.05$) than the negative control. Treatment of the HCT116 with MW and DCM extracts of *P. weinbergi* and DCM extract of *Ircinia* sp. resulted in a statistically higher ($p < 0.05$) luminescence than the negative control. Relative to dimethyl sulfoxide, the extracts increased caspase-3/7 activity more than onefold. This suggests the proapoptotic activity through the activation of the caspase-3/7. Lastly, the MW extract of *Ircinia* sp. and the H extract of *P. weinbergi* significantly ($p < 0.05$) inhibited the migration of HCT116 after wounding. Thus, *Ircinia* sp. and *P. weinbergi* were found to show potential antiproliferative, proapoptotic, and antimigration properties against HCT116. Further investigation of compounds and the mechanism of action involved in their anticancer property are thus recommended.

INTRODUCTION

Cancer is an aberrant proliferation of cells and tissues which produces tumor mass, vascularization, and the subsequent spread to other parts of the body (Kumar and Akdi, 2018). All cancer forms exhibit several characteristics in common, including the capacity to sustain proliferation-associated signals, evade growth suppressors, resist cell death, enable replicative immortality, induce angiogenesis, and activate invasion and metastasis (Hanahan and Weinberg, 2011). Three characteristics of cancer—sustained

proliferation, resistance to apoptosis, and activating invasion by cell migration—were taken into account in this study.

The proliferation of cancer cells substantially impacts the development and spread of cancer. This is due to modifications in the expression and/or activity of proteins related to the cell cycle or the constitutive activation of several signal transduction pathways that stimulate cell growth (Feitelson *et al.*, 2015). Growth and metastasis will eventually result from continued cancer cell multiplication. Therefore, it is crucial to find and develop drugs that will block tumor formation and progression early on. On the other hand, resistance to apoptosis is among the important challenges in cancer treatment. It allows cancer cells to survive longer and provides them more time to develop mutations that will speed up the progression, invasiveness, and metastasis of the tumor (Hassan *et al.*, 2014). As a result, finding metabolites that target apoptotic pathways has become an important strategy

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for creating chemotherapeutic agents. The activation of caspase-3 or -7 is one of the molecular processes displayed by proapoptotic substances (Yadav *et al.*, 2021). Executioner caspase-3 and -7 are downstream effectors of apoptosis. Their activation causes cancer cells to undergo apoptosis, which eventually killed them (Clark and MacKenzie, 2008). Lastly, tumor invasion and metastasis are two pathogenic processes in which cell migration plays a role. Malignant cells invading surrounding tissue and the vascular system constitute the initial stage of tumor metastasis. Cancer cells must migrate to achieve this. The migration of cells, according to Yamaguchi *et al.* (2005), involves the protrusion of the cell membrane and its adherence to the extracellular matrix (ECM). As a result, several studies focus on looking for substances that can eventually be developed into cancer treatment medications and stop cell migration and growth. For this, the scratch wound assay is frequently utilized to comprehend the molecular mechanisms that regulate cell migration (Walter *et al.*, 2010) and to find pharmaceuticals that can influence cell migration and therefore support therapeutic strategies (Decaestecker *et al.*, 2007).

In recent years, the prevalence of colorectal cancer (CRC) has been rising dramatically across the globe. In 2020, about 1.93 million new cases were diagnosed, and 0.94 million people died from CRC, accounting for 9.4% of all cancer-related deaths and 10% of the world's 19.29 million new cases of cancer. This makes CRC the third leading cause of cancer-related deaths in both genders worldwide (Xi and Xu, 2021). Furthermore, given the population growth, aging, and human development projections, the number of cases is predicted to increase to 3.2 million in 2040 (Surti *et al.*, 2022). Thus, various studies devoted to the early detection, diagnosis, prevention, and treatment of CRC are being conducted. HCT116 is a commonly used cell line for CRC studies (Zhou *et al.*, 2017).

The majority of conventional cancer treatments have been created to specifically target one of the seven hallmarks (Ercolano *et al.*, 2019). Although there exist cancer treatment choices, their efficacy restrictions and the severe side effects that cancer patients must endure have prompted increased efforts to look for alternative treatments. The development of a novel type of anticancer medication with fewer side effects and enhanced effectiveness would be a breakthrough in cancer therapeutics.

