



Spray-dried self-nanoemulsifying drug delivery systems as carriers for the oral delivery of piperine: Characterization and *in vitro* evaluation

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ABSTRACT

Piperine is the primary alkaloid compound of white pepper, which has many therapeutic benefits. However, piperine has limitations, including low bioavailability, being photosensitive, and being unstable in liquid preparations. This study aims to increase the dissolution rate and stability of isolated piperine in solid preparations using the solid- self-nanoemulsifying drug delivery systems (S-SNEDDS) formulation approach. The S-SNEDDS formulation was made with Miglyol 812 N, PEG400, and Cremophor RH40 as oil, cosurfactant, and surfactant with a ratio of 2.25:2.60:5.15, respectively. The S-SNEDDS solidification process used mesoporous mannitol as an inert adsorbent. It implemented the spray drying method. S-SNEDDS characterization was performed through droplet size distribution, vibration spectroscopy, morphological characteristics, thermal analysis, X-ray diffraction, and *in vitro* drug release. The characterization results showed no observed crystallization and interaction of isolated piperine with pore carriers in the S-SNEDDS formulation. It was confirmed by vibrational spectroscopy and thermal analysis. S-SNEDDS morphology results showed liquid-SNEDDS contained in mesoporous mannitol pore carriers. In addition, S-SNEDDS increased the dissolution rate of isolated piperine up to 10 times compared to pure isolated piperine. The S-SNEDDS formulation is helpful for dealing with active substances with low solubility and is photosensitive because of their ease of handling and flexibility for further formulation.

INTRODUCTION

Piperine, also known as International Union of Pure and Applied Chemistry 1-(5-[1,3-benzodioxol-5-yl]-1-oxo-2,4-pentadienyl), is the primary alkaloid component of white pepper. The Ayurvedic system of medicine has traditionally used black pepper as an antipain, amenorrhea, antiasthmatic, and cough, and cold medicine. Piperine compounds have several pharmacological effects, including antioxidant, hepatoprotective, antimetastatic (Koul and Kapil, 1993), antidepressant (Li *et al.*, 2007), analgesic, anti-inflammatory, antipyretic (Mujumdar *et al.*, 1990), antithyroid, immunomodulatory, antitumor, inhibit P-glycoprotein expression (Ee *et al.*, 2010; Han *et al.*, 2008; Srinivasan, 2007), and

increase the bioavailability of other drugs through cell membranes by increasing the vasodilation of gastrointestinal tract (GIT) membrane (Khajuria *et al.*, 2002). Although piperine compounds have many therapeutic benefits, they have several disadvantages: photosensitive, unstable in the ultraviolet (UV) light, oxidative stress, and low solubility in water (Kotte *et al.*, 2014). The results of low solubility (22.34 mg/l at 25°C) and low dissolution rate lead to low bioavailability of orally administered drugs (Wu *et al.*, 2012). Thus, piperine compounds have been identified as Biopharmaceutics Classification System (BCS) class II.

Many researchers have attempted to increase the dissolution rate, solubility, and oral bioavailability of piperine. In general, to increase the bioavailability of Active Pharmaceutical Ingredients (APIs) in BCS class II, one needs to increase the dissolution rate of APIs that are poorly soluble in water. Several piperine formulations have been proposed, including solid dispersions (Thenmozhi and Yoo, 2017), nanoparticles (Bhalekar *et al.*, 2017), microemulsions (Etman *et al.*, 2018), hot-melt extrusion technology (Ashour *et al.*,

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2016), liposomes (Dutta and Bhattacharjee, 2017; Pentak, 2016; Priprem *et al.*, 2011), complex with α - and γ -cyclodextrin (Ezawa *et al.*, 2016, 2018), piperine encapsulation with polyethylene glycol poly lactide-co-glycolide (Pachauri *et al.*, 2015), piperine encapsulation with soy-lecithin multilamellar vesicles (Aukunuru and Bonepally, 2017), and a lipid-based self-emulsifying drug delivery system (Shao *et al.*, 2015).

Liquid self-nanoemulsifying drug delivery system (L-SNEDDS) is a lipid-based formulation consisting of a combination of anhydrous isotropic, active substances, liquid/solid surfactants, cosurfactants, and natural/synthetic oils/lipids. It can spontaneously form an oil in water emulsion (o/w) by gentle agitation in GIT fluids. This droplet emulsion is advantageous for drug absorption due to the large interfacial surface area. Therefore, it can improve the bioavailability of APIs with low solubility in water (Kim *et al.*, 2014; Rashid *et al.*, 2015). However, L-SNEDDS has several limitations, such as stability during handling and storage, interactions between L-SNEDDS and soft/hard capsule shells, and production problems that often hinder the L-SNEDDS application in the pharmaceutical industry (Ahmad *et al.*, 2014; Cole *et al.*, 2008).

Thus, along with technological developments, the solid- self-nanoemulsifying drug delivery systems (S-SNEDDS) formulation has been extensively explored to overcome the shortcomings of the L-SNEDDS formulation (Kang *et al.*, 2012; Kim *et al.*, 2017; Mustapha *et al.*, 2017). The S-SNEDDS formulation was developed by converting L-SNEDDS into powder form through spray drying, freeze-drying (Kang *et al.*, 2012; Seo *et al.*, 2013), spray cooling (Cavallari *et al.*, 2005; Passerini *et al.*, 2006), melt granulation, melt extrusion/extrusion spheronization (Tang *et al.*, 2008), or solid carrier adsorption (Ameeduzzafar *et al.*, 2019). The S-SNEDDS dosage form provides many benefits, such as high stability, ease of scale-up, uniformity of content, accurate doses, and increase in patient's compliance (Bari *et al.*, 2015; Hu *et al.*, 2012). The S-SNEDDS formulation provides a promising drug delivery system for active substances that are poorly soluble in water, photosensitive, and unstable in liquid preparations. S-SNEDDS provides advantages by combining the L-SNEDDS advantages (enhanced solubility and bioavailability) with solid dosage forms (high stability during handling and storage in a large selection of dosage forms) (Nazzal and Khan, 2006; Wang *et al.*, 2009).

This study was aimed at solidifying L-SNEDDS containing isolated piperine through a mesoporous mannitol carrier and spray-dried to produce S-SNEDDS powder. It was to overcome the solubility and stability of piperine in liquid preparations. Miglyol 812 N, polyethylene glycol 400 (PEG400), and Cremophor RH40 at a ratio of 2.25:2.60:5.15 were chosen as the optimum formula for L-SNEDDS isolated piperine. The three ingredients are proven safe for oral formulations. Furthermore, the surfactant Cremophor RH40 has been successfully used in formulating SNEDDS products, including Neural[®] (Novartis) product containing cyclosporine A (Badran *et al.*, 2014; Krstić *et al.*, 2018; Mohd *et al.*, 2015). Mesoporous mannitol made of D-mannitol and templating agent ammonium bicarbonate in a ratio of 1:1 w/w% was selected as a high porous adsorbent for the solidification process of L-SNEDDS by spray drying technique.

MATERIAL AND METHODS

Isolated piperine (purity 95%) was obtained by isolating white pepper (*Piper nigrum* L) (Sorowako, South Sulawesi, Indonesia), Cremophor RH40 (Clariant, Indonesia), Miglyol 812 N (IOI Oleochemical, Germany), PEG 400 (Brataco, Indonesia), Tween 80 (Brataco, Indonesia), olive oil (Brataco, Indonesia), virgin coconut oil (VCO), soybean oil, isopropyl myristate (Brataco, Indonesia), propylene glycol (Brataco, Indonesia), water for injection (IKA Pharmindo, Indonesia), methanol for high performance liquid chromatography (HPLC) (Merck, Germany), D-mannitol (Merck, Germany), and ammonium bicarbonate (Merck, Germany).

