



Synthesis and evaluation of novel 4-anilinocoumarin derivatives as potential antimicrobial agents

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ABSTRACT

The development of potential antimicrobial agents is one of the critical needs to obviate microbial infection. Coumarins are acknowledged as the potential therapeutic regime for several ailments including microbial infection. Due to the microbial resistance of the existing pharmaceuticals, the present aim of the study is associated with the development of novel 4-anilinocoumarins and their exploration against several Gram-positive and negative microbial stains. A series of 4-anilinocoumarins derivatives was synthesized through Schiff base reaction and characterized by mass spectrometric, FT-IR, and ¹HNMR analytical techniques. Antibacterial activity of each synthesized compound was determined against *Staphylococcus aureus*, *Bacillus subtilis*, *Escherichia coli*, and *Pseudomonas aeruginosa*. The results showed that compound **4a'**, **4d'**, and **4h'** were showing significant ($p < 0.05$) inhibitory action with average zones of inhibition: 5.905 ± 1.011 , 6.145 ± 1.138 , and 6.595 ± 0.021 mm against *S. aureus*; 4.82 ± 0.042 , 3.97 ± 0.014 , and 5.335 ± 0.021 mm against *B. subtilis*; 3.8 ± 0.056 , 5.805 ± 0.728 , and 3.755 ± 0.091 mm against *E. coli*; and 5.51 ± 0.381 , 5.61 ± 0.001 , and 5.66 ± 0.014 mm inhibition, respectively. Hence, it can be demonstrated that 4-anilinocoumarins exhibit the potential role as antimicrobial agents.

INTRODUCTION

The World Health Organization (WHO) is an international organization responsible for maintaining the highest level of health and well-being. As per the WHO, microorganisms are one of the challenging threats that are affecting global health, exponentially. Antimicrobial resistance of modern medicine is a continuously growing crisis accredited because of their overuse or misapplication of these medications. In addition, economic burden and challenging regulatory requirements are the biggest lacking reasons that are obstacles to the development of new drugs in the pharmaceutical industry. Over the years, bacterial resistance to the existing pharmaceuticals has continuously increased, thus to minimize resistance and maximize the biological effectiveness

of antimicrobial agents, it is necessary to explore the mechanism of antibacterial resistance through another biomolecular approach and negate the effectiveness of the therapeutic drug (Alshibl *et al.*, 2020; Breijyeh *et al.*, 2020).

Staphylococcus aureus (*S. aureus*), *Bacillus subtilis* (*B. subtilis*), *Escherichia coli* (*E. coli*), and *Pseudomonas aeruginosa* (*P. aeruginosa*) are the most frequent etiological agents of bacteremia, the bacterial pathogens which are responsible for several ailments and even shut down the immune system of the body system. However, some of the common risk factors from such pathogens are neutropenia, cystic fibrosis, pneumonia, septicemia, and severe burns (Yayan *et al.*, 2015).

To evade such pathogenic microorganisms, it has become a challenging endeavor to the emerging healthcare system worldwide. These strains resist a wide class of existing antibacterial drugs such as fluoroquinolones, β -lactam antibiotics, glycopeptides, macrolides, and oxazolidinones (Fair and Tor, 2014).

However, the risk factors for an individual's morbidity and mortality are continuously increasing due to such pathogenic microbes. Pneumonia is one of the severe diseases caused by

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the transmission of *P. aeruginosa* in healthcare centres or even through medical sinks, disinfectants, equipment, and food. It has become a serious problem due to the difficult elimination of *P. aeruginosa* as it is resistant to many antibiotics such as penicillin G, aminopenicillin, β -lactamase inhibitors, cephalosporins (first generation), piperacillin, cefepime, aminoglycosides, carbapenems, colistin, etc. (Bush and Bradford, 2016). Therefore, it is important for choosing the right antibiotic to alleviate the trends in resistance of *P. aeruginosa* (Akter *et al.*, 2014). In addition, *E. coli* is a Gram-negative bacteria which is well recognized as a part of the normal intestinal flora; in case the strength becomes disbalance, it causes severe life-threatening sequelae (Bobak and Guerrant, 2014).

Coumarins are naturally and synthetically processed organic compounds and their derivatives have been poeticised for several past years for the treatment of microbial infection. Coumarins are acknowledged with excellent therapeutic action as antioxidant, anti-inflammatory, anticoagulant, antifungal, anticancer, and antibacterial activity (Pereira *et al.*, 2018). Because of their noteworthy biological importance, the synthesis of new coumarins derivatives is being assessed continuously as antibacterial and anti-inflammatory and are applied as modern therapy against the drugs which show antibacterial resistance (Lin *et al.*, 2012). Several synthesized coumarin derivatives have been explored to their pharmacological potential, and many of them have exhibited potential biological activity such as antibacterial and antifungal. The attention to such important compounds has been increased significantly because it is suggested that these compounds work against several viral infections such as HIV activity, etc. Furthermore, potential cytostatic activity is being exhibited by coumarin derivatives and therefore can be deliberated for anticancer therapy as potential candidates. Recently, coumarin derivatives of dimer and tetramer type were reported to possess HIV-1 integrase inhibitory activity. These facts urged the researchers to commence the synthesis of coumarin derivatives and explore their biological potential (López-Rojas *et al.*, 2018).

