Journal of Applied Pharmaceutical Science Vol. 12(06), pp 156-163, June, 2022 Available online at http://www.japsonline.com DOI: 10.7324/JAPS.2022.120615 ISSN 2231-3354



Hypolipidemic effect of [6]-Gingerol-loaded Eudragit polymeric nanoparticles in high-fat diet-induced rats and Gamma scintigraphy evaluation of gastric-retention time

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ARTICLE INFO

Received on: 12/09/2021 Accepted on: 02/01/2022 Available Online: 05/06/2022

Key words:

[6]-Gingerol, Nanoformulation, Hypolipidemic, Atorvastatin, Gamma scintigraphy.

ABSTRACT

[6]-Gingerol is the potent pharmacological active phytocompound of ginger. The essential role of [6]-gingerol in drug therapy is limited due to its poor water solubility and low bioavailability. Hence, [6]-gingerol-loaded Eudragit nanoparticles were prepared by using a single emulsification solvent evaporation method to overcome hepatic first-pass metabolism and to increase oral bioavailability. The study was designed to investigate the anti-hyperlipidemic potential of [6]-gingerol in rats fed a high-fat diet (HFD). The hypolipidemic potential of [6]-gingerol at a dose of 25 and 50 mg/kg was evaluated in HFD-induced hyperlipidemic rats. The results of the study were surprisingly intriguing with [6]-gingerol significantly ($p \le 0.005$), altering the serum total cholesterol (TC), triglycerides (TG), low-density lipoprotein cholesterol, and high-density lipoprotein cholesterol levels to near-normal in HFD-induced hyperlipidemic rats. The TC and TG were also significantly reduced after treatment with 25 and 50 mg/kg of [6]-gingerol as compared to standard atorvastatin treatment. Gamma scintigraphy results were promising with 24-hour gastric-retention time. The novel formulation [6]-gingerol-loaded Eudragit polymeric nanoparticle has a significant potential to be used as a green antihyperlipidemic agent.

INTRODUCTION

Atherosclerosis is the accumulation of lipids and fibrous elements in large arteries. It is an important contributor to the growing burden of cardiovascular diseases. The three major processes involved in the development of atherosclerosis include inflammation, oxidation, and hypercholesterolemia/hyperlipidemia (Katselou *et al.*, 2014). The elevated levels of total cholesterol (TC) along with high and low-density lipoprotein concentrations in the plasma are the major risk factors for ischemic heart disease by developing atherosclerosis, whereas an increased level of highdensity lipoprotein in the plasma has a protective role against atherosclerosis (Lei *et al.*, 2014). Hyperlipidemia is treated with therapeutic agents like statins, fibrates, niacin, and resins. However, these are also associated with several adverse effects. Therefore, alternative therapy serves as a better approach than conventional therapy in terms of safe and effective use. (Hu *et al.*, 2006).

The management of hyperlipidemia focuses on reducing the elevated plasma concentration of lipids by altering the biosynthesis and elimination of cholesterol. The various experimental studies have demonstrated the role of ginger (*Zingiber officinale*) as an antihyperlipidemic agent in high-fat diet (HFD) animal models. Ginger is the most commonly used

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species worldwide. [6]-Gingerol is the major bioactive, pungent, and pharmacological active component of ginger.

The active constituents of ginger are divided into volatile and nonvolatile components. The volatile constituents of ginger are different types of terpenoids, while the nonvolatile constituents include gingerols, shogaols, paradols, and zingerone (Ahmad *et al.*, 2015). Gingerol has a series of chemical analogs due to the presence of the hydroxyl group at different positions. The ginger analogs are [4]-gingerol, [6]-gingerol, [8]-gingerol, [10]-gingerol, and [12]-gingerol (Bhattarai *et al.*, 2007). The presence of aliphatic chain moiety with the hydroxyl group is responsible for the biological activity.