Instrumentation

The instruments used in this study included stirrer (Stuart Hotplate Stirrer CB162, UK), analytical balance (Ohaus PA 323, China), semi-micro-analytical balance (Shimadzu AP135W, Japan), vortex (D-Lab MX-S, USA), centrifuge (Hettich, EBA 8, Germany), HPLC Hitachi L-2420 UV-Vis detector with Luna[®] 5 μ m C₁₈ 100 Å LC Column 250 × 4.6 mm Phenomenex (Hitachi, Japan), UV-Vis spectrophotometer (Genesys 10S, United States Pharmacopeia), spray dryer (Buchi Mini Spray Dryer B-290, Swiss), scanning electron microscopy (SEM) (JSM-6510LA, Japan), Quadrasorb Evo surface area analyzer (SAA) (Boynnton Beach FL, USA), Differential scanning calorimetry (DSC) (DSC-60 Plus Shimadzu, Japan), Fourier transform infrared (FTIR) (Thermo Scientific Nicolet iS10, USA), Malvern Zetasizer (Malvern Instruments, UK), and Dissolution Tester RC-1 (Hanchen Instrument).

Extraction and purification of piperine compounds

White pepper fruit powder was extracted using a proanalytical n-hexane solvent using the Soxhlet method. The extracted filtrate was then evaporated at room temperature of 25°C up to half portion and stored in the refrigerator for 2 days. The crystals formed were filtered and dried at room temperature of 25°C. The formed yellow crystals were washed using cyclohexane. Then the insoluble crystals were dried at room temperature of 25°C. The results obtained were isolated piperine with a purity of $\geq 95\%$ and had been previously published by Kusumorini *et al.* (2021b).

Analytical method

Determination of isolated piperine was measured by a validated method using the HPLC system (Hitachi L-2420, Tokyo, Japan) with column conditions C₁₈ Phenomenex[®], size 4.6 × 250 mm; particle size 5 μ m; mobile phase methanol:water with a ratio of 75:25% v/v; flow rate 1 ml/minute; sample injection volume 20 μ l; and temperature of 25°C. The wavelength of the UV detector is 340 nm. The validation result of this method has been published in Kusumorini *et al.* (2021a).

Preparation of S-SNEDDS formulations

The S-SNEDDS formulation was prepared in two steps. First, L-SNEDDS was made from the ratio of oil: S_{mix} (ratio of surfactant and cosurfactant) selected from the optimization results using D-Optimal design, Design-Expert software (DX ver.10, State Ease Inc, Minnesota). Next, the S-SNEDDS formulation

was prepared using a mesoporous mannitol hydrophilic carrier with spray drying technology.

Solubility studies of isolated piperine

Solubility tests of isolated piperine in various oils (VCO, olive oil, canola oil, and Miglyol 812 N), surfactants (Cremophor RH40 and Tween 80), and cosurfactants (PEG400 and propylene glycol) were carried out by dissolving 10.0 mg of isolated piperine in 10.0 ml of the carrier. The mixture was vortexed for 10 minutes and stirred for 2 hours at $37^{\circ}\text{C} \pm 1^{\circ}\text{C}$. Furthermore, the insoluble active substance was separated through centrifugation at a speed of 3,000 rpm for 30 minutes. A precipitate, the rest of the active substance, was then separated from the supernatant. Furthermore, the supernatant dissolved in methanol and the levels were determined using HPLC in a validated method. The carriers with the highest solubility (oils, surfactants, and cosurfactants) were selected and used for SNEDDS liquid.

Construction of the pseudoternary phase diagrams

The pseudoternary phase diagram of nanoemulsion was made by mixing surfactants, cosurfactants, and oil titrated with water until they were homogeneous. The surfactant and cosurfactant (S_{mix}) mixture was prepared at different volume ratios (1:1, 1:2, 2:1, and 1:3). Next, the oil phase was added to the S_{mix} at the volume ratio of 1:9, 2:8, 3:7, 4:6, 5:5, 6:4, 7:3, 8:2, and 9:1, respectively. The monophasic mixture was slowly titrated with water and stirred using a magnetic stirrer. After achieving a homogeneous equilibrium mixture until the mixture was translucent in color, it was titrated with water until it became cloudy or turbid. The tendency to emulsify spontaneously and the development of emulsion droplets were visually observed (Khan *et al.*, 2015). The ternary phase diagram was plotted using the Triplot version 4.1.2 software (Todd A. Thompson, LA).

Preparation of isolated piperine loaded in L-SNEDDS

SNEDDS containing isolated piperine was made by mixing isolated piperine with Cremophor RH40, PEG400, and Miglyol 812N. The SNEDDS was stirred for 30 minutes and then stored at room temperature at 30°C for 48 hours (Khan *et al.*, 2015). The number of the three components was determined based on the optimization results of the three components using the D-Optimal design and Design-Expert software (DX version 10, State Ease Inc., MN). The results have been previously published in Kusumorini *et al.* (2021a). The optimization range of the three components was obtained from the results of the emulsion region of the ternary phase diagram. The optimum formula L-SNEDDS obtained consisted of Miglyol 812 N (oil), PEG400 (cosurfactant), and Cremophor RH40 (surfactant) at a ratio of 2.25:2.60:5.15, respectively (Kusumorini *et al.*, 2021a).

Determination of the amount of isolated piperine loaded in L-SNEDDS formulation

The maximum amount of isolated piperine contained in the L-SNEDDS formulation was determined by a validated method using the HPLC system (Hitachi L-2420, Tokyo, Japan), as described in the "Analytical method." A total of 50, 100, 150, 200, and 250 mg of isolated piperine were mixed with 10 g of L-SNEDDS optimum formula, homogenized using vortex for 5 minutes, and stirred for 30 minutes. Furthermore, the SNEDDS

mixture was centrifuged at 3,000 rpm for 15 minutes. The supernatant was taken and filtered through a 0.45 μl cellulose membrane. The filtered solution was diluted with methanol and measured by the validated HPLC method (Vu *et al.*, 2020).

Characterization of L-SNEDDS isolated piperine

Determination of percent transmittance

The transmittance percentage of the L-SNEDDS isolated piperine formulation was measured by dissolving 100 μl of L-SNEDDS isolated piperine in 10 ml of distilled water, homogenized with a stirrer. Afterward, the clarity of the isotropic system was read using a Shimadzu UV spectrophotometer (Genesys 10S, USA) at a wavelength of 650 nm (Khan *et al.*, 2015).

Determination of emulsification time

The emulsification time of L-SNEDDS isolated piperine was measured by dropping 1 ml of L-SNEDDS isolated piperine into 500 ml of distilled water at $37^{\circ}\text{C} \pm 1^{\circ}\text{C}$, stirred using a magnetic stirrer at a speed of 120 rpm. The emulsification process was plotted against the time required to form a homogeneous SNEDDS droplet mixture in the medium (Khan *et al.*, 2015).

Determination of droplet size, polydispersity index, and zeta potential

Particle size, polydisperse index, and zeta potential were analyzed by diluting 1 ml of L-SNEDDS isolated piperine in 100 ml of distilled water. The dilution was homogenized using a magnetic stirrer. A particle size analyzer (Malvern Instruments, UK) was used to monitor light scattering at 25°C and an angle of 172° (Salem *et al.*, 2019).

Preparation of mesoporous mannitol

Mesoporous mannitol powder was the mixture of 50 g of D-mannitol and 50 g ammonium bicarbonate dissolved with 1 l of distilled water at 30°C and stirred for 10 minutes (Song *et al.*, 2018). Then, the solution was dried using a spray dryer Buchi B-290 with an inlet and outlet temperature of 120°C and 80°C , an aspirator of 90%, a pump on a scale of 25%, and a pressure of 50 mBar. As a result, mesoporous mannitol powder obtained characterized attributes including specific surface area, pore size, pore volume, and surface morphological shape.