Marine sponges (phylum Porifera) are among the richest sources of chemically diverse and pharmacologically active metabolites (Sipkema *et al.*, 2005). Natural products from sponges have an amazing variety of chemical components, including uncommon nucleosides, bioactive terpenes, sterols, cyclic peptides, alkaloids, fatty acids, peroxides, and derivatives of amino acids which are frequently halogenated (Blunt *et al.*, 2013). The biological actions of sponges, including their antiviral, antioxidant, antiobesity, antihypertensive, antidiabetic, anticancer, and antiproliferative capabilities, are also attributed to these chemicals (Kumar and Adki, 2018). The anticancer potential of marine sponge-derived metabolites relies on a variety of cellular and molecular pathways, including growth inhibition, the induction of apoptosis and autophagy, antiangiogenic effects, and antimigration (Ruiz-Torres *et al.*, 2017).

Philippine marine biodiversity has been evaluated as one of the richest in the world (Carpenter and Springer, 2005). However, the Philippines' exceptional biodiversity remains

largely unexplored (Scheffers *et al.*, 2012). Thus, marine sponges from Iligan Bay, Misamis, Oriental, Philippines, i.e., *Ircinia* sp., *Agelas* sp., and *Petrosia weinbergi*, were collected and studied. Their antiproliferative, proapoptotic, and antimigration activities against colorectal carcinoma cells (HCT116) were evaluated.

MATERIALS AND METHODS

Materials

The marine sponges *Agelas* sp., *Ircinia* sp., and *Petrosia weinbergi* were collected off the coast of Iligan Bay, Calangahan, Lugait, Misamis Oriental, Philippines, by hand using scuba at a depth of 6–7 m. Authentication and identification of *Agelas* sp. were done by Prof. Angelo Responde, a marine biologist from Mindanao State University-Iligan Institute of Technology (MSU-IIT), Iligan City, Philippines, based on its morphological characteristics. For *Ircinia* sp. and *Petrosia weinbergi*, deoxyribonucleic acid (DNA) barcoding was employed. The DNA extraction and amplification were done using a commercial genomic DNA extraction kit (QIAGEN's DNeasy Blood and Tissue Kit) and PCR kit (Dongsheng Biotech), respectively. Amplification using C2 forward and D2 reverse primers was carried out under standard conditions. PCR product purification and DNA sequencing were carried out by Macrogen Inc. (Seoul, South Korea). The BLASTN search software was used to analyze and compare the sequence results with known sequences in the GenBank database to ascertain the samples' identity. Voucher specimens of *Agelas* sp., *Ircinia* sp., and *P. weinbergi* with voucher specimen numbers KL23, KL26, and KL28, respectively, are kept at Natural Products and Drug Discovery Laboratory, Premier Research Institute of Science and Mathematics (PRISM), MSU-IIT, Iligan City, Philippines.

The HCT116 cancer cells were purchased from the American Type Culture Collection. McCoy's 5A media and the fetal bovine serum were obtained from Gibco, while the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) (Celltiter96® Nonradioactive Cell Proliferation Assay) and the Caspase-Glo® 3/7 Assay kits were purchased from Promega Corporation. In the assays, sodium butyrate (Sigma-Aldrich), digitonin (Sigma-Aldrich), and dimethyl sulfoxide (DMSO) (Sigma-Aldrich) were used.

Sample preparation and extraction

The samples were washed thoroughly with distilled water, chopped, and freeze-dried. Each dried sample was powdered and sequentially soaked for 72 hours in H, DCM, and MW (1:1 v/v). After 72 hours, filtration and concentration of each solvent extract under vacuum were done. The obtained crude nonpolar H, mid-polar DCM, and polar MW extracts were weighed and stored at –20°C until further analysis.

Cell culture

The HCT116 cells were cultured in McCoy's media supplemented with 10% fetal bovine serum (FBS). The cell cultures were incubated and kept at 37°C under a 5% CO₂ atmosphere.

MTT assay

The MTT assay was conducted to determine the cytotoxicity of the sponge extracts against the carcinoma cell lines according to the method described by Roy *et al.* (2020) with

modifications. The provided protocol in the Promega kit (Cell Titer 96[®] Nonradioactive Cell Proliferation Assay) was followed. A 90 µl cell suspension containing 2×10^4 cells was seeded in a 96-well microplate except for the blank wells and incubated overnight at 37°C and a 5% CO₂ atmosphere.