The fascinating properties of different coumarins can be attributed to their chemical reactivity as 2H-chromen-2-one core; its aromatic ring comprised a series of hydrophobics, π - π , CH- π , and cation- π interactions and the two oxygen atoms in the lactone ring can hydrogen-bond to a sequence of amino acid remains in different classes of enzymes and receptors. Furthermore, the double bond in the lactone provides assistance to create the planar system and consents for charge delocalization between the aromatic ring and the carbonyl group of the lactone which confers the characteristic fluorescence of this class of compounds (Stefanachi *et al.*, 2018). Considering the facts, the present aim of the study is associated with synthesis and exploration of 4-anilinocoumarin derivatives as a potential antimicrobial agent against the drugs-causing antibacterial resistance, even the development of a new antibacterial agent would facilitate an effective and economic antibacterial drug.

MATERIALS AND METHODS

Chemicals and microbial strains

Thin-layer chromatography (TLC) silica gel 60 F254 (Merck KGaA, 64271 Darmstadt, Germany), CDCl_3 as the solvent, TMS as an internal standard, *S. aureus* (MTCC No.

3161), *B. subtilis* (MTCC No. 441), *E. coli* (MTCC NO. 1687), and *P. aeruginosa* (MTCC NO. 424) and the reagents, solvents, and catalysts were of analytical grade and used directly for the analysis.

Chemistry

The synthesis process was accomplished in four steps: in the first step, compound **1a** was prepared via reaction of aniline with 4-hydroxycoumarin. Furthermore, this compound was treated with ethylchloroacetate in the presence of $\text{KOH}/\text{K}_2\text{CO}_3$, yielding an N-alkylated product (**2a**). The reaction of this product with hydrazine hydrate yielded desired hydrazide compound (**3a**). Schiff base derivatives (**4a–4j**) were obtained by reaction of **3a** with different aromatic aldehydes in the presence of glacial acetic acid in alcohol. The percentage purity of the compounds was checked by TLC using appropriate solvent systems where a single spot was considered the indication of a completed reaction. The structures of all the synthesized derivatives were elucidated by mass, IR, and ^1H NMR spectroscopical analysis. The steps included in the synthesis are described below.

Synthesis of 4-anilino-2H-1-benzopyran-2-one(1a')

A mixture of 4-hydroxycoumarin (0.01 mol) and aniline (0.01 mol) was stirred at 160°C for 20 minutes. The resulting mixture was dissolved in methanol (30 ml) and then 0.1 M aq. NaOH was added dropwise with stirring. After 5–10 minutes, the precipitate formed was washed, dried, and recrystallized. The completion of the reaction was confirmed by TLC using ethyl acetate and benzene as a mobile system.

Synthesis of ethyl [(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]acetate (2a')

4-anilino-2H-1-benzopyran-2-one (0.01 mol) was treated with ethylchloroacetate (0.01 mol) in the presence of $\text{KOH}/\text{K}_2\text{CO}_3$ and dry DMF (70 ml) was heated at 92°C – 94°C with stirring for 4 hours. After that, the solution was poured into ice water; the precipitate was filtered, washed with water, and dried to yield the N-substituted compound. The completion of the reaction was confirmed by TLC using acetone and n-hexane as a mobile system.

Synthesis of 2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]acetohydrazide (3a')

Compound **3'** was reacted with hydrazine (0.01 mol) in presence of ethanol and refluxed for 2 hours. The completion of the reaction was confirmed by TLC and the yield of the compound.

Synthesis of Schiff bases (4a'–4o')

Compound **4'** (0.01 mol) was reacted with a different aromatic aldehyde (0.01 M) in the presence of glacial acetic acid in alcohol and refluxed for 2–4 hours to yield corresponding Schiff bases. The steps involved in chemistry for the synthesis of Schiff base coumarin derivatives are shown in Figure 1.

Spectroscopical analysis of synthesized compounds

Mass spectrometric (MS) analysis

The MS analysis was carried out to identify the synthesis compounds based on their molecular mass obtained from the MS

