

3D-QSAR analysis of pyrimidine derivatives as AXL kinase inhibitors as anticancer agents

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ABSTRACT

AXL kinase receptor belongs to the TAM family of receptor tyrosine kinases (RTKs). Different types of cancer namely breast cancer, osteosarcoma, acute myeloid leukemia, colorectal cancer and non-small cell lung cancer (NSCLC) manifest overexpression of AXL receptor. Moreover, AXL kinase overexpression leads to tumor angiogenesis & resistance to chemotherapeutic agents and reduces the antitumor immune response. Therefore, AXL kinase has emerged as the potential and attractive target for the treatment of cancer. The present study is based on the correlation between the structural parameter and biological activity of the compounds using the 3D-QSAR technique. In this technique, pyrimidine derivatives and their inhibitory activity against AXL kinase receptor were chosen as independent and dependent variables respectively. Based on the investigation, the structural requirements for AXL kinase inhibition were recognized. Here, CoMFA and CoMSIA analysis were used for the execution of the 3D-QSAR model. The training and the test set pyrimidine derivatives were used for the generation and validation of QSAR model respectively. Dataset alignment was performed using the lowest energy conformer of the most active compound. CoMFA, as well as CoMSIA, model have encouraging values of the cross-validation coefficient (q^2) 0.700 and 0.622 and conventional correlation coefficient (r^2) 0.911 and 0.875 independently. Furthermore, values of r^2_{pred} were obtained as 0.709 and 0.668 respectively. Outcomes of the QSAR models and contour maps may be used for discovery of new AXL kinase inhibitors as potent anticancer agents.

INTRODUCTION

Receptor tyrosine kinases (RTKs) are the type of multi-domain transmembrane proteins. These proteins are the sensor for extracellular ligands (O'Bryan *et al.*, 1991). When ligands bind to the active site of the receptor, the receptor dimerization and activation of intracellular kinase domain were achieved. This biochemical process leads to the recruitment, phosphorylation, and activation of multiple downstream signaling cascades. The AXL tyrosine kinase receptor is a member of the TAM (TYRO3-AXL-MER) group of RTKs and emerged as the promising therapeutic target for cancer therapy (Graham *et al.*, 2014; Lai *et al.*, 1994). Other than AXL kinase, TYRO-3 (also known as Brt, Dtk, Rse, Sky and Tif), and MER (also known as Eyk, Nym and Tyro12)

receptors are also part of TAM receptor kinase. The overexpression of AXL has been reported in different types of cancer namely non-small cell lung cancer (NSCLC), osteosarcoma, breast cancer, acute myeloid leukemia, prostate and colorectal cancer (Zhang *et al.*, 2013; Paccetz *et al.*, 2013; Ou *et al.*, 2011; Yuen *et al.*, 2013). Further, epithelial-mesenchymal transition (EMT) because of AXL signaling mechanism in tumor cells leads to the development of drug resistance to targeted therapies and chemotherapy (Hong *et al.*, 2008; Zhang *et al.*, 2012; Zhou *et al.*, 2016). Most of AXL kinase inhibitors were discovered due to the similarity in the kinase domain of AXL receptor with MET or MER receptors (Gay *et al.*, 2017). Bemcentinib (BGB324 or R428) is one of the most advance AXL selective inhibitor. It is an oral small molecule in the second phase of the clinical trial for NSCLC management (Ben-Batalla *et al.*, 2013). Non-selective AXL kinase inhibitors include foretinib, carbozantinib, merestinib, amuvatinib, bosutinib, gilteritinib etc. (Eder *et al.*, 2010; Hart *et al.*, 2013; Lee *et al.*, 2013; Yan *et al.*, 2013; Mahadevan *et al.*, 2007; Mori *et al.*, 2014). The structures of AXL kinase receptor inhibitors are depicted in Figure 1.

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The selective and potent inhibitory activity of the series of novel pyrimidine derivatives towards AXL kinase has been reported (Mollard *et al.*, 2011). The correlation of the structural parameter (independent variable) with the biological activity (dependent variable) of the derivatives was performed using the 3D-QSAR technique. Here, CoMFA and CoMSIA methods were accomplished for 3D-QSAR model development. Comparative molecular field analysis (CoMFA) is the type of 3D-QSAR technique where, steric and electrostatic fields were used as an independent variable against the biological activity of compounds (Cramer *et al.*, 1988). Whereas, CoMSIA is improved method of CoMFA in which other than steric and electrostatic, hydrophobic, hydrogen bond donor and hydrogen bond acceptor were used as an independent variable for the correlation with the biological activity of the compounds (Klebe *et al.*, 1994).