The next day, 10 µl of 300 µg/ml test samples and positive control (digitonin) were added to their corresponding wells in a 96-well plate to produce a 30 µg/ml assay concentration of test solutions. A vehicle DMSO-treated cell in a medium was used as a negative control, while a vehicle DMSO and cell culture medium without the cells was used as a blank. Incubation at 37°C and 5% CO₂ atmosphere was done overnight before the addition of 15 µl MTT reagent and further incubation for 4 hours under 5% CO₂ atmosphere at 37°C. A 100 µl of solubilization solution/stop mix was added to all wells which were then incubated for an hour at 37°C and 5% CO₂ atmosphere. After incubation, the absorbance was measured and recorded at 570 nm using a spectrophotometer (Perkin Elmer Victor X3 multimode spectrophotometer). The cytotoxicity, expressed as percent cell viability, was then calculated as follows:

$$\text{Cell viability, \%} = \frac{\text{Absorbance of sample} - \text{Absorbance of blank}}{\text{Absorbance of negative control} - \text{Absorbance of blank}}$$

Caspase-Glo[®] 3/7 assay

A commercial kit (Promega G8090 Caspase-Glo[®] 3/7 Assay, Promega, Madison, WI) was used to measure the proapoptotic property in terms of caspase-3/7 cleavage. The recommended protocol by the manufacturer was followed with minor modifications on the amount of Caspase-Glo 3/7 reagent utilized. In a 96-well flat bottom plate, a 90 µl of cell suspension (1×10^4 cells) was seeded. The plate was then incubated at 37°C and 5% CO₂ for 6 hours. The cells were then treated with 10 µl of 300 µg/ml sample extracts to yield a 30 µg/ml final assay concentration and were incubated for a period of 20 hours. After 20 hours, a 10 µl Caspase-Glo 3/7 reagent was added to each well. Incubation for 2 hours at 37°C in 5% CO₂ followed. Luminescence was quantified using a luminometer (Perkin Elmer Victor X3 multimode spectrophotometer). A 5 mM sodium butyrate was used as a positive control. On the other hand, DMSO-treated cells in a medium served as a negative control, while a vehicle DMSO and cell culture medium without the cells were used as a blank.

The proapoptotic activity of the samples in terms of caspase-3/7 activity was reported as a mean of the corrected relative light units (RLUs). Higher RLU values are indicative of a stronger caspase-3/7 activity. To further evaluate the proapoptotic activity of the sample, fold change (FC) against the negative control was calculated.

$$FC = \frac{RLU \text{ test sample or positive control}}{RLU \text{ negative control}}$$

In vitro scratch wound assay

The HCT116 at 5×10^4 density was seeded in a 96-well plate and cultured for 3 days at 37°C under a 5% CO₂ atmosphere to attain 90%–100% confluency. A sterile 10 µl tip was used to make a scratch and the culture media were removed carefully. Washing of the cells with 100 µl of PBS to remove detached cells

and other cellular debris was then done. The culture media were replaced with 90 µl of media consisting of lower FBS content (2%). The image of the wound area or scratch was taken using an inverted microscope (time 0). Treatment of the cells with 10 µl of the sample extracts (300 µg/ml) and incubation for 18 hours (time 18) at 37°C under a 5% CO₂ atmosphere followed. Vehicle DMSO-treated cells in a medium were used as a negative control. The cell migration was observed in the images that were taken using an inverted microscope (AmScope) equipped with a digital camera. ImageJ software was used to assess the wound area at time 0 (A_0) and time 18 (A_{18}). Using the formula below, cell migration was calculated as a percentage of wound closure (Almeida *et al.*, 2019):

$$\text{Wound Closure, \%} = \frac{[A_0 - A_{18}]}{A_0} \times 100$$

Statistical analysis

All analysis was done in a three-by-three format (three trials, three replicates per trial). Results are reported as mean values. Significant differences between the negative control (DMSO) and the sample extracts were determined by *T*-test at a 5% significance level.