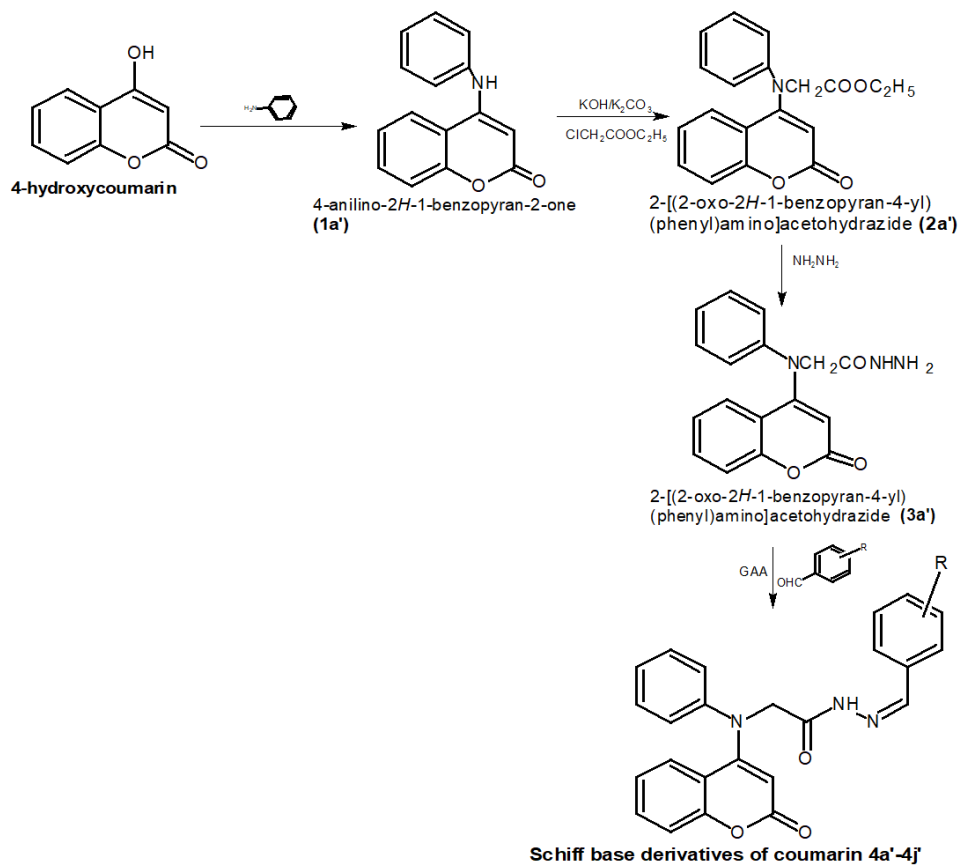


Figure 1. Schematic representation of the reaction process for the synthesis of Schiff base coumarin derivatives.

spectral data. The MS system is attached with Water's ACQUITY UPLCTM system (Waters Corp., MA) and adjoined with an auto-sampler, binary solvent delivery system, column manager, and MS detector. In brief, 1 mg/ml concentration were prepared in LCMS grade solvent for each sample; acetonitrile (A: 85%) and water (B: 15%) were used as chromatographic solvent run throughout a monolithic capillary silica-based C18 column [ACQUITY UPLC(R) BEH C18 1.7 μ m, 2.1 \times 100 mm]; the injection volume for the sample was set at 2 μ l. The flow rate of the nebulizer gas and cone gas was set at 500 l/hours and 50 l/hours, respectively. In mass analysis, electrospray ionization (ESI) was used as the ion source and the temperature for mass source was fixed at 120°C, while capillary and cone voltage was set at 3.0 and 40 KV, respectively. For collision, argon was active at a pressure of 5.5×10^{-5} torr. The obtained spectral data were interpreted and tentatively identified (Gaurav *et al.*, 2020).

FT-IR spectroscopy analysis of isolated compounds

The spectral analysis for synthesized compounds was carried out by using Win-IR, Bio-Rad FTS spectrophotometer. In brief, each sample was mixed with potassium bromide individually and proceeded ahead for spectroscopical analysis in the range of 4,000–400 cm^{-1} (Hosseini *et al.*, 2019).

¹H-NMR spectroscopic analysis of isolated compounds

NMR spectroscopical analysis of synthesized compounds was carried out as per the described method using Bruker Avance 500 MHz NMR spectrometer. Each sample (5 mg) was

dissolved in deuterated methanol (CD_3OD) as an NMR solvent, while tetramethylsilane was used as an internal standard. The samples proceeded for spectroscopic analysis by NMR technique (Prasad and Sati, 2011).

Antibacterial activity

In vitro antibacterial activity of the synthesized coumarin derivatives compounds was conducted through zone inhibition assay/well-diffusion assay against *S. aureus*, *B. subtilis*, *E. coli*, and *P. aeureginosa* bacterial strains using the standard protocol with some modification (Kumar *et al.*, 2020). In brief, the bacterial inoculum from each secondary strain culture (prepared from 100 μ l of primary suspension containing approx. 1×10^8 CFU/ml pathological tested bacteria) was evenly spread over the surface of the agar petri dish plates using a sterile cotton swab. Thereafter, four holes of 5 mm diameter were made using a sterile tip. 10 μ l of drug solution (1 mg/ml) was added into the wells and the controls well were treated with 10 μ l of autoclaved double-distilled water which was considered the control treatment. Plates were incubated for 72 hours under aerobic conditions and at 37°C temperature. The antimicrobial effect was determined by the clear zone in the agar, which was measured after the completion of treatment.

RESULTS AND DISCUSSION

Chemistry

During the synthesis process, 10 coumarin derivatives were synthesized through Schiff base reaction and characteristically

monitored by TLC to evaluate that there was no by-products formed. The completion of the reaction was checked by the TLC method. The percentage yield and melting points of each synthesized compound were determined. The resulted outcomes of the study are summarized in Table 1.

Spectroscopic analysis of synthesized compounds

In spectroscopic analysis, MS, FT-IR, and ¹H-NMR analyses were conducted for the characterization of synthesized coumarin derivatives. The predicted common coumarin derivative structure with atom numbering is shown in Figure 2.