Characterization of mesoporous mannitol

Scanning electron microscopy

The spray drying of the mixture of D-mannitol with ammonium bicarbonate and pure D-mannitol was tested for their morphological form using SEM. Samples were prepared by coating with platinum for 130 seconds. Furthermore, the sample was observed using SEM with several magnifications to obtain a suitable morphological image (Kuncahyo *et al.*, 2019).

Surface area analyzer

The spray drying of a mixture of D-mannitol and ammonium bicarbonate and pure D-mannitol was tested for surface area, pore size, and pore volume using a Quadrasorb Evo SAA (Boynton Beach, FL). The sample was vacuumed for 16 hours before the measurement process. The measurements used nitrogen gas sorption and desorption techniques. The specific area,

pore size, and pore volume were based on the Brunauer-Emmet-Teller (BET) analysis (Kuncahyo *et al.*, 2019).

Preparation of S-SNEDDS using spray drying technique

The spray drying technique was for the preparation of S-SNEDDS. Solidification of S-SNEDDS was done by dissolving the mesoporous mannitol as a hydrophilic carrier (5 g) in 200 ml of distilled water using a stirrer to produce a clear solution. Next, L-SNEDDS isolated piperine (5 ml) was added to the hydrophilic solution and stirred using a stirrer at 100 rpm to achieve a homogeneous solution. The solution was dried with a Buchi B-290 spray dryer at 120°C inlet and 80°C outlet temperatures, an aspirator of 90%, a pump on a scale of 25%, and a pressure of 50 mBar (Kuncahyo *et al.*, 2019).

Characterization of S-SNEDDS

Measurement of globule size

The formation of S-SNEDDS was determined by measuring the globule size. A total of 50.0 mg of S-SNEDDS was weighted and added to 100.0 ml of distilled water at 37°C ± 2°C and rotated at 100 rpm until nanoemulsion disperse was obtained. Afterward, the globule size was measured using a Malvern Zetasizer globule size measuring device SZ-100 (Malvern Instruments, UK).

Fourier transform infrared

Functional group analysis of isolated piperine samples, mesoporous mannitol, and S-SNEDDS used attenuated total reflectance-FTIR with ZnSe crystals equipped with a deuterated triGlycine sulfate detector. Samples were read as absorbance at the wavelength of 400–4,000 cm⁻¹ and the resolution of 8 with 32 iterations (Kuncahyo *et al.*, 2019).

Scanning electron microscopy

Morphology was conducted on isolated piperine, mesoporous mannitol, and S-SNEDDS samples. Samples were prepared by coating with platinum for 130 seconds. Samples were observed using SEM at an acceleration voltage of 5 kV with several magnifications to obtain a suitable morphological image of mesoporous mannitol (Kuncahyo *et al.*, 2019).

Differential scanning calorimetry

Thermal analysis of isolated piperine samples, mesoporous mannitol, and S-SNEDDS were done using DSC. A total of 5.0 mg of the specimen was placed in an aluminum pan and heated at a rate of 10°C/minute with a nitrogen rate of 30 ml/minute and measured at a temperature of 25°C–30°C. The temperature and enthalpy of the DSC tool were calibrated with iodine standard (Kuncahyo *et al.*, 2019).

Powder X-ray diffraction (PXRD)

The crystallinity pattern characterization used PXRD with Cu- α radiation (wavelength of 1.5406 Å and a voltage of 40 kV). The samples analyzed included isolated piperine, mesoporous mannitol, and S-SNEDDS. Readings were at a rate of 3°C/minute

in the range of 2 θ (diffraction angle) of 3°C–40°C with a step size of 0.02° (Kuncahyo *et al.*, 2019).

Test of the content of S-SNEDDS isolated piperine

100.0 mg of S-SNEDDS was dissolved in 10.0 ml of distilled water using a stirrer until it became clear. Observation of isolated piperine content in S-SNEDDS was determined using HPLC with a previously validated method.

In vitro dissolution study

The *in vitro* dissolution test of isolated piperine, L-SNEDDS isolated piperine, and S-SNEDDS isolated piperine (containing 50.0 mg of isolated piperine) used simulated gastric fluids (SGF) at pH 1.2 and simulated intestinal fluids (SIF) at pH 6.8 at 37°C ± 0.5°C of 900 ml. The dissolution tool used USP type II (paddle type) at 50 rpm. Isolated piperine, L-SNEDDS isolated piperine, and S-SNEDDS isolated piperine were put into a hard-shell capsule size of 0. After that, a 5 ml sample was taken using a volume pipette at intervals of 5, 10, 20, 30, 45, 60, 90, and 120 minutes. At each interval time, the medium was replaced with the same volume of fresh medium to preserve the sample volume. The specimen obtained was filtered using a membrane filter; then, the levels were determined using HPLC with a validated method after filtering through a 0.45 μ m Millipore filter. Data are presented as mean ± standard deviation (SD) of triplicate samples. To assess the comparative rate of increased dissolution of the L-SNEDDS and S-SNEDDS formulations, the mean dissolution time or mean dissolution time (MDT) (minute) and dissolution efficiency (DE) were calculated with Microsoft Excel version 10.0 (Microsoft, USA).

Data analysis and statistics

Each experiment was carried out with three replications. The results were expressed as mean ± SD. The computerized data were described statistically using Microsoft Excel v.10.0 (Microsoft, USA). The results of the L-SNEDDS and S-SNEDDS particle sizes were analyzed using a paired *t*-test, Statistical Package for the Social Sciences (SPSS) v.24 software (IBM, AS).

RESULTS AND DISCUSSION

Screening of components

The solubility of isolated piperine in various excipients includes oils (VCO, olive oil, canola oil, and Miglyol 812 N), surfactants (Cremophor RH40 and Tween 80), and cosurfactants (PEG400 and propylene glycol), as shown in Figure 1. The components used in the SNEDDS formulation must be capable of maximally dissolving the isolated piperine and have a larger self-emulsification area in the ternary phase diagram (Marasini *et al.*, 2012). The vehicle was selected on considerations of safety, compatibility, and dissolving capacity. Among the tested oils, Miglyol 812 N showed the highest solubility of isolated piperine (3.93 ± 0.37 mg/ml) and was selected as the oil phase for further investigation. Isolated piperine dissolved highly in semisynthetic oil derivatives compared to natural oils. It may be associated with the polarity of poorly soluble drugs that prefer to dissolve in oil volume with small/medium molar, such as medium-chain monoglycerides, diglycerides, or triglycerides (Lawrence and