RESULTS AND DISCUSSION

Antiproliferative activity

All extracts showed cytotoxicity against the HCT116 cells except for the *Ircinia* sp. MW extract (Fig. 1). The viability of HCT116, although greater than 50%, was found significantly lower ($p < 0.05$) than those treated with the vehicle control DMSO. The DCM extract of *Ircinia* sp. gave the lowest percent cell viability (75.81%) followed by the H extract of *Agelas* sp. (77.67%).

Cell proliferation is tightly regulated in normal cells, while cancer cells are characterized by their ability to sustain proliferation-related signals as a consequence of stroma or specific gene expression

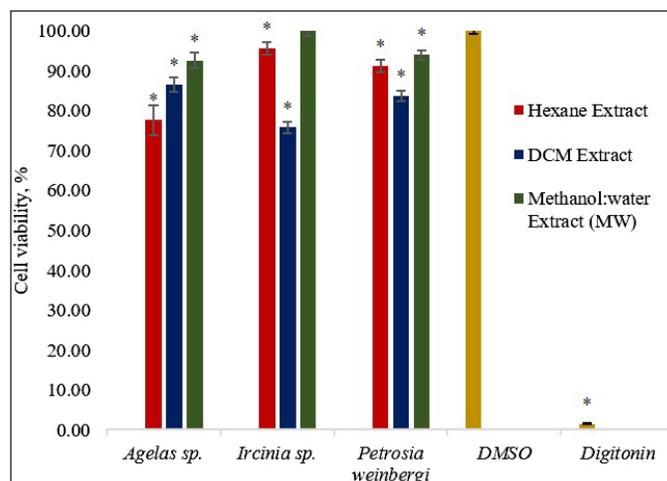


Figure 1. Percent cell viability of HCT116 using the MTT assay after treatment with 30 µg/ml of the sponge extracts. The asterisk (*) indicates significantly lower percent cell viability than the vehicle control (DMSO) based on the *T*-test ($p < 0.05$). Digitonin was used as a positive control. Error bars are SEM ($n = 9$).

patterns (Deshpande *et al.*, 2005). Marine sponges are known prolific producers of secondary metabolites that have exhibited intriguing chemopreventive and chemotherapeutic properties (Calcabrini *et al.*, 2017). According to Calcabrini *et al.* (2017), several marine sponge-produced chemicals show antiproliferative activity that targets the regulators of cell-cycle progression.

Previous studies have reported the cytotoxicity of *Ircinia* and *Agelas* sponges against various carcinoma cells. Their cytotoxicity potential against cancer cells is even more evident with the isolation of bioactive secondary metabolites. The genus *Ircinia* has been a known source of a wide array of steroids, terpenoids, and other metabolites with various pharmacological properties (Heidary *et al.*, 2021). The methanol and diethyl extracts of *I. mutans* from the Persian Gulf have been found strongly active on KB and HUT cells as determined through XTT assay (Nazemi *et al.*, 2020). Moreover, tedanolide C, an 18-membered macrolide ring, which was obtained from the chloroform partition of the methanolic extract of *Ircinia* sp., also exhibited potent cytotoxicity against HCT116 cells. Further, treatment of cells with 0.2 µg/ml of tedanolide C led to the arrest of the cell cycle at the S-phase (Chevallier *et al.*, 2006). On the other hand, *Agelas* sponges have been investigated since the beginning of the 1970s (Zhang *et al.*, 2017). Studies revealed that *Agelas* sponges produce bioactive compounds belonging to alkaloids (bromopyrrole derivatives), terpenoids, fatty acids, glycosphingolipids, and carotenoids (Bickmeyer *et al.*, 2004). The genus *Agelas* has been discovered as the most lucrative producer of bioactive alkaloids (Elissawy *et al.*, 2021). Chu *et al.* (2017) reported the identification of diterpene alkaloids from *A. nakamurai* collected from the South China Sea. Evaluation of the cytotoxicity of the compounds demonstrated weak to moderate effects against HL60, K560, and HCT116. Two new sceptrin derivatives were also obtained from the combined methanol and DCM extracts of *A. kosrae*. The sceptrin alkaloids have shown weak cytotoxicity against carcinoma cells including HCT116 (Kwon *et al.*, 2018). Lastly, oroidin, a pyrrole alkaloid from *A. oroides*, was known to possess moderate cytotoxic activity against many cells, i.e., MCF-7, A2780, and HT29 (Dyson *et al.*, 2014). *Ircinia* sp. DCM and *Agelas* sp. H extracts, thus, can be further purified to determine the compounds responsible for their cytotoxicity.