2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-[(E)-phenylmethylidene] acetohydrazide (4a')

MS (ESI) m/z: 397.46 (M+1); (FTIR, ν_{\max} , cm⁻¹): 3,378.59, 3,315.11 (N/NH), 3,112.07 (CH₂), 1,735.99 (C=C/C=N), 1,650.43 (C=O), 1,583.01 (diketones), 1,492.48 (C-O-C), 1,339.97 (CH), 1,145.42, 1,087.35, 989.88, and 723.39 (aromatic, aliphatic amines, and aromatic C-H vibrations). ¹H-NMR (CD₃OD, 500MHz): δ 10.473, 8.421 (2H, 2s, NH, CH-16), 8.348 (1H, m, Ar, CH-5), 7.952 (2H, m, Ar, CH-2', 6'), 7.762, and 7.612, (4H, 2m, Ar, CH-7, Ar, CH-3', 4', 5'), 7.372 (2H, m, Ar, CH-6, 8), 7.037, 6.692, and 6.502 (5H, 3m, Ar, Aniline), 5.112 and 3.942 (3H, 2s, Ar, CH-3, CH₂).

2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-[(E)-(4-chlorophenyl)methylidene] acetohydrazide (4b')

MS (ESI) m/z: 432.07 (M+1); (FTIR, ν_{\max} , cm⁻¹): 3,455.70, 3,347.62 (=N/-NH), 3,042.91(CH₂), 1,722.95 (C=C/C=N), 1,638.17 (C=O), 1,473.29, and 1,357.36 cm⁻¹ (C-O-C and monosubstituted alkynes), 1,135.01, 1,023.55, 1,087.35, 989.88, and 725.83 (aromatic, aliphatic amines, and aromatic C-H and C-Cl). ¹H-NMR (CD₃OD, 500MHz): δ 10.473 and 8.421 (2H, 2s, NH, CH-16), 8.346 (1H, m, Ar, CH-5), 7.952 (2H, d, J=6.74 Hz, Ar, CH-2', 6'), 7.752 (1H, m, Ar, CH-7), 7.332 (4H, m, Ar, CH-6, 7, CH-3', 5'), 7.021, 6.695, and 6.537 (5H, 3m, Ar, Aniline), 5.664 and 3.925 (3H, 2s, Ar, CH-3, CH₂).

2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-[(E)-(3-chlorophenyl)methylidene] acetohydrazide (4c')

MS (ESI) m/z: 431.78 (M+1); (FTIR, ν_{\max} , cm⁻¹): 3,421.85, 3,318.74 (=N/-NH), 3,107.21 (CH₂), 1,635.32 (C=O), 1,553.77, 1,419.99, 1,335.48 (diketones, C-O-C, aromatic C-H, and mono-substituted alkynes), 1,093.48, 1,001.37, 830.95, and 759.32 (aromatic, aliphatic amines and aromatic C-H, C-Cl vibrations). ¹H-NMR (CD₃OD, 500MHz): δ 10.473 and 8.421 (2H, 2s, NH, CH-16), 8.346 (1H, m, Ar, CH-5), 7.632 and 7.385 (7H, m, Ar, CH-6, 7, 8; CH-2', 4', 5', 6'), 7.023, 6.68, and 6.515 (5H, 3m, Ar, Aniline), 5.676 and 3.937 (3H, 2s, Ar, CH-3, CH₂).

(E)-N'-(3-aminobenzylidene)-2-((2-oxo-2H-chromen-4-yl)(phenyl)amino)acetohydrazide (4d')

MS (ESI) m/z: 412.58 (M+1); (FTIR, ν_{\max} , cm⁻¹): 3,421.29 (=N/-NH), 3,053.01 (CH₂), 1,647.13 (C=O), 1,543.55, 1,413.93, and 1,329.17 (C-O-C, aromatic C-H, and monosubstituted alkynes), 1,125.36, 1,007.22, and 847.28 cm⁻¹ (aromatic, aliphatic amines, and aromatic C-H vibrations). ¹H-NMR (CD₃OD, 500MHz): δ 10.473 and 8.421 (2H, 2s, NH, CH-16), 8.346 (1H,

m, Ar, CH-5), 7.751 (1H, m, Ar, CH-7), 7.275 (3H, m, Ar, CH-6, 8; CH-5'), 7.275, 7.025, 6.901, 6.794, 6.695, and 6.539 (8H, 6m, Ar, aniline and aminobenzylidene), 5.672, 5.377, and 3.926 (5H, 3s, Ar, CH-3, NH₂, CH₂).

2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-[(E)-(4-hydroxyphenyl)methylidene] acetohydrazide (4e')

MS (ESI) m/z: 413.64 (M+1); (FTIR, ν_{\max} , cm⁻¹): 3,397.07 (=N/-NH), 3,355.58 (OH), 2,981.23 (CH₂), 1,648.13 (C=O), 1,523.55, and 1,405.68 cm⁻¹ (C-O-C, aromatic C-H / monosubstituted alkynes), 1,183.95, 1,063.79, 1,007.02, and 739.74 (aromatic, aliphatic amines, and aromatic C-H vibrations). ¹H-NMR (CD₃OD, 500MHz): δ 10.473, 9.521, and 8.421 (3H, 3s, NH, Ar-OH, CH-16), 8.346 (1H, m, Ar, CH-5), 7.752 (3H, m, Ar, CH-7, CH-2', 6'), 7.385 (2H, m, Ar, CH-6, 8), 7.155, 6.901, 6.804, and 6.521 (7H, 4m, Ar, aniline, and phenol), 5.672 and 3.926 (3H, 2s, Ar, CH-3, CH₂).