[6]-Gingerol-loaded nanoparticles were prepared with the combination of Eudragit RS100, polyvinyl alcohol, and organic phase. The Box–Behnken design was used to evaluate the role of independent variables on responses, including particle size, % entrapment efficiency (%EE), and zeta potential. The physicochemical properties of the optimized formulation were stable with an acceptable spherical nanoparticle. The particle size, %EE, and zeta potential of the optimized formulation were $323 \pm$ 2.36 nm, 82 ± 0.89 , and $+24 \pm 1.11$ mV, respectively. The present study investigates the hypolipidemic potential of [6]-gingerolloaded Eudragit nanoparticles in the HFD-induced hyperlipidemic animal model.

MATERIALS AND METHODS

Materials

[6]-Gingerol was purchased from Shaanxi Pioneer Biotech Co. Ltd., and polymer, solvents, and reagents were purchased from Sigma-Aldrich, India. All the other solvents and chemicals used in the experiment were purchased from Fisher Scientific, Mumbai, India.

Male Wistar albino rats (160-200 g) were used. Animals were procured from Central Drug Research Institute (CDRI), Lucknow, and approved by the Institutional Animal Ethics Committee (IAEC/SHUATS/PA/2017III/SVKS02, dated: April 10, 2017). The animals were kept under standard housing conditions in a polyacrylic cage (humidity = 60%-65% and room temperature = 24°C-27°C, with 12:12 light:dark cycle). A normal animal diet with purified water was provided. The experiment was conducted as per the guidelines provided by the Committee for the Purpose of Control and Supervision of Experiments on Animals.

Methods

Formulation of [6]-Gingerol-loaded Eudragit nanoparticles

We prepared [6]-gingerol-loaded Eudragit nanoparticles using the emulsification solvent evaporation method (Vardhan *et al.*, 2017). [6]-Gingerol (100 mg) and Eudragit (446 mg) were dissolved in acetone to form an organic phase that was added dropwise in an aqueous phase consisting of an aqueous solution of polyvinyl alcohol (4% w/v) and was stirred continuously to form a uniform mixture." The mixture was homogenized by high shear homogenizer (IKA) at 15,000 rpm for 10 minutes. The homogenized mixture was sonicated via an Ultra Probe Sonicator (UP50H, Hielscher) for 12.5 minutes. The obtained product was left overnight for evaporation of the organic solvent. The residue was centrifuged, lyophilized, and stored in an airtight container for further use.

In-vitro drug release profile

The dialysis membrane of 12,000–14,000 Da was used for the *in-vitro* drug release profile. The optimized formulation was kept in a dialysis membrane (bag) and immersed in a beaker containing phosphate buffer at pH 7.4. The beaker was placed on a magnetic stirrer for uniform mixing and temperature was maintained at $37^{\circ}C \pm 0.5^{\circ}C$. The samples were collected at 1, 2, 4, 8, 12, 16, 20, and 24 hours. The collected samples were analyzed using UV-visible spectroscopy at a wavelength of 430 nm.

Induction of hyperlipidemia

The efficacy of the prepared compound was tested against hyperlipidemia using a HFD animal model.

Hyperlipidemia was induced in animals by feeding cholesterol (500 mg/kg) and cholic acid (250 mg/kg) suspension in groundnut oil (10 ml/kg) once daily for 28 days (Bidkar *et al.*, 2012; Yan *et al.*, 2006).

The animals were divided into five different groups, with six animals in each group. Animals in Group I (non-hyperlipidemic control) were fed a normal pellet diet and treated with distilled water (vehicle); animals in Group II (hyperlipidemic control) were fed a HFD and treated with vehicle; and animals in Group III, IV, and V (treated groups) were fed a HFD and treated with [6]-gingerol of 25 and 50 mg/kg body weight and atorvastatin of 20 mg/kg body weight, respectively. Treatment was given orally, once daily for 28 days. On day 0, 7, 14, and 28, the blood samples were collected via retro-orbital sinus and triglycerides (TG), TC, high-density lipoprotein cholesterol (HDL-C), low-density lipoprotein cholesterol (VLDL-C) levels were analyzed using an automated analyzer (Beckman Coulter AU480).

Concluding the experiment, the animals were sacrificed. The heart, liver, and kidney organs were isolated for histopathological examination.