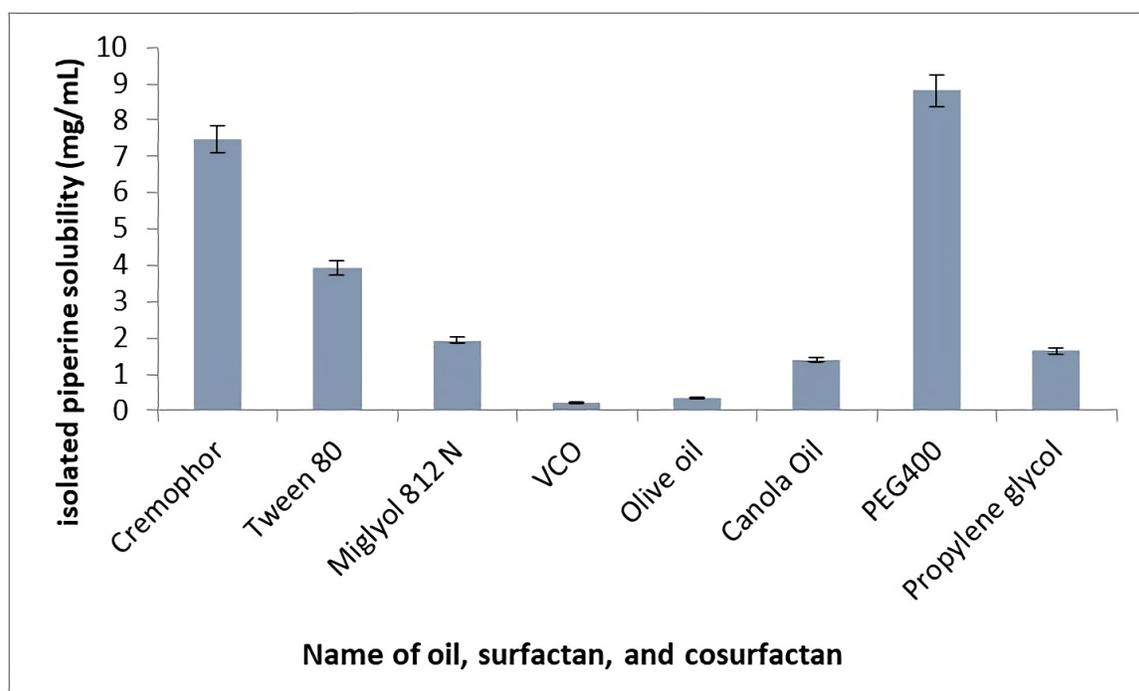


Figure 1. Solubility study of isolated piperine. Each value represents the means \pm SD ($n = 3$).

Rees, 2000). Miglyol 812 N was a medium-chain triglyceride (MCT) oil, a triglyceride ester from saturated coconut/palm kernel oil derived from caprylic, capric, and glycerol-derived fatty acids. MCT oil has a greater dissolving capacity than long-chain triglyceride (LCT), does not undergo oxidation, and can increase the ability to emulsify (Pouton and Porter, 2008).

Screening of components in the manufacture of L-SNEDDS was not only based on the highest solubility of isolated piperine in oil but also with surfactants and cosurfactants. Among the tested surfactants and cosurfactants, Cremophor RH40 (7.48 ± 0.09 mg/ml) and PEG400 (8.82 ± 0.43 mg/ml) showed the highest solubility of isolated piperine. Therefore, both were selected for further research. Cremophor was a nonionic surfactant that was generally considered the safest to take orally and most widely used for lipid-based delivery systems (Pouton and Porter, 2008). Based on the results of previous research, the usage of Cremophor with MCT oil dissolved more active substances than LCT oil (Thomas *et al.*, 2012). PEG400 was the most commonly used cosolvent for the manufacture of L-SNEDDS. Cosolvents serve to help increase the solubility capacity of drugs that dissolve in cosolvents.

The use of these three components in the L-SNEDDS isolated piperine formulation for oral administrations was safe. Moreover, the surfactant Cremophor RH40 (polyoxyl 40 hydrogenated castor oil) has been successfully used to formulate SNEDDS products on the market, namely, the Neural[®] product (Novartis). It contains cyclosporine A and is given 2–4 times a day, so a patient consumes approximately 2–3 g daily (Badran *et al.*, 2014; Krstić *et al.*, 2018; Mohd *et al.*, 2015).

Construction of pseudoternary phase diagrams

The relationship between the phase behavior and the L-SNEDDS mixture's composition was studied using a pseudoternary phase diagram. A pseudoternary phase diagram

was constructed without the active substance to identify the self-emulsification region and optimize the percentages of oil, surfactant, and cosurfactant for the L-SNEDDS formulation. The pseudoternary phase diagram of the system containing Miglyol 812 N as oil, Cremophor RH40 as a surfactant, and PEG400 as a cosurfactant is shown in Figure 2. The results show that Miglyol 812N, PEG400, and Cremophor RH40 have high nanoemulsion region capacity. Large emulsion regions were formed with the proportion of Miglyol 812 N in the range of 10%–65%, PEG400 in the span of 5%–65%, and Cremophor RH40 in the span of 5%–78%. The ternary phase diagram was constructed from the three constituent components of L-SNEDDS. The nanoemulsion region was determined based on the emulsion formed after adding a certain amount of water to the mixture of the three components in the ternary phase diagram. Based on the range of emulsion formation in the ternary phase diagram, the mixture of oil, surfactant, and cosurfactant was further optimized using the D-Optimal design, Design-Expert software (DX v.10, State Ease Inc., MN). Based on the optimization results published in Kusumorini *et al.*, (2021a), the ratio of oil (Miglyol 812N), cosurfactant (PEG 400), and surfactant (Cremophor RH40) chosen was 2.25:2.60:5.15. Therefore, it was used as the constituent of L-SNEDDS isolated piperine.

Determination of the amount of isolated piperine loaded in L-SNEDDS formulation

Determining the maximum amount of isolated piperine contained in L-SNEDDS is needed to determine the weight of isolated piperine used for the L-SNEDDS formulation. The results of the study using isolated piperine weight series 50.0, 100.0, 200.0, and 250.0 mg were 5.03 ± 0.01 , 9.48 ± 0.27 , 19.66 ± 0.35 , and 19.62 mg/ml, each. Isolated piperine with a 20 mg/ml weight