2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-[(E)-(3,4-dihydroxyphenyl)methylidene]acetohydrazide (4f')

MS (ESI) m/z: 430.26 (M+1); (FTIR, ν_{\max} , cm⁻¹): 3,463.05 (=N/-NH), 3,365.28 (OH), 3,082.75 (CH₂), and 1,879.36 (C=N), 1,673.42 (C=O), 1,588.07, 1,507.19, and 1,395.53 cm⁻¹ (C-O-C, aromatic C-H / monosubstituted alkynes), 1,068.27, 1,007.25, 827.53, and 791.39 cm⁻¹ (aromatic, aliphatic amines, and aromatic C-H vibrations). ¹H-NMR (CD₃OD, 500MHz): δ 10.473, 9.521, and 8.421 (4H, 3s, NH, OH, CH-16), 8.346 (1H, m, Ar, CH-5), 7.752 (1H, m, Ar, CH-7), 7.395 (2H, m, Ar, CH-6, 8), 7.175, 7.051, 6.752, and 6.536 (8H, 4m, Ar, aniline, and phenol), 5.672 and 3.926 (3H, 2s, Ar, CH-3, CH₂).

2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-[(E)-(4-dimethylaminophenyl)methylidene]acetohydrazide (4g')

MS (ESI) m/z: 440.82 (M+1); (FTIR, ν_{\max} , cm⁻¹): 3,341.16 (=N/-NH), 3,006.72 and 2,968.83 (CH₂/CH₃), 1,639.49 (C=O), 1,507.33, 1,439.05, and 1,396.47 (C-O-C, aromatic C-H / monosubstituted alkynes), 1,078.11, 1,006.25, 783.91, and 654.32 (aromatic, aliphatic amines, and aromatic compounds *cis*- and *trans*-substitution stretching vibrations). ¹H-NMR (CD₃OD, 500MHz): δ 10.473 and 8.421 (2H, 2s, NH, CH), 8.346 (1H, m, Ar, CH-5), 7.752 (1H, m, Ar, CH-7), 7.395 (4H, m, Ar, CH-6, 8; CH-2', 6'), 7.155, 6.891, 6.702, and 6.526 (7H, 3m, Ar, aniline, and dimethylamino-benzylidene), 5.772, 3.926, and 2.893 represent nine protons (9H, 3s, Ar, CH-3, CH₂, 2CH₃).

2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-[(E)-(3,4,5-trimethoxyphenyl)methylidene]acetohydrazide (4h')

MS (ESI) m/z: 487.34 (M+1); (FTIR, ν_{\max} , cm⁻¹): 3,427.62 (=N/-NH), 3,001.02, 2,965.22 and 1,681.05 (CH₂/CH₃ and C=O), 1,560.77, 1,501.08, 1,424.89, and 1,347.26 (diketones, C-O-C, aromatic C-H / monosubstituted alkynes), 1,058, 979.43, and 777.35 (aromatic, aliphatic amines, and aromatic compounds *cis*- and *trans*-substitution stretching vibrations). ¹H-NMR (CD₃OD, 500MHz): δ 10.473 and 8.421 (2H, 2s, NH, CH), 8.346 (1H, m, Ar, CH-5), 7.752 (1H, m, Ar, CH-7), 7.395 (2H, m, Ar, CH-6, 8), 7.211 (2H, s, Ar, CH-2', 6'), 7.114, 6.813, and 6.532 (5H, 2m, Ar-aniline), 5.672, 3.936, 3.846, and 3.873 (12H, 4s, Ar, CH-3, CH₂, 3CH₃).

Table 1. Chemical structures/IUPAC name and physicochemical characters of synthesized compounds.

Compound number	Structure	Name	m.p (°C)	Yield (%)	R _f value
4a'		2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-(E)-phenylmethylidene]acetohydrazide	200–202	63.25	0.65
4b'		2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-(E)-(4-chlorophenyl)methylidene]acetohydrazide	203–204	78.17	0.67
4c'		2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-(E)-(3-chlorophenyl)methylidene]acetohydrazide	195–196	78.86	0.73
4d'		(E)-N'-(3-aminobenzylidene)-2-((2-oxo-2H-chromen-4-yl)(phenyl)amino)acetohydrazide	201–203	70.19	0.69
4e'		2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-(E)-(4-hydroxyphenyl)methylidene]acetohydrazide	210–213	77.40	0.74
4f'		2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-(E)-(3,4-dihydroxyphenyl)methylidene]acetohydrazide	223–224	67.85	0.66
4g'		2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-(E)-(4-dimethylaminophenyl)methylidene]acetohydrazide	201–203	59.65	0.58
4h'		2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-(E)-(3,4,5-trimethoxyphenyl)methylidene]acetohydrazide	218–219	62.35	0.59
4i'		2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-(E)-(3,4-dimethoxyphenyl)methylidene]acetohydrazide	187	73.45	0.57
4j'		2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-(E)-(4-bromophenyl)methylidene]acetohydrazide	217–219	68.32	0.71