Cardiac risk index and Atherogenic index (AI)

AI = (TC – HDL-C)/HDL-C (Dobiasova and Frohlich, 2001; Lemieux *et al.*, 2001).

Coronary risk index (CRI) = TC / HDL-C (Lemieux *et al.*, 2001).

Histopathology

For histopathological analysis, the transverse sections of the organ were prepared with a thickness of 5 μ m. The hematoxylin and eosin-dyed sections were mounted with Canada Balson. The organ section was fixed in 10% v/v formalin and placed in paraffin. The slides were viewed by an optical microscope at 200× magnification (Donates, 2014).

Radiolabeling and scintigraphy evaluation

Gamma scintigraphy is a noninvasive technique to determine the release characteristics of the radiolabeled formulation and is a measure of gastrointestinal (GI) performance (Davis *et al.*, 1992; Wilding *et al.*, 2001). The optimized formulation was labeled with the radioactive substance ^{99m}Technetium (^{99m}Tc) using a stannous-based reduction method. ^{99m}Tc forms a covalent complex with the optimized formulation (Sharma *et al.*, 2017). The radiolabeling binding was determined by thin-layer chromatography using acetone and chloroform in a 7:3 ratio as mobile phase. The quantification of radiolabeling was done by gamma counter (Captintec, USA). The formulation–^{99m}Tc complex was administered orally to the rat for investigating the gastric-retention time of the radiolabeled formulation.

Statistical analysis

The results are presented in mean \pm SEM. The data was analyzed using one-way ANOVA, followed by Dunnett's test (GraphPad Prism 5). *p*-value ≤ 0.005 was considered significant.

RESULT AND DISCUSSION

The drug release kinetics were studied by the Higuchi model and Peppas–Korsmayer's model by comparing the regression coefficient value for zero-order and first-order (Fazil *et al.*, 2012; Masood *et al.*, 2013; Zhang *et al.*, 2013). The values closer to 1 are considered standard drug release kinetic data values. The release of [6]-gingerol from Eudragit-loaded nanoparticles was calculated using Peppas–Korsmeyer's kinetic model (Fig. 1). The regression coefficient of 0.918 was found to be significant for pharmacological action.

The plasma TC, TG, HDL-C, LDL-C, and VLDL-C levels of the non-hyperlipidemic control, hyperlipidemic control, [6]-gingerol (25 and 50 mg/kg), and atorvastatin-treated groups on day 0, 7, 14, 21, and 28 are shown in Table 1.

The plasma TC of Group (GP)-I (non-hyperlipidemic control) was found to be uniform and the average mean value was 67.42 ± 0.33 mg/dl from day 0 to 28, whereas the plasma TC of GP-II (hyperlipidemic control) was found to be 67 ± 2.63 mg/dl on day 0, which gradually increased to 119.83 ± 5.06 , 178.33 ± 4.08 and 210.33 ± 4.55 mg/dl on day 7, 14, and 28, respectively.

However, the plasma TC of GP-III (the hyperlipidemic-treated group with 25 mg/kg of [6]-gingerol) was found to be varied from 65.33 ± 4.409 to 92.83 ± 3.32 mg/dl and the plasma TC of GP-IV (the hyperlipidemic-treated group with 50 mg/kg of [6]-gingerol) varied from 67.5 ± 2.14 to 99.17 ± 2.104 mg/dl. The plasma TC of GP-V (the hyperlipidemic-treated group with 20 mg/kg of atorvastatin) was in the range of 65.17 ± 3.68 to 97.17 ± 4.29 mg/dl. Both the groups treated with [6]-gingerol, (GP-III and GP-IV) and the group treated with standard atorvastatin (GP-V) showed a marked increase in TC levels on day 7 and a significant reduction on day 14 and 28 of treatment.