We have generated the quantitative structure-activity relationship (QSAR) model of pyrimidine derivatives for the prediction of biological activity. Based, on the QSAR model 3D contour maps of steric, electrostatic, hydrophobic, H-bond donor and H-bond acceptor fields were generated and SAR was

established. These details could be applied to modify the structure and improve the biological activity of pyrimidine derivatives against AXL kinase receptor.

MATERIALS AND METHODS

Dataset preparation

We have collected the pyrimidine derivatives as AXL kinase inhibitors for this study from the literature (Mollard *et al.*, 2011). The pyrimidine derivatives were bifurcated into approximate 70% of the training set (18 derivatives) and approximate 30% of the test set (7 derivatives). Compounds in the test set have a range of biological activity similar to the training set. Derivatives present in the training set and test set were used for the generation and validation of QSAR model respectively. The IC_{50} value of all derivatives was transformed into pIC_{50} ($-\log IC_{50}$) value. These transformed pIC_{50} values were utilized as a dependent variable for CoMFA and CoMSIA model generation (Table 1) (Liu *et al.*, 2011). Table 1 depicts the structure of all pyrimidine derivatives along with their biological activity against AXL kinase receptor.

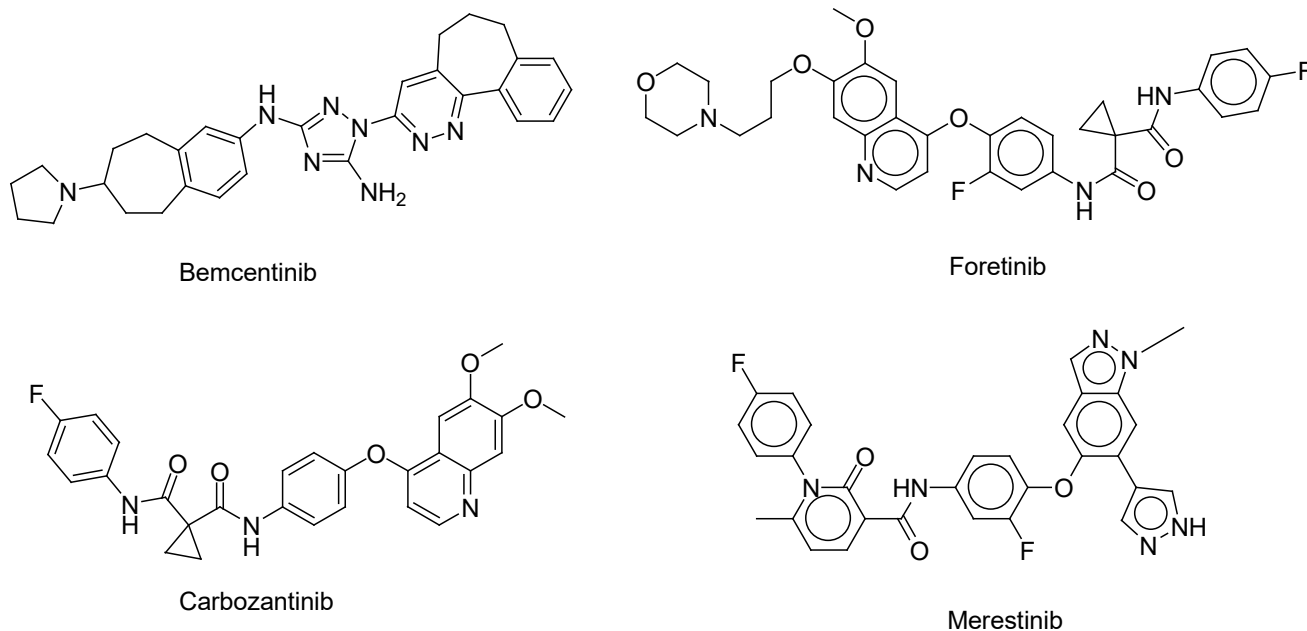


Fig. 1: Chemical structure of AXL tyrosine kinase inhibitors.

Computational study and alignment of database

SYBYL-X 2.0 from Tripos Inc. was used to perform the 3D-QSAR study (SYBYL X Molecular Modeling Software). In the SYBYL software, SKETCH function was utilized for construction of the 3D structure of all pyrimidine derivatives. Gasteiger Huckel (GH) charges along with the TRIPOS force field was applied to all designed derivatives for structure optimization process. Further, minimization was carried out by Powell conjugated gradient algorithm method (Clark *et al.*, 1989). Structure alignment is the utmost important step in QSAR study. Here, Distill alignment function was carried out. Alignment was executed by selecting highly potent pyrimidine derivative from the dataset as template structure (Raichurkar *et al.*, 2003). Figure 2 and Figure 3 represent

a common fragment of AXL kinase inhibitors and all aligned compounds respectively.