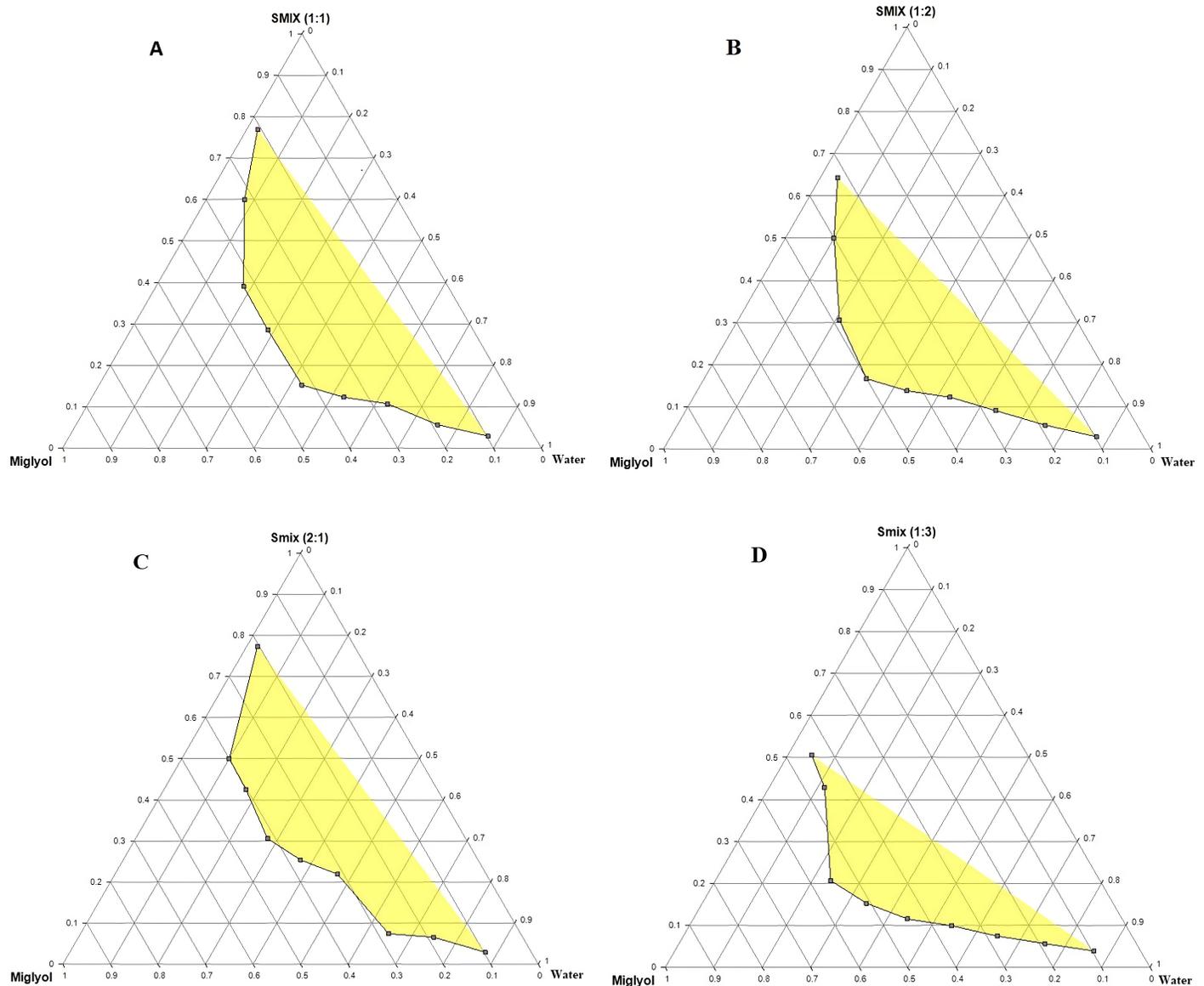


Figure 2. Pseudoternary phase diagram of S_{mix} , oil, and water in four different ratios of S_{mix} , including (A) S_{mix} 1:1, (B) S_{mix} 1:2, (C) S_{mix} 2:1, and (D) S_{mix} 1:3.

was chosen as the weight of the active substance used for the L-SNEDDS formulation.

Formulation and characterization of L-SNEDDS isolated piperine

L-SNEDDS isolated piperine formula in the ratio of oil (Miglyol 812N), cosurfactant (PEG 400), and surfactant (Cremophor RH40), 2.25:2.60:5.15, produced a particle size of 33.35 ± 1.97 nm, zeta potential of -22.87 ± 3.31 mV, percent transmittance of $96.34\% \pm 0.33\%$, and emulsification time of 14.91 ± 0.10 seconds. In the SNEDDS formulation, droplet size was the most crucial factor affecting the determination of the rate of drug release and absorption (Parmar *et al.*, 2011), followed by the zeta potential value, which played a role in the stability of the colloid disperse. The increase in electrostatic repulsion forces between globules could prevent coalescence/fusion between droplets, while the decrease in electrostatic repulsion could cause phase separation (Khan *et al.*,

2015). The zeta potential value limit for SNEDDS ranged from ± 30 mV (Cherniakov *et al.*, 2015).

The results of particle size and zeta potential showed that the characteristics of L-SNEDDS in this study met the requirements for the characteristics of nanoemulsions (<200 nm). The reduction of particle size in the L-SNEDDS preparation allowed isolated piperine to be easily permeated. In addition, negatively charged L-SNEDDS was also permeated faster in mucus than positively charged L-SNEDDS. Thus, both characteristics enhance biological transport through mucus as a barrier to oral absorption (Griesser *et al.*, 2018).

Mesoporous mannitol

In the solidification of L-SNEDDS isolated piperine, solid excipients played a significant role in the L-SNEDDS drug's successful loading. The carrier with high porosity and surface area allows numerous L-SNEDDS to be contained in the pore. Mesoporous silica is an attractive excipient for L-SNEDDS solidification because of its high surface area and large pore

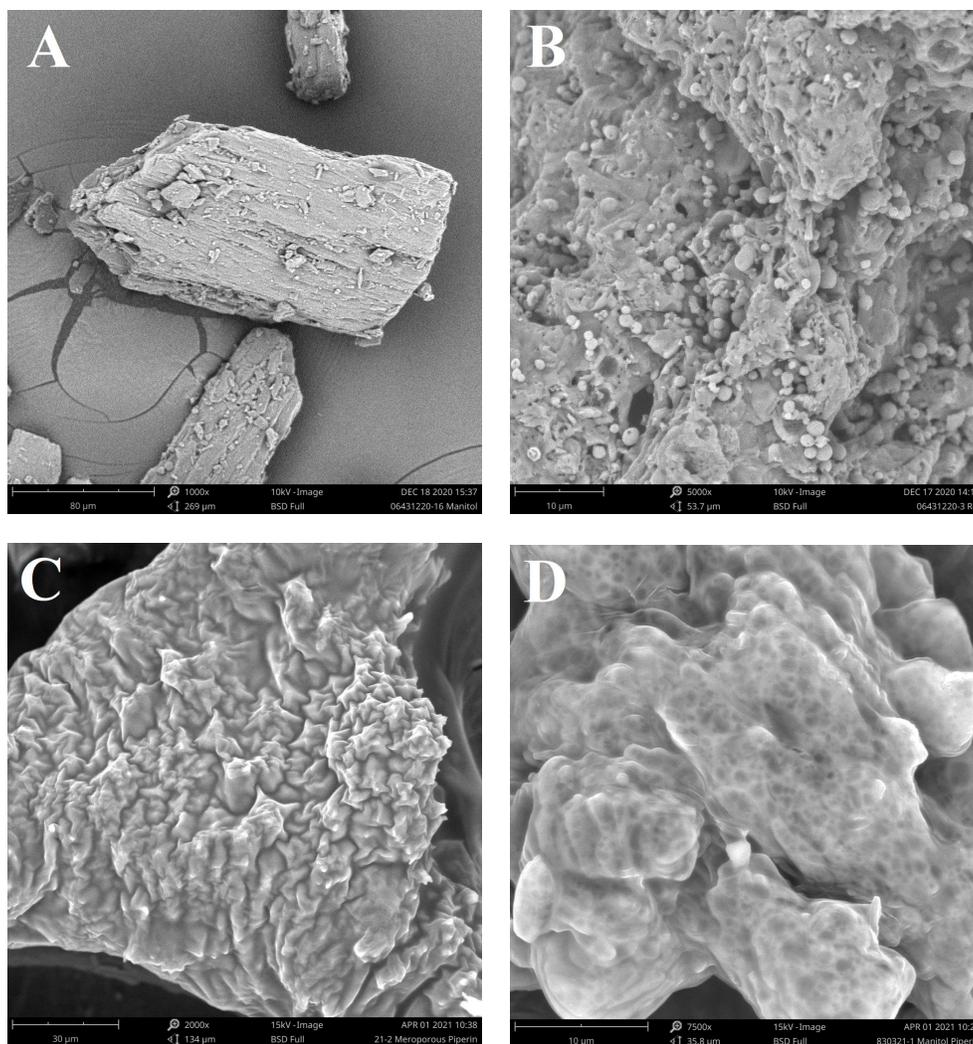


Figure 3. Scanning electron microscopy imaging of (A) D-mannitol, (B) mesoporous mannitol, (C) S-SNEDDS isolated piperine, and (D) mannitol loaded L-SNEDDS isolated piperine.

volume; however, mesoporous silica is expensive and has low water solubility (Millqvist-Fureby *et al.*, 2014). Mannitol is a porous adsorbent with high water solubility and can potentially be used as a carrier for controlled drug release in drugs with low water solubility. However, mannitol has a much lower specific surface area and a smaller pore volume than mesoporous silica. The solidification process produces highly porous mannitol with a high surface area of mesoporous mannitol by mixing D-mannitol with ammonium bicarbonate as a sublimation agent using a spray drying process to maximize D-mannitol porosity. Ammonium bicarbonate, which sublimes above approximately 36°C by breaking down into ammonia, carbon dioxide, and water, was used to induce pore formation during the drying process (Song *et al.*, 2018).