The plasma TG levels of GP-I (non-hyperlipidemic control) was found to be uniform and the average mean value was 81.38 ± 0.20 mg/dl from day 0 to 28, whereas the plasma TG of GP-II (hyperlipidemic control) was found to be 83.67 ± 2.2 mg/ dl on day 0, which gradually increased to 131.83 ± 2.57 , $169 \pm$ 3.58, and 198.33 ± 4.39 mg/dl on day 7, 14, and 28, respectively. However, the plasma TG of GP-III (the hyperlipidemic-treated group with 25 mg/kg of [6]-gingerol) was found to be varied from 81.33 ± 1.58 to 116.17 ± 2.72 mg/dl and the plasma TG of GP-IV (the hyperlipidemic-treated group with 50 mg/kg of [6]-gingerol) varied from 82.33 ± 2.08 to 124 ± 2.34 mg/dl. The plasma TG of GP-V (the hyperlipidemic-treated group with 20 mg/kg of standard atorvastatin) was in the range of 78.5 ± 1.89 to $106.33 \pm$ 2.25 mg/dl. Both the groups treated with [6]-gingerol, (GP-III and GP-IV) and the group treated with standard atorvastatin (GP-V) showed a marked increase in TG levels on day 7 and a significant reduction on day 14 and 28 of treatment.

The plasma HDL-C levels of GP-I (non-hyperlipidemic control) was found to be uniform and the average mean value was 33.79 ± 0.54 mg/dl from day 0 to 28, whereas the plasma HDL-C of GP-II (hyperlipidemic control) was found to be 32.17 ± 1.11 mg/dl on day 0, which gradually decreased to 29 ± 1.06 , 27.17 ± 1.14 , and 25.33 ± 1.14 mg/dl on day 7, 14, and 28, respectively. However,



Cumulative drug release (%)

Figure 1. In-vitro release profile of [6]-gingerol from nanoformulation in phosphate buffer (pH 7.4).

Parameters	Groups	0 day	7th day	14th day	28th day
TC	GP I	66.67 ± 3.01	66.83 ± 2.36	67.83 ± 2.12	68.33 ± 1.89
	GPII	67 ± 2.63	119.83 ± 5.06	178.33 ± 4.08	210.33 ± 4.55
	GP III	65.33 ± 4.409	92.83 ± 3.32***	88.5 ± 2.012***	85.17 ± 2.182***
	GP IV	67.5 ± 2.14	99.17 ± 2.104**	96.33 ± 1.819***	93.17 ± 1.851***
	GP V	65.17 ± 3.68	97.17 ± 4.29***	92.83 ± 3.65***	87.33 ± 3.88***
TG	GP I	81.17 ± 1.19	80.83 ± 1.28	82 ± 1.32	81.5 ± 1.34
	GPII	83.67 ± 2.2	131.83 ± 2.57	169 ± 3.58	198.33 ± 4.39
	GP III	81.33 ± 1.58	116.17 ± 2.72***	106 ± 2.37***	96.67 ± 1.45***
	GP IV	82.33 ± 2.08	124 ± 2.34	115.83 ± 2.31***	109.67 ± 2.25***
	GP V	78.5 ± 1.89	106.33 ± 2.25***	98.17 ± 1.96***	$94 \pm 1.88^{***}$
HDL-C	GP I	32.17 ± 1.11	33.5 ± 0.67	35.33 ± 1.36	34.17 ± 1.25
	GPII	32.17 ± 1.11	29 ± 1.06	27.17 ± 1.14	25.33 ± 1.14
	GP III	31.83 ± 1.05	29.17 ± 0.95	27 ± 0.63	28.17 ± 0.4
	GP IV	36 ± 1.39	$32.83 \pm 0.17*$	31.33 ± 0.76	33.17 ± 1.25***
	GP V	34.17 ± 1.25	32.5 ± 1.18	32.33 ± 1.5*	34.33 ± 1.2***
LDL-C	GP I	22.33 ± 1.12	23 ± 1.29	22.5 ± 1.26	23.17 ± 1.33
	GPII	24.5 ± 1.87	92.83 ± 3.32	126 ± 3.36	149.83 ± 4.85
	GP III	23.17 ± 1.58	37.5 ± 0.62***	31.33 ± 0.76 ***	$29.17 \pm 0.87 ***$
	GP IV	22.17 ± 1.25	37.5 ± 1.52***	35 ± 1.24***	33.83 ± 1.01***
	GP V	23.83 ± 1.54	34.33 ± 1.09***	31.5 ± 0.67***	31 ± 0.82 ***
VLDL-C	GP I	14.67 ± 0.88	14.33 ± 0.8	14.17 ± 0.79	14.83 ± 0.4
	GPII	14.5 ± 1.41	18 ± 1.57	18.67 ± 1.28	21.17 ± 1.89
	GP III	15 ± 1.15	17.17 ± 0.87	$14.83 \pm 0.75*$	12.83 ± 0.91***
	GP IV	14.17 ± 0.79	16.83 ± 0.48	$14.67 \pm 0.42*$	14.17 ± 0.48 ***
	GP V	16.5 ± 0.67	15.67 ± 0.61	$14.67 \pm 0.88*$	12.83 ± 0.48 ***