Proapoptotic activity

The proapoptotic activity of the extracts against HCT116 carcinoma cells was monitored by measuring the luminescence produced after the addition of the caspase-3/7 reagent into the cells and sponge extract reaction mixture. The amount of luminescence recorded is proportional to the intensity of caspase activity exhibited by the sample (Payne *et al.*, 2013). To further describe the degree of changes of the measured signals in the treatments relative to the negative control, FC was calculated.

Treatment of the HCT116 carcinoma cells with MW and DCM extract of *P. weinbergi* and DCM extract of *Ircinia* sp. led to a significantly higher ($p < 0.05$) luminescence signal than the vehicle control (Fig. 2). The said extracts enhanced the caspase-3/7 activity greater than onefold relative to DMSO. Among the three extracts, the DCM extract of *Ircinia* sp. gave the highest RLU and FC value of 563 RLU and 2.69, respectively. The data suggest that the DCM extract of *Ircinia* sp. may have induced the activation of the caspase-3/7 leading to the apoptosis of HCT 116 cells.

Caspase-3 and -7, which are cysteine-aspartic proteases, are downstream effector caspases in the apoptosis pathway. The caspase-3 regulates DNA fragmentation and morphologic changes of apoptotic cells while caspase-7 was found important in the loss of cellular viability (Lakhani *et al.*, 2006). During apoptosis, caspase-3 and caspase-7 are both activated. Regardless of the death-initiating stimulus, both caspases are considered to coordinate during apoptosis by cleaving a diverse array of protein substrates (Timmer and Salvesen, 2007). The induction of caspase-3/7 activity is proportional to the activation of apoptosis (Butterick *et al.*, 2014).

Only a few literature works have reported on the proapoptotic activity of *Ircinia* sponges. A compound furanoterpenoid, 10-acetylirciformonin B, was isolated from the ethyl acetate extract of *Ircinia* sp. This compound exhibited cytotoxicity against HL-60 human leukemia cells by inhibiting cell growth. Interestingly, it also showed proapoptotic properties through the activation of caspase 8, 9, and 3 and induction of PARP cleavage (Su *et al.*, 2012). Chinen *et al.* (2010) have isolated irciniastatin A (ISA) from *I. ramose*. ISA, apart from being selectively cytotoxic against certain human cancer cell lines, activates stress-activated JNK in human leukemia Jurkat