Scanning electron microscopy

SEM images of isolated piperine, D-mannitol, mesoporous mannitol, and S-SNEDDS can be seen in Figure 3. The surface morphology of isolated piperine and D-mannitol powder was most likely a characteristic description of the crystalline particles, irregular in shape with an uneven surface. The pore surface of

the mixture of D-mannitol with ammonium bicarbonate resulted in the number of open pores compared to without the addition. Sublimation and decomposition of ammonium bicarbonate during the spray drying process resulted in different pore sizes. The larger pore size was due to the space filled by ammonium bicarbonate before sublimation and ultimately changed the pore shape of D-mannitol (Song *et al.*, 2018).

Surface area analyzer

As shown in Table 1, the specific surface area of mesoporous mannitol (4.192 m²/g) was two times greater than that of D-mannitol (2.096 m²/g). The SEM test results in morphological studies show that mesoporous mannitol has a larger pore volume than D-mannitol. The larger specific surface area of mesoporous mannitol was due to a change in pore size. It was larger because the space of D-mannitol was filled with ammonium bicarbonate before the sublimation process (Song *et al.*, 2018). The larger the pore surface area of the carrier, the greater the number of L-SNEDDS contained in the pore carrier (Chavan *et al.*, 2015; Peng *et al.*, 2017). Based on these considerations, mesoporous mannitol adsorbent was used for the

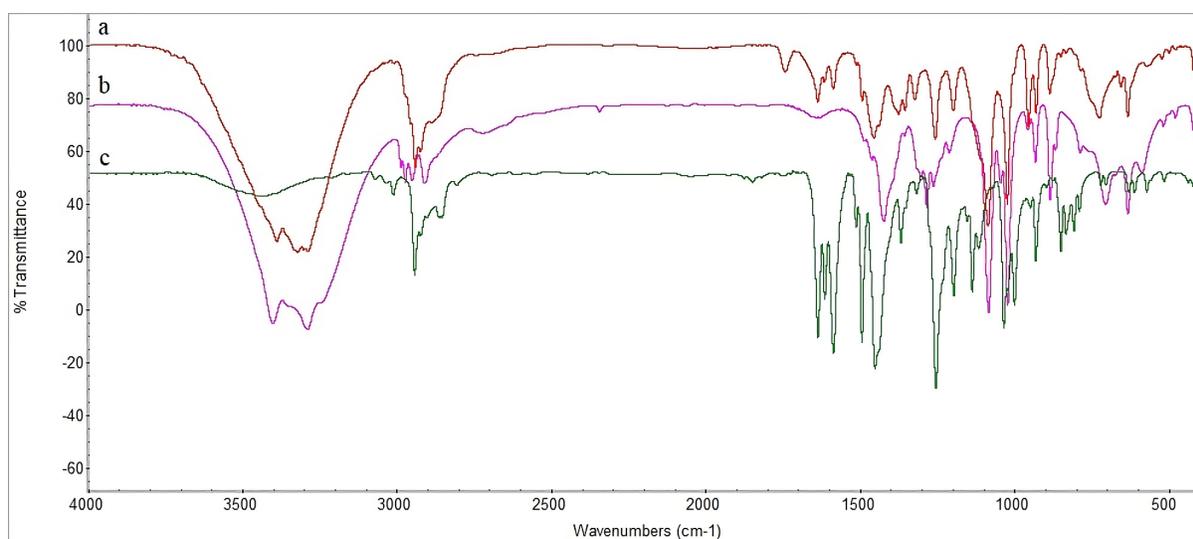
Table 1. Surface area analysis of D-mannitol and mesoporous mannitol by the BET method.

Powders	Specific surface area (m ² /g)	Pore volume (cc/g)	Pore radius (Å)
D-mannitol	2.096	0.009	26.889
Mesoporous mannitol	4.192	0.015	26.924

Table 2. The result of globule size, zeta potential, and statistical analysis S-SNEDDS in water, SGF pH 1.2, and SIF pH 6.8 compared with L-SNEDDS (*n* = 3).

Category	Globule size (nm)	Zeta potential (mV)	<i>p</i> -value emulsion droplet size (compared with L-SNEDDS)	<i>p</i> -value zeta potential (compared with L-SNEDDS)
S-SNEDDS in water	41.20 ± 10.17	-30.47 ± 2.74	0.289	0.133
S-SNEDDS in SGF pH 1.2	43.42 ± 0.58	+3.72 ± 0.46	0.017	0.006*
S-SNEDDS in SIF pH 6.8	33.09 ± 0.30	-8.78 ± 0.08	0.860	0.019

* *p*-value ≤ 0.05: significant.

**Figure 4.** FTIR analysis of (A) isolated piperine, (B) mesoporous mannitol, and (C) S-SNEDDS isolated piperine.

solidification process of L-SNEDDS isolated piperine in the hope that mesoporous mannitol was able to contain large amounts of L-SNEDDS isolated piperine.

Previous studies have reported the use of ammonium bicarbonate to increase the surface area of D-mannitol using the spray drying technique (Kuncahyo *et al.*, 2019; Peng *et al.*, 2017). Ammonium bicarbonate decomposes during the spray drying process, so it can act as a pore-forming agent. Peng *et al.* (2017) proved that the results of spray drying have no N atoms detected. Two factors must be considered in making porous mannitol: the outlet temperature during the spray drying process and the ratio of the mixture of D-mannitol and ammonium bicarbonate. The higher the outlet temperature used, the more the porous mannitol produced cracks, and the less ammonium bicarbonate used, the smaller the pore size formed. In addition, the more ammonium

bicarbonate is used, the more the small pore size is produced (Peng *et al.*, 2017).

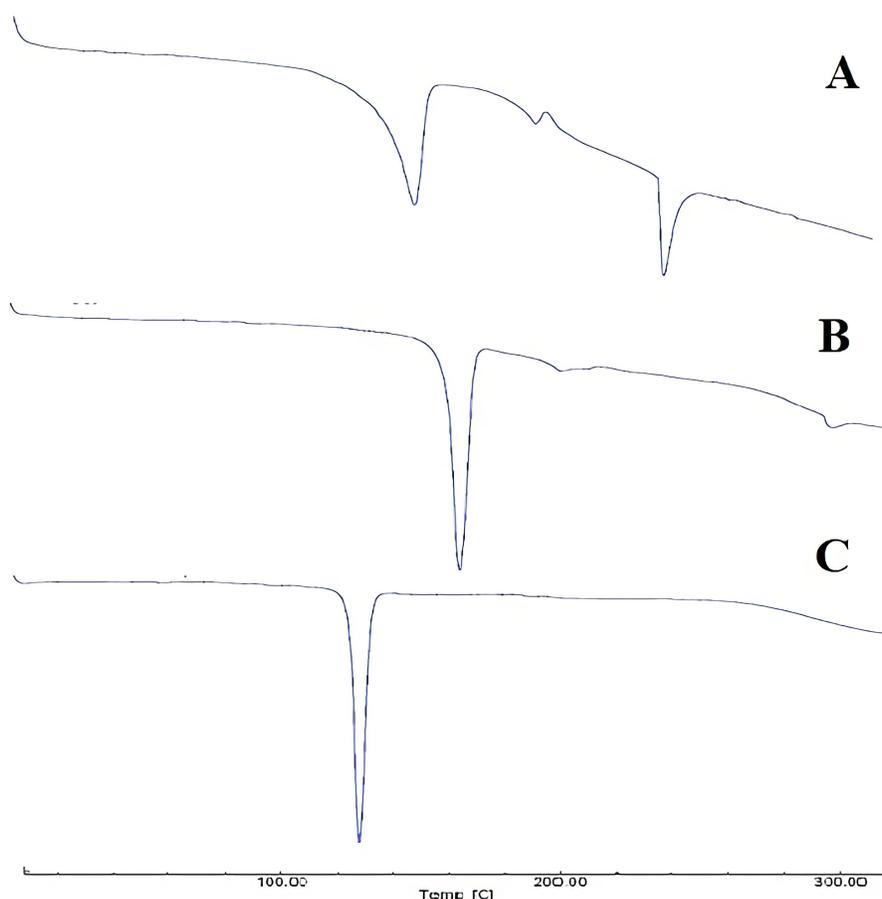
Formulation and characterization of S-SNEDDS