 Table 1. Effect of [6]-gingerol on serum TC, TG, HDL-C, LDL-C, and VLDL-C levels of treated animals on day 0, 7, 14, and 28 of the HFD-induced animal (rat) model.

Values are expressed in mean \pm SEM (n = 6).

 $p \le 0.005$ (*statistically significant) compared with hyperlipidemic control group (non-treated) and with other treated groups.

the plasma HDL-C of GP-III (the hyperlipidemic-treated group with 25 mg/kg of [6]-gingerol) was found to be varied from 31.83 \pm 1.05 to 27 \pm 0.63 mg/dl and the plasma HDL-C of GP-IV (the hyperlipidemic-treated group with 50 mg/kg of [6]-gingerol) varied from 36 \pm 1.39 to 32.83 \pm 0.17 mg/dl. The plasma HDL-C of GP-V (the hyperlipidemic-treated group with 20 mg/kg of standard atorvastatin) was in the range of 34.17 \pm 1.25 to 32.33 \pm 1.5 mg/dl. In the GP-III (the hyperlipidemic-treated group with [6]-gingerol 25 mg/kg), no significant changes were observed in the plasma HDL-C levels as compared to the hyperlipidemic non-treated GP-III group; in both the non-treated GP-II group and treated GP-III group the plasma (HDL-C) levels showed a significant decline on day 7, 14, and 28, respectively. However, in the GP-IV (the hyperlipidemic-treated group with [6]-gingerol 50 mg/kg), the serum HDL-C levels significantly reduced on day 7 and 14 and showed a slight increase on day 28 of treatment. The GP-V (the hyperlipidemic-treated group with 20 mg/kg of standard atorvastatin) showed equal value reduction of HDL-C levels on day 7 and 14 with an increase on day 28, which was nearly similar to day 0 (HDL-C) levels. Hence, the plasma HDL-C levels in the [6]-gingerol-treated groups, GP-IV group, was found to be more promising than the GP-III group.

The plasma LDL-C levels of GP-I (non-hyperlipidemic control) were found to be uniform and the average mean value was 67.42 ± 0.33 mg/dl from day 0 to 28, whereas the plasma LDL-C of GP-II (hyperlipidemic control) was found to be 24.5 ± 1.87 mg/dl on day 0, which gradually increased to 92.83 ± 3.32 , 126 ± 3.36 , and 149.83 ± 4.85 mg/dl on day 7, 14, and 28, respectively. However, the plasma LDL-C of GP-III (the hyperlipidemic-treated group with 25 mg/kg of [6]-gingerol) was found to be varied



Figure 2. AI of GP-I (non-hyperlipidemic control), GP-II (hyperlipidemic control), GP-III (treated with [6]-gingerol 25 mg/kg), GP-IV (treated with [6]-gingerol 50 mg/kg), and GP-V (treated with atorvastatin 20 mg/kg). Values expressed as mean (n = 6). ** $p \le 0.005$ (statistically significant) when compared with GP II.

from 23.17 \pm 1.58 to 37.5 \pm 0.62 mg/dl and the plasma LDL-C of GP-IV (the hyperlipidemic-treated group with 50 mg/kg of [6]-gingerol) varied from 22.17 \pm 1.25 to 37.5 \pm 1.52 mg/dl. The plasma LDL-C of GP-V (the hyperlipidemic-treated group with 20 mg/kg of standard atorvastatin) was in the range of 23.83 \pm 1.54 to 34.33 \pm 1.09 mg/dl. Both the groups treated with [6]-gingerol, GP-III and GP-IV, and the group treated with standard atorvastatin, GP-V, showed a marked increase in LDL-C levels on day 7 and a significant reduction on day 14 and 28 of treatment.