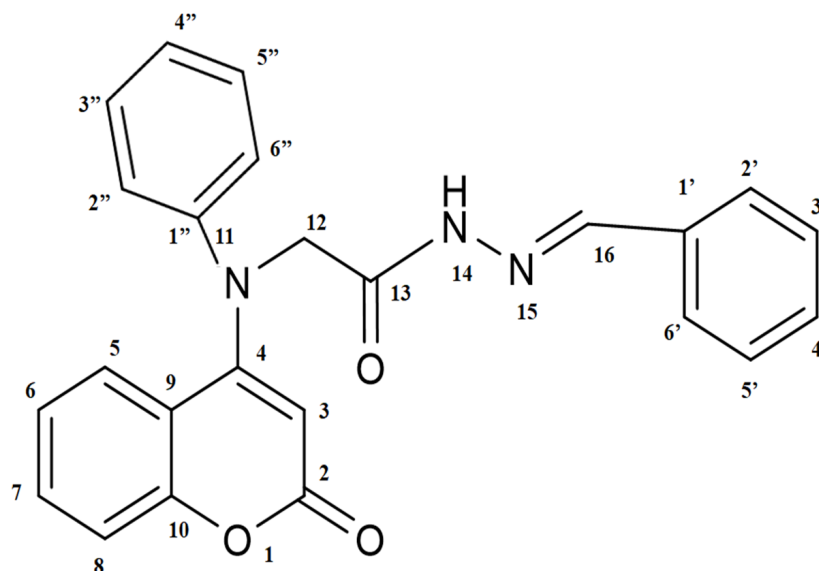


Figure 2. Predicted common structure of coumarin derivative with atomic numbering.

2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-[(E)-(3,4-dimethoxyphenyl)methylidene]acetohydrazide (4i')

MS (ESI) m/z : 458.06 (M+1); (FTIR, ν_{\max} , cm^{-1}); 3,345.11 (=N/-NH), 3,019.45, 2,929.05 ($-\text{CH}_2/\text{CH}_3$), 1,663 (C=O), 1,632.14, 1,527.33, 1,472.28, and 1,376.77 (diketones, C-O-C, aromatic C-H / monosubstituted alkynes), 1,198.46, 1,117.25, and 864 (aromatic, aliphatic amines, and C-H stretching vibrations). $^1\text{H-NMR}$ (CD_3OD , 500MHz): δ 10.473 and 8.387 (2H, 2s, NH, CH), 8.335 (1H, m, Ar, CH-5), 7.762 (1H, m, Ar, CH-7), 7.413 (3H, m, Ar, CH-2; CH-6, 8), 7.191 and 6.873 (2H, 2m, Ar, CH-5', 6'), 7.014, 6.703, and 6.532 (5H, 3m, Ar-aniline), 5.672, 3.925 and 3.716 (9H, 3s, Ar, CH-3, CH_2 , CH_3).

2-[(2-oxo-2H-1-benzopyran-4-yl)(phenyl)amino]-N'-[(E)-(4-bromophenyl)methylidene]acetohydrazide (4j')

MS (ESI) m/z : 476.01 (M+1); (FTIR, ν_{\max} , cm^{-1}); 3,391.15 (=N/-NH), 3,089.47 (CH_2), 1,721.35 (C=O), 1,539.92, 1,412.73 and 1,358.06 (diketones, C-O-C, aromatic C-H / monosubstituted alkynes) 1,099.63, 900.37, and 702.93 (aromatic, aliphatic amines, C-H and C-Br stretching vibrations). $^1\text{H-NMR}$ (CD_3OD , 500MHz): δ 10.473 and 8.387 (2H, 2s, NH, CH-16), 8.335 (1H, m, Ar, CH-5), 7.755 represents five proton (5H, m, Ar, CH-7, bromobenzylidene), 7.392 (2H, m, Ar, CH-6, 8), 7.163, 6.801, and 6.532 (5H, 2m, Ar-aniline), 5.672 and 3.925 (3H, 2s, Ar, CH-3, CH_2).

The structures of synthesized compounds with the melting point, percentage yield, and TLC retention factors (Rf) are summarized in Table 1.

Antibacterial activity of compounds

In vitro antibacterial activity of the synthesized Schiff base coumarin derivatives was carried out through zone inhibition assay/well-diffusion assay against *S. aureus*, *B. subtilis*, *E. coli*, and *P. aeruginosa* bacterial strain. The inhibitory action to

bacterial strain was considered with the direct proportion of obtained zone observed in the agar plate. The resulted data revealed that compounds **4a'**, **4c'**, **4d'**, **4g'**, and **4h'** were showing good activity, while other compounds exhibited lower activity against the targeted strains of microorganism. The zones of inhibition for the potentially active compounds were found as follows: 5.905 ± 1.011 , 5.595 ± 0.728 , 6.145 ± 1.138 , 5.285 ± 0.176 , and 6.595 ± 0.021 mm against *S. aureus*; 4.82 ± 0.042 , 4.6 ± 0.367 , 3.97 ± 0.014 , 5.25 ± 0.028 , and 5.335 ± 0.021 mm against *B. subtilis*; 3.8 ± 0.056 , 5.295 ± 0.063 , 5.805 ± 0.728 , 3.255 ± 0.021 , and 3.755 ± 0.091 mm against *E. coli*; and 5.51 ± 0.381 , 5.63 ± 0.226 , 5.61 ± 0.001 , 5.435 ± 0.121 , and 5.66 ± 0.014 mm inhibition, respectively. The resulted outcomes of the selected compounds may be the potential agents for antimicrobial activity. The outcomes of the antimicrobial study are shown in Figures 3 and 4.