Reconstitution studies

The reconstitution rate is the time required for the S-SNEDDS to be uniformly dispersed in the aqueous medium. From the research, the reconstitution rate of S-SNEDDS isolated piperine was 40.03 ± 3.13 seconds. The S-SNEDDS isolated piperine reconstitution studies were dispersed in simulated gastric and intestinal fluids. Since S-SNEDDS was intended for oral drugs administration, it provided an overview of the particle size and zeta potential formed in gastric and intestinal fluids. The result analysis using paired *t*-test on SPSS v.24 software (IBM, USA) shows that the S-SNEDDS form did not change the particle size of L-SNEDDS in water, SGF pH 1.2,

Table 3. Dissolution efficiency 60 minutes (DE60) and MDT of isolated piperine, L-SNEDDS, and S-SNEDDS at SGF pH 1.2 and SIF pH 6.8.

Category	DE60 (%)	MDT (minute)
Isolated piperine at SGF pH 1.2	4.71 ± 0.03	61.44
Isolated piperine at SIF pH 6.8	6.19 ± 0.25	62.82
L-SNEDDS at SGF pH 1.2	54.72 ± 1.24	56.40
L-SNEDDS at SIF pH 6.8	54.82 ± 1.70	55.50
S-SNEDDS at SGF pH 1.2	62.18 ± 0.68	47.23
S-SNEDDS at SIF pH 6.8	63.88 ± 0.08	45.94

**Figure 5.** Thermal DSC profiles of (A) S-SNEDDS isolated piperine, (B) mesoporous mannitol, and (C) isolated piperine.

and SIF pH 6.8 (Table 2). Conversion of L-SNEDDS to solid dosage form did not significantly change the particle size of L-SNEDDS.

Fourier transform infrared

The FTIR spectrum serves to explain the interaction between L-SNEDDS and porous carriers. The vibrational spectra of isolated piperine and S-SNEDDS can be seen in Figure 4. The FTIR spectrum of the mesoporous mannitol shows that the host (Fig. 4) was determined by the specific peak vibrations, namely, the OH hydrogen bonds and the OH vibrations occurring at wavenumbers of 3,286 and 3,463 cm^{-1} , respectively. The peaks at 1,022 and 1,084 cm^{-1} vibrated from the C–O stretching. The alkyl vibrations of the

mesoporous mannitol were increased from 1,450–1,300 cm^{-1} . In addition, the peak vibrations of isolated piperine (Fig. 4) were determined by the presence of the functional group's CH, C–H₂, O=CN, C=C, =COC, and COC at the peaks of 2,800–3,000, 1,635, 1,495–1,589, 1,030–1,257, and 1,134 cm^{-1} , respectively.

The vibration spectrum showed that S-SNEDDS had a similar pattern with the spectrum pattern of isolated piperine and mesoporous mannitol, although the peak vibration of mesoporous mannitol was the most dominant. Thus, the specific peak vibration of the isolated piperine was not well observed. The FTIR spectrum results revealed no observed vibrational interactions in the incorporation of L-SNEDDS isolated piperine with pore carriers.

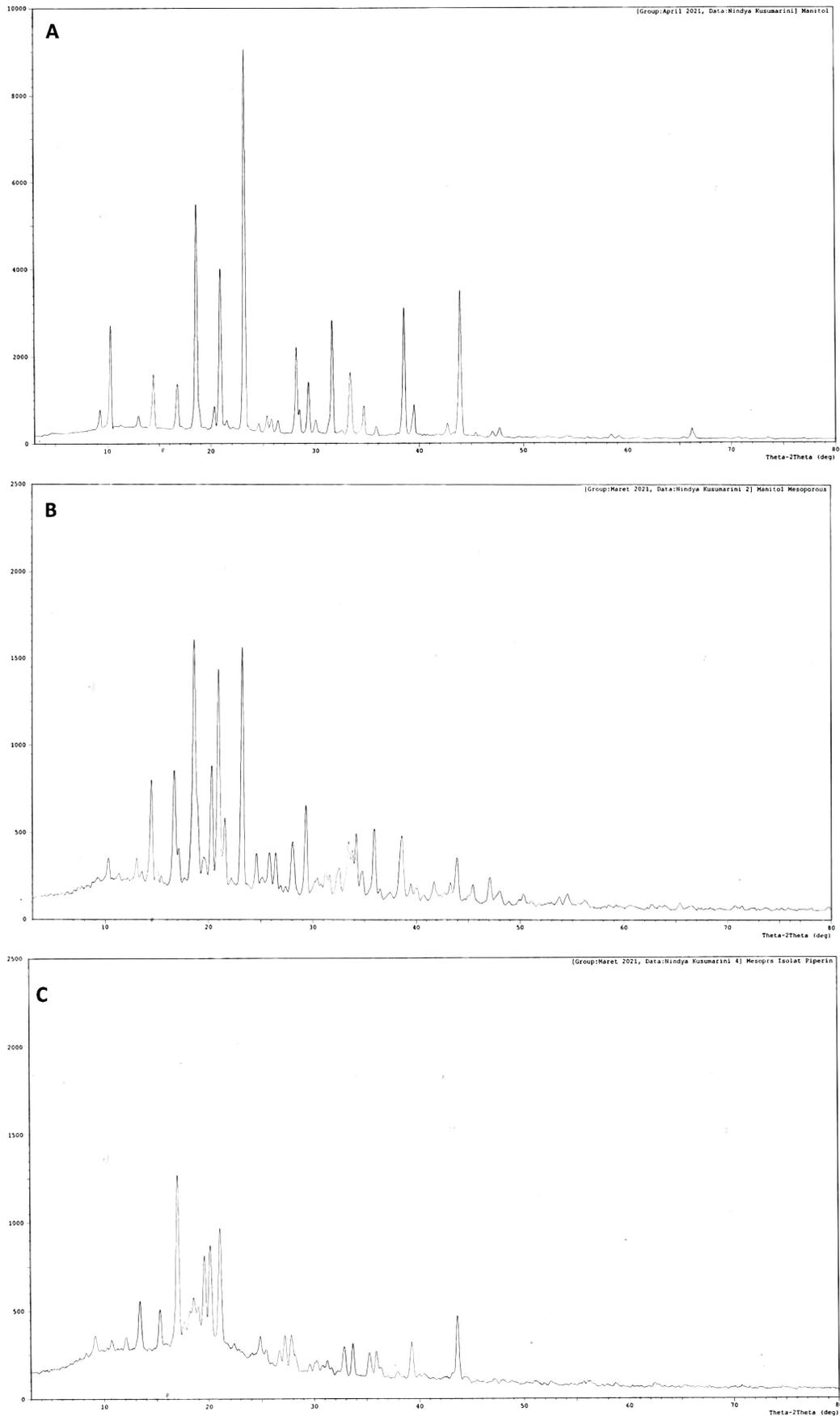


Figure 6. XRD diffraction patterns of (A) D-mannitol, (B) mesoporous mannitol, and (C) S-SNEDDS isolated piperine.

Further characterization was carried out to see the interaction between the two.