The plasma VLDL-C levels of GP-I (non-hyperlipidemic control) were found to be uniform and the average mean value was 14.50 ± 0.12 mg/dl from day 0 to 28, whereas the plasma VLDL-C of GP-II (hyperlipidemic control) was found to be 14.5 ± 1.41 mg/dl on day 0, which gradually increased to $18 \pm$ $1.57, 18.67 \pm 1.28$, and 21.17 ± 1.89 mg/dl on day 7, 14, and 28, respectively. However, the plasma VLDL-C of GP-III (the hyperlipidemic-treated group with 25 mg/kg of [6]-gingerol) was found to be varied from 12.83 ± 0.91 to 17.17 ± 0.87 mg/dl and the plasma VLDL-C of GP-IV (the hyperlipidemic-treated group with 50 mg/kg of [6]-gingerol) varied from 14.17 ± 0.48 to 16.83 \pm 0.48 mg/dl. The plasma VLDL-C of GP-V (the hyperlipidemictreated group with 20 mg/kg of standard atorvastatin) was in the range of 12.83 ± 0.48 to 16.5 ± 0.67 mg/dl. Both the groups treated with [6]-gingerol, GP-III and GP-IV, showed a marked increase in VLDL-C levels on day 7, whereas the GP-IV group treated with standard atorvastatin showed a slight increase. However, VLDL-C levels were found to be gradually decreasing on day 14 and 28 in the [6]-gingerol-treated groups, GP-III and GP-IV, and standard atorvastatin group, GP-V, with a marked decrease was observed on day 28 of treatment.

AI and CRI are the predictive markers for evaluating the risk of atherosclerosis and coronary heart disease. The AI of the experimental groups is shown in Figure 2. In our study, it was found that the AI of the treated groups (GP-III and GP-IV) was significantly decreased as compared to the non-treated group (GP-II) and no significant difference was seen between the treated groups (GP-III and GP-IV) with the standard group (GP-V). The CRI of the experimental groups is shown in Figure 3. In the present study, it was found that the CRI of the treated groups (GP-III and GP-IV) showed a marked decrease as compared to the non-treated group (GP-II) and no significant difference was observed between the treated groups (GP-III and GP-IV) with the standard group (GP-V). However, among the treated groups, GP-IV exhibited greater significance as compared to GP-III. We found that AI and CRI had an affirmative correlation with the lipid surrogate markers.

Histopathology was performed on the group that had received the highest dose (50 mg/kg) of [6]-gingerol. The transverse sections of kidney, heart, and liver were prepared and observed. Figure 4 shows the observational findings of the kidney section. The slide showed normal glomeruli, focal hemorrhage, dilated blood vessels, tubules with minimal cloudy change, vessels normal, interstitial tissue minimal, and no inflammation/abnormality. Figure 5 shows the observational findings of the heart section. The slide showed congested blood vessels, focal myocardial inflammation, pericardium, and endocardium within normal limits. Figure 6 shows the observational findings of the liver section. The slide showed lobular architecture, portal triad normal, mild lymphocytic infiltrate in periportal area, and trabeculae show normal thickness. We found that the histopathological findings were nonsignificant.

Evaluation of gastric-retention time of formulation using scintigraphy technique

The radiolabeling efficiency of covalent complex formulation–^{99m}Tc was 93% at 4 hours, 91.7% at 8 hours, and 89% after 24 hours. The GI performance was determined by gamma



Figure 3. CRI of GP-I (non-hyperlipidemic control), GP-II (hyperlipidemic control), GP-III (treated with [6]-gingerol 25 mg/kg), GP-IV (treated with [6]-gingerol 50 mg/kg), and GP-V (treated with atorvastatin 20 mg/kg). Values expressed as mean (n = 6). * $p \le 0.005$ (statistically significant) when compared with GP II.