DISCUSSION

Schiff bases are the ketone or aldehyde organic compounds that are abundantly used in the synthesis of several organic compounds together with coumarins. Although the classical synthesis mainly involves condensation of carbonyl compounds and is used for highly electrophilic carbonyl and nucleophilic amine compounds (Dixit *et al.*, 2010; Ronad *et al.*, 2008), in our study several coumarin derivatives were synthesized through a Schiff base reaction. During the synthesis process, the reaction was characteristically monitored by TLC to determine the purity of compounds. The synthesized coumarin derivatives were screened as a potential antimicrobial agent against the survivability of several *B. subtilis*, *S. aureus*, *E. coli*, and *P. aeruginosa* are considered as the major factor of several severe deleterious onsets inside the body (Ramachandran, 2014). Among the tested 10 coumarin derivatives, 3 compounds were found to exhibit significant ($p < 0.05$) inhibitory activity against each bacterial strain. The

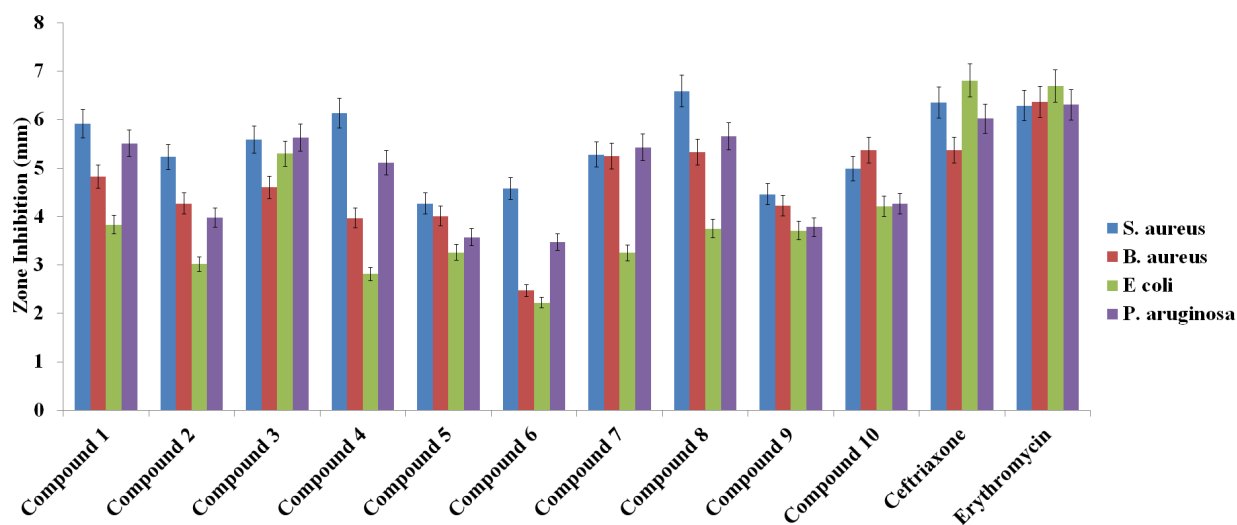


Figure 3. A bar graph representation of the antibacterial activity of synthesized compounds evaluated through zone inhibition assay against different strains of microorganisms.