Scanning electron microscopy

Figure 3 presents D-mannitol, mesoporous mannitol, L-SNEDDS isolated piperine, and S-SNEDDS to understand their morphological characteristics. The morphology of mesoporous mannitol shows a rough, irregular surface shape, and it has holes in the carrier core space. It is different from the form of the D-mannitol structure that is irregular with a flat crystal surface. The morphology of S-SNEDDS shows that L-SNEDDS was contained in the empty space of the mesoporous mannitol pore. It produced larger particles. Meanwhile, the morphological form of directly mixed L-SNEDDS with D-mannitol resulted in small irregular particles. So that fewer L-SNEDDS was contained in the pores.

Differential scanning calorimetry

The thermal DSC profiles of isolated piperine, mesoporous mannitol, and S-SNEDDS can be seen in Figure 5. The thermal profile of the S-SNEDDS samples showed a shift in the endothermic peak of mesoporous mannitol at a temperature of 166.62°C, the melting point of mannitol, and shifted to a temperature of 154.07°C. Meanwhile, isolated piperine that melted at a temperature of 126.37°C–139.27°C did not appear in S-SNEDDS. It indicated a change in the physical state to nonamorphous crystals or irregular forms of molecular dispersions. In the thermal profile of the S-SNEDDS sample, there was a new endothermic peak with a melting point of 234.69°C, which was most likely a component of the SNEDDS constituent. The melting point of D-mannitol in this study was similar to previous studies on the melting point of D-mannitol, where the spray drying process did not change the melting point of D-mannitol (Burger *et al.*, 2000; Hulse *et al.*, 2009).

Powder X-ray diffraction

PXRD is the fastest and easiest method for determining basic information about the structure of a crystalline material (Brittain, 2001). Through X-ray diffraction, it could determine the presence of amorphous properties, polymorphisms, and crystal forms. Thus, this method was used to evaluate the polymorph content of each sample. The XRD reference pattern of each polymorph refers to research from Hulse *et al.* (2009). Based on his research, mannitol had three polymorphic forms, namely, α , β , and δ . The α -polymorph form had a peak position at 2θ 9.57 and 13.79, β -polymorph at 10.56, a relatively intense peak at 14.71, δ -polymorph with a very sharp peak at 9.74, and no peak until 14.66.

Based on the XRD results of the mesoporous mannitol, D-mannitol, and S-SNEDDS samples (Fig. 6), the D-mannitol sample contained two forms of polymorphism, namely, α -polymorph and β -polymorph. The spray drying process of a mixture of D-mannitol and ammonium bicarbonate of mesoporous mannitol samples produced one polymorph form, namely, β -polymorph, since the intensity was relatively reduced compared to the mannitol sample. In addition, it showed that samples containing α -polymorphs underwent a polymorphic transition into β -polymorphs during the spray drying process. The

finding is similar to Hulse *et al.* (2009). Finally, the XRD results of S-SNEDDS showed one polymorphic form, namely, β -polymorph, with an intensity lower than mesoporous mannitol. The not-sharp intensity in the S-SNEDDS may be because the drug crystals were converted to an amorphous state.

Test the content of isolated piperine in S-SNEDDS

The isolated piperine content determination in S-SNEDDS was carried out by dissolving 100.0 mg of S-SNEDDS into 10.0 ml of distilled water. Furthermore, the solution obtained was determined using HPLC with a validated method, using the regression equation $y = 148.11x + 8,208.5$ ($r = 0.9999$). This resulted in 9.53 ± 0.54 mg/ml of isolated piperine content in 100.0 mg S-SNEDDS. Then, the results were used to equalize the dose of S-SNEDDS in the dissolution test.

DISSOLUTION STUDY

Dissolution profiles of piperine, L-SNEDDS isolated piperine, and S-SNEDDS isolated piperine (containing 50.0 mg isolated piperine) in SGF pH 1.2 and SIF pH 6.8 are shown in Figure 7, and the DE60 and MDT values are shown in Table 3. The dissolution profile shows that S-SNEDDS and L-SNEDDS at pH 1.2 and 6.8 dissolution were faster than pure isolated piperine at pH 1.2 and 6.8. S-SNEDDS at pH 1.2 and 6.8 showed faster drug release than L-SNEDDS at pH 1.2 and 6.8. It is possible that L-SNEDDS has emulsified longer when in the dissolution medium than S-SNEDDS. The presence of mesoporous mannitol with high porosity is possible to accelerate the dissolution and emulsification between the dissolution medium and the SNEDDS carrier (Inugala *et al.*, 2015).

In the dissolution medium, S-SNEDDS disintegrates first and forms a fine emulsion in the GIT with mild agitation due to the normal mobility of the GIT. Once disintegrated, the fate of S-SNEDDS in the body will be the same as that of L-SNEDDS. The droplet size of the emulsion generated by S-SNEDDS should be equal to the resulting emulsion droplet size of L-SNEDDS. Based on the paired t-test analysis using SPSS v.24 software (IBM, USA), as shown in Table 2, the droplet size of S-SNEDDS emulsion was the same as L-SNEDDS at both pH 1.2 and 6.8. This shows that the change from liquid to solid did not change the droplet size of the emulsion, and the emulsion did not bind to the mesoporous mannitol pore carrier.

A previous study on the pharmacokinetics of SNEDDS preparations shows that linearly the SNEDDS preparations, which were able to increase dissolution *in vitro* when tested for pharmacokinetics, also showed an increase in peak maximum plasma concentrations (C_{max}) and area under the curve (Shao *et al.*, 2015). The pharmacokinetic profile of most SNEDDS preparations will have a double peak phenomenon. This is due to the presence of hepato-enteric circulation caused by bile elimination (Shao *et al.*, 2015). Piperine is rapidly absorbed through the GIT and can be detected in plasma 15 minutes after administration to rats (Bajad *et al.*, 2003). This is in line with Shao *et al.* (2015) on the study of intestinal absorption *in situ*. Suresh and Srinivasan (2010) stated that piperine does not undergo metabolic changes during the absorption process in the intestine but is metabolized rapidly in the liver and other tissues. In the body of piperine, piperine undergoes four metabolisms through

et al., 2015). Physical modification of D-mannitol has succeeded in increasing the surface area and pore characteristics. Thus, the higher the surface area, the more likely the L-SNEDDS is loaded into the porous carrier.

CONCLUSION

In conclusion, the solid formation of L-SNEDDS isolated piperine succeeded in increasing the dissolution of piperine and protecting piperine compounds from photosensitivity. The change in the form of L-SNEDDS to S-SNEDDS did not change the size of SNEDDS particles in SGF pH 1.2 and SIF pH 6.8. S-SNEDDS release of isolated piperine was slightly faster observed than L-SNEDDS at both pH 1.2 and 6.8. This may be the effect of increasing the specific surface area provided by the pore carrier. Mesoporous mannitol pore carriers have succeeded in forming good S-SNEDDS, which are amorphous and do not show interaction with L-SNEDDS constituent components as evidenced by compatibility assessment by FTIR, XRD, and DSC. The surface morphology of S-SNEDDS revealed that L-SNEDDS was successfully adsorbed on highly porous amorphous carriers. S-SNEDDS features a slightly rough surface indicating that the drug is loading efficiently. Thus, the mesoporous mannitol pore carrier can be used for the development of S-SNEDDS.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

AUTHOR CONTRIBUTIONS

All authors made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; took part in drafting the article or revising it critically for important intellectual content; agreed to submit to the current journal; gave final approval of the version to be published; and agree to be accountable for all aspects of the work. All the authors are eligible to be an author as per the international committee of medical journal editors (ICMJE) requirements/guidelines.

ETHICAL APPROVAL

This study does not involve experiments on animals or human subjects.

DATA AVAILABILITY

All data generated and analyzed are included within this research article.

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