Figure 4. Transverse section of the kidney of animal treated with [6]-gingerol 50 mg/kg.

scintigraphy (Siemens, Germany). The real-time serial scanning was done at 4, 8, 12, and 24 hours after the oral administration of the complex. The changing intensity of impressions was recorded with time. The captured real-time image (Fig. 7) showed the 24-hour gastric-retention time.

The results of our study are substantiated by the study conducted by Nammi *et al.* (2009), in which the ethanolic extract of *Zingiber officinale* showed a protective effect against the HFD animal model on rats at doses of 100, 200, and 400 mg/ kg body weight. The elevated levels of glucose, TG, TC, LDL-C phospholipids, and free fatty acids after HFD treatment were decreased significantly when treated with ethanolic extract of *Zingiber officinale*, but there was no significant change in the HDL-C level. The study results are also corroborated by a study conducted by Kadnur and Goyal (2005), in which they observed



Figure 5. Transverse section of the heart of animal treated with [6]-gingerol 50 mg/kg.



Figure 6. Transverse section of the liver of animal treated with [6]-gingerol 50 mg/kg.



Figure 7. Gamma scintigraphy showing the serial scanning at 4, 8, 12, and 24 hours after oral administration of radiolabeled nanoformulation.

a significant decrease in fructose-induced elevation in lipid levels when treated with methanolic extract of *Zingiber officinale*. The study outcomes also bear resemblance with the study conducted by Bhandari *et al.* (1998), in which they investigated the hypolipidemic potential of ethanolic extract of *Zingiber officinale* (200 mg/kg, p.o.) in cholesterol-fed rabbits. The increased levels of TC, TG, phospholipids, and lipoproteins in serum and tissue after cholesterol feeding was significantly decreased by ethanolic extract of ginger in comparison with gemfibrozil as standard.

CONCLUSION

The present study demonstrated significant а antihyperlipidemic action [6]-gingerol of containing nanoformulation in HFD-induced rats. The [6]-gingerol-loaded Eudragit polymeric nanoparticles showed significant lipidlowering potential with reduced TC, TG, LDL-C, VLDL-C, and increased HDL-C levels. After 28 days of treatment with nanoformulation, the lipid profile of both nanoformulation-treated groups ($p \le 0.005$) significantly altered the plasma lipid levels to the normal range. No significant differences were observed in the antihyperlipidemic profile between the nanoformulationtreated groups (GP-III and GP-IV). However, GP-IV with a dose of 50 mg/kg was found to give more encouraging results as compared to GP-III with a dose of 25 mg/kg. Further studies are suggested for investigating the possible mechanism(s) of action and to develop a new sustained release dosage form. However, the findings are of great interest and clinical significance.

ACKNOWLEDGMENT

The authors express their gratitude to Prof. Avinash Chandra Pandey, Nano-Application Centre (NAC), Allahabad University, for his continued guidance and support.

CONFLICT OF INTEREST

The authors declare that the research work is novel and they have no conflict of interest.

FUNDING

The study was not sponsored by any government or industrial organization.

ETHICAL APPROVAL

The study was approved by the Institutional Animal Ethics Committee (IAEC) with Reg No.: IAEC/SHUATS/ PA/2017III/SVKS02.

AUTHORS' CONTRIBUTION

All authors have significantly contributed to the concept, design, acquisition, analysis, and interpretation of data; draft and critical revision of the manuscript for relevant intellectual content; and gave final approval for publication in the current issue of the journal. All authors are eligible to be an author as per the International Committee of Medical Journal Editors (ICJME) requirement/guidelines and are accountable for all aspects of the work.

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How to cite this article:

Singh V, Pandey H, Misra V, Tiwari V, Srivastava P, Singh D. Hypolipidemic effect of [6]-Gingerol-loaded Eudragit polymeric nanoparticles in high-fat diet-induced rats and Gamma scintigraphy evaluation of gastric-retention time. J Appl Pharm Sci, 2022; 12(06):156–163.