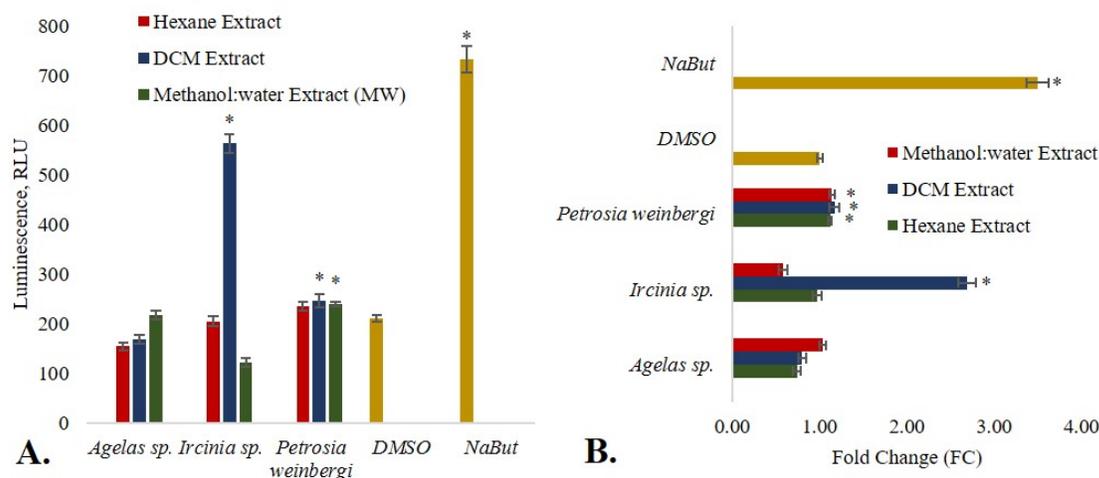


Figure 2. The proapoptotic activity of the sponge extracts is expressed in terms of A. RLU and B. FC. The asterisk (*) indicates a significantly higher RLU or FC than the vehicle control (DMSO) based on the *T*-test ($p < 0.05$). Sodium butyrate (NaBut) was used as a positive control. Error bars are SEM ($n = 9$).

cells leading to apoptosis. The observed proapoptotic property of the DCM extract of *Ircinia* sp. on HCT116 carcinoma cells would warrant further investigation to elucidate both the structure of the bioactive compound and the mechanism of action involved in its exhibited activity.

Antimigration property

The MW extracts of *Agelas* sp. and *Ircinia* sp., the DCM extract of *Ircinia* sp., and all extracts of *P. weinbergi* showed antimigration properties against HCT116 carcinoma cells (Fig. 3). The change in the wound area 18 hours after wounding of the cell layer is smaller in the above-mentioned extracts (Fig. 4). When compared against the vehicle control DMSO, the said extracts gave significantly lower ($p < 0.05$) percent wound closure. The MW extract of *Ircinia* sp. and the H extract of *P. weinbergi* exhibited the highest inhibitory activity against the migration of HCT116 carcinoma cells after wounding. The results imply that *Ircinia* sp. and *P. weinbergi* may have produced compounds that possess antimigration activity.

Cell migration is associated with several physiological and pathological processes. Cell migration facilitates embryonic morphogenesis, aids in tissue repair and regeneration, and promotes the development of diseases including cancer, mental retardation, osteoporosis, atherosclerosis, and arthritis (Ridley *et al.*, 2003). The primary characteristics of tumor biology are cell migration and invasion, which are essential elements of metastasis (van Zijl *et al.*, 2011). As a cyclic process, cell migration is initiated by polarization and extension of protrusions in the direction of migration when a cell responds to a migration-promoting agent. Typically, actin polymerization stimulates the protrusions, which are then stabilized by attachment to the ECM or neighboring cells via transmembrane receptors. These adhesions serve as grip sites for migration as the cells move forward leading to cell movement (Ridley *et al.*, 2003). Wound healing and cancer have been long associated. The mechanisms which regulate wound healing have been found involved in the promotion of the growth and transformation of malignant cells (Arnold *et al.*, 2015). Thus, inhibition of wound healing in cancer cells leads to the suppression of cell migration or invasion (Liew *et al.*, 2017).

The integrins and intracellular signaling proteins found at the regions of focal adhesions stimulate extracellularly and initiate the migration process. The regulation of integrin 1 signaling may prevent cell migration. Basement membranes and extracellular matrices allow tumor cells to move as they spread from the initial site to other subsequent organs (Liew *et al.*, 2017). A study by Yang *et al.* (2021) reported that astragalin dramatically reduced HCT116 cell migration by preventing matrix metalloproteinases (MMP-2 and MMP-9) from being expressed. MMPs are among the most important regulatory molecules in cancer and metastasis (Egeblad and Werb, 2002), and the expression of these molecules is associated with the emergence and growth of tumors (Song *et al.*, 2016). Cantharidin also suppresses HCT116 cell migration by changing the cytoskeleton structure. Many aspects of cell migration are linked to the dynamic movement of the cytoskeleton network (Zhang *et al.*, 2018).

To the best of our knowledge, however, this is the first report on the antimigration property of *Agelas* sp., *Ircinia* sp., and *P. weinbergi* extracts against HCT116 carcinoma cells. Thus, the results of this study may serve as baseline data for future-