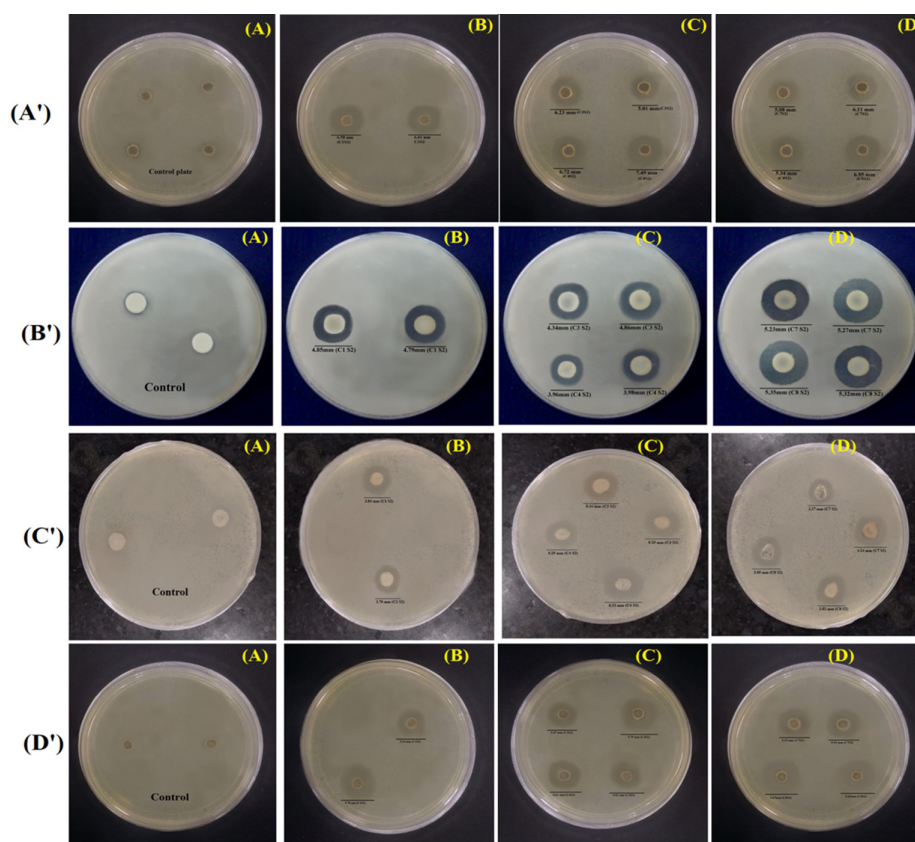


Figure 4. Antibacterial activity of coumarins derivatives (4a', 4c', 4d', 4g', and 4h'). (A') the control plate and drug-treated plates against *S. aureus*, whereas (A) shows the control plate; (B) shows the test plate of compound 4a'; and (C and D) show the test plate of compounds 4c', 4d', 4g', and 4h'. (B') shows the control plate and treated plates against *B. subtilis*. (A) shows the control plate; (B) shows the test plate of compound 4a'; and (C and D) show the test plate of compounds 4c', 4d', 4g', and 4h'. (C') shows the control plate and treated plates against *E. coli*. (A) shows the control plate; (B) shows the test plate of compound 4a'; and (C and D) show the test plate of compounds 4c', 4d', 4g', and 4h'. (D') shows the control plate and treated plates against *P. aruginosa*. (A) shows the control plate; (B) shows the test plate of compound 4a'; (C and D) show the test plate of compounds 4c', 4d', 4g', and 4h'.

outcomes revealed that compounds **4a'**, **4d'**, and **4h'** are the potential antibacterial agents against the survivability of Gram-positive and negative bacterial strains.

Furthermore, the outcomes were matched with another reported study which conferred that the antimicrobial activity of coumarins is symmetrized in a diversified array in the field of pharmaceutical sciences. In a study cited by [Završnik *et al.* \(2008\)](#), it is reported that tetramer 3,3',3'',3'''-(1,4-dimethylenphenyl)tetra (4-hydroxycoumarin) derivatives showed potential antimicrobial activity against several Gram-positive and Gram-negative bacteria such as *B. subtilis*, *S. aureus*, *E. coli*, and *P. aeruginosa* ([Završnik *et al.*, 2008](#)). In a reported study, 45 coumarin derivatives were tested against the same bacterial strains and reported the inhibitory properties of coumarins. So far, the most effective antibacterial coumarin derivative was represented as osthenol with MIC values ranging between 125 and 62.5 µg/ml. Furthermore, the authors claimed that the synthesized coumarins are the potential candidate for antibacterial against these strains ([De Souza *et al.*, 2005](#)). A study conducted by [Kawase *et al.* \(2001\)](#) evaluated that the substituents ester or carboxylic acids on the coumarin ring exhibits potent inhibitory activity against both several Gram-positive and negative microbial strains. In addition, the presence of phenolic hydroxyl or carboxylic acid functional group of coumarins are necessary to possess higher activity against *Helicobacter pylori* ([Kawase *et al.*, 2001](#)).

A study conducted by [Alshibl *et al.* \(2020\)](#) evaluated the antioxidant, antibacterial, and anti-inflammatory activity of the several synthesized compounds. The outcomes of the study revealed that among a total series of the coumarin derivatives, Coumarin-sulfonamide compounds **8a–d** confirmed with the significant antioxidant activity against DPPH free radicals, while **7c,d**, **8c,d**, and **9c,d** unveiled antimicrobial activity almost equal or greater than the antimicrobials against used as standards for tested microorganism. Because of the anti-inflammatory evaluation, pyranocoumarins and coumarin-sulfonamide compound **9a** exhibited more significant antiproteinase activity than aspirin. Furthermore, the authors claimed that five compounds exhibited the potent activity than aspirin. *In vivo* outcomes for evaluation of anti-inflammatory activity assessed pharmacologically on formaldehyde-induced rat paw edema and revealed potent inhibitory activity against the induced edema. In addition, pyranocoumarin derivative **5a** and coumarin-sulfonamide derivative **8d** were found more active toward COX-2 isozyme ([Alshibl *et al.*, 2020](#)).

Therefore, it can be demonstrated that coumarins derivative is the most potential candidate as an antioxidant, anti-inflammatory, and antibacterial agent. In addition, the outcome from the present study revealed coumarin derivatives as another alternative for antimicrobial activity. The developed coumarins derivatives can be assisted as auspicious candidates for more potent and highly effective antimicrobial agents.

CONCLUSION

The present study enlightens that among the series of different synthesized 4-anilinocoumarin derivatives, compounds **4a'**, **4d'**, and **4h'** are the most potent antimicrobial agents against Gram-positive and negative bacterial strains which exhibit significant even higher antibacterial effect than the standard drugs.

In addition, it can be demonstrated that the synthesized coumarin derivatives cover those antimicrobial agents that shows resistance even restricted to provide significant therapeutic effect.

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All authors made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; took part in drafting the article or revising it critically for important intellectual content; agreed to submit to the current journal; gave final approval of the version to be published; and agree to be accountable for all aspects of the work. All the authors are eligible to be an author as per the international committee of medical journal editors (ICMJE) requirements/guidelines.

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The authors report no financial or any other conflicts of interest in this work.

ETHICAL APPROVALS

This study does not involve experiments on animals or human subjects.

DATA AVAILABILITY

All data generated and analyzed are included within this research article.

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