CoMFA and CoMSIA model

Three-dimensional cubic lattice with 2 Å grid space, which was extended to 4 Å for the aligned dataset in X, Y, and Z three axis, was defined automatically to calculate the CoMFA and CoMSIA fields. Here, Sp^3 carbon atom with the radius of 1.52 Å and +1 charge was used as probe atom under CoMFA analysis (Sridhar *et al.*, 2011). In CoMFA analysis, for the derivation of steric and electrostatic fields, interaction energy was calculated using the Tripos force field. ± 30 kcal/mol cut-off value was used for the steric and electrostatic fields respectively. At the lattice

intersection, Lennard-Jones and Columbic potentials were used (Bush *et al.*, 1993).

In the CoMSIA, five descriptors namely steric, electrostatic, hydrophobic, H-bond donor and H-bond acceptor were used (Myint *et al.*, 2010). For hydrophobic, H-bond donor

and H-bond acceptor, probe atom with +1 charge was used by default. While all other parameters are similar as in the CoMFA model, only the value of attenuation factor (α) was set to 0.3 for ease of functioning (Buolamwini *et al.*, 2002).

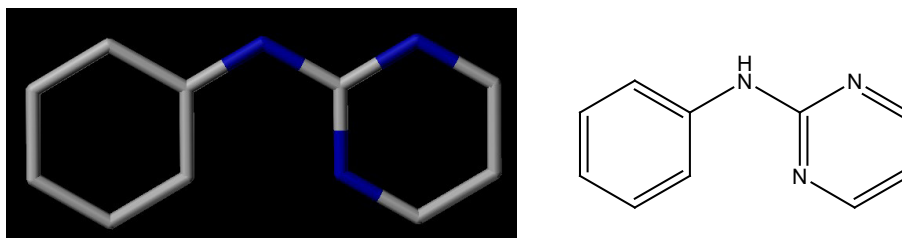


Fig. 2: The common fragment of AXL kinase inhibitors for alignment of the database to perform QSAR study.

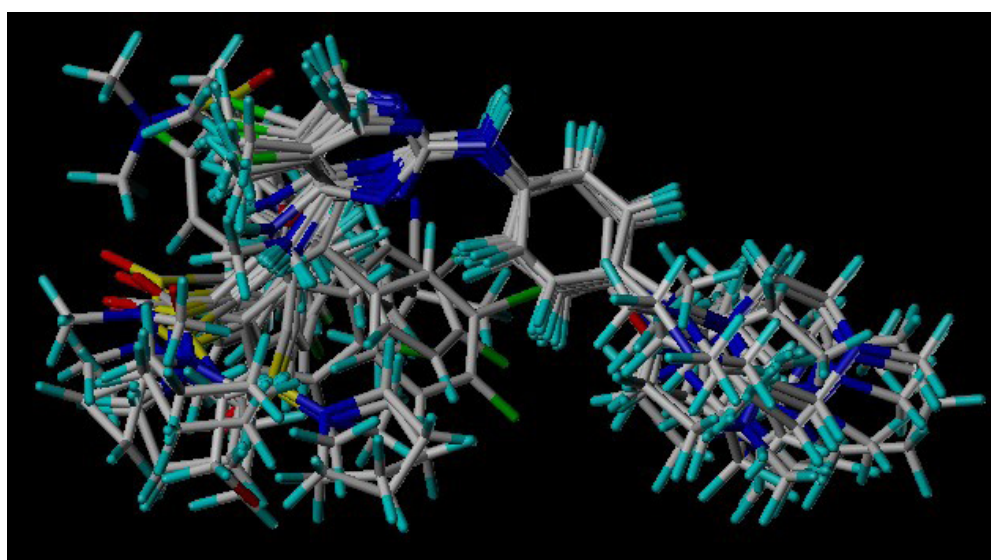


Fig. 3: Dataset alignment for 3D-QSAR (CoMFA and CoMSIA) study.

PLS analysis

PLS (Partial Least Square) analysis is useful when so many highly colinear factors are present. It is an extension of multiple linear regression analysis (MLR) method. The 3D-QSAR model was built using PLS analysis. Here, the values of CoMFA and CoMSIA were used as an independent variable, while AXL kinase inhibitory activity was used as a dependent variable for the correlation of the structural parameters with its biological activity. If the QSAR model is developed using the optimum number of components with a high value of cross-validated coefficient (q^2) and low standard errors, then chances of overfitted models are very less. In cross-validated coefficient (q^2) method, one compound was removed from the set and by use of all other compounds, the model was generated. The activity of the elided compound was predicted using the developed QSAR model. If methods give a q^2 value > 0.5 and $r^2 > 0.616$, models are termed as acceptable (Golbraikh *et al.*, 2002). It is estimated by the following