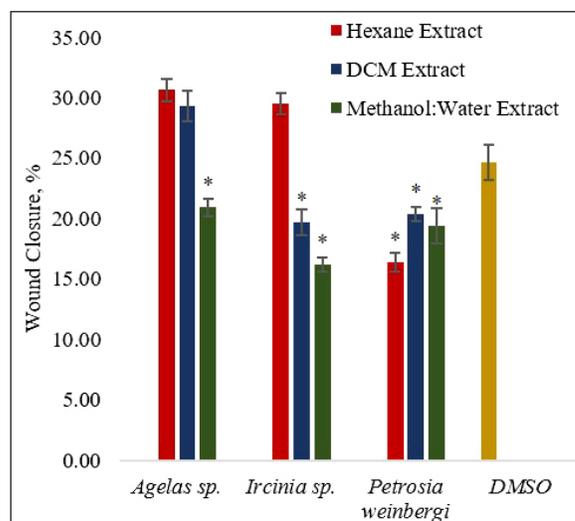


Figure 3. The antimigration activity of the sponge extracts was expressed in terms of percent wound closure 18 hours after treatment of the wound. The asterisk (*) indicates a significantly lower percent wound closure than the vehicle control (DMSO) based on the *T*-test ($p < 0.05$). Sodium butyrate (NaBut) was used as a positive control. The percent wound area of the HCT116 after treatment with NaBut was less than 0 after 18 hours. Error bars are SEM ($n = 9$).

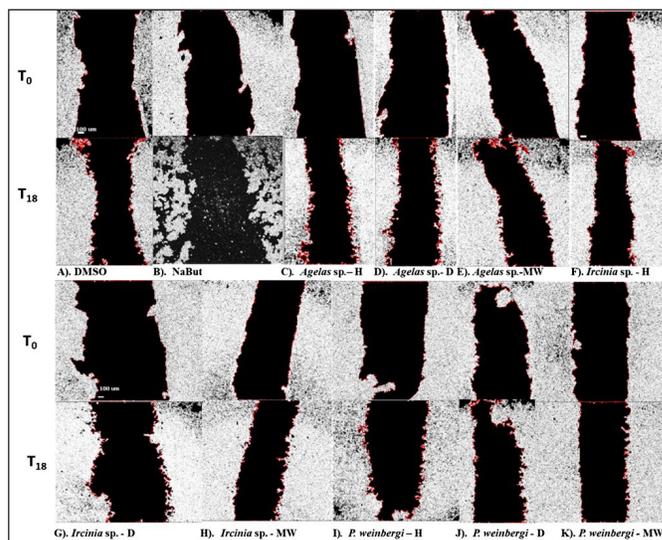


Figure 4. Images of the wound of the HCT116 cells were taken at 0 hours (T_0) after wound and 18 hours (T_{18}) after wound and treatment with (A) DMSO (negative control), (B) NaBut (sodium butyrate as positive control), and (C–K) sponge extracts H, D-DCM, MW). The images and wound area were processed and determined using ImageJ software.

related evaluation of their anticancer properties, especially their antimigration activity.

CONCLUSION

Ircinia sp. and *P. weinbergi* were found to show potential antiproliferative, proapoptotic, and antimigration properties against human colorectal carcinoma cells HCT116. Moreover, this is the first report on the antimigration properties of the *Agelas* sp., *Ircinia* sp., and *P. weinbergi* against HCT116. The results of this

study warrant the isolation of the bioactive compounds from the studied sponges as well as the elucidation of the mechanism of action responsible for their anticancer property.

LIST OF ABBREVIATIONS

The following abbreviations are used in this manuscript:

HCT: human colorectal carcinoma cell line;
FC: fold change; H: hexane; RLU: relative light unit;
DCM: Dichloromethane; MW: methanol: water; MTT:
(3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide).

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AUTHOR CONTRIBUTIONS

The conceptualization and design of the study were done by M.M. Uy. Acquisition, analysis, and interpretation of the data were performed by A.M.G. Ang. The authentication and identification of samples through DNA barcoding experiments were done by S.R.M. Tabugo and A.M.G. Ang. A.M.G. Ang drafted the article. All authors took part in revising the article; agreed to submit it to the current journal; gave final approval of the version to be published; and agree to be accountable for all aspects of the work. All the authors are eligible to be an author as per the international committee of medical journal editors (ICMJE) requirements/guidelines.

CONFLICTS OF INTEREST

The authors declare that there are no conflicts of interest.

ETHICAL APPROVALS

This study does not involve experiments on animals or human subjects.

DATA AVAILABILITY

All data generated and analyzed are included in this research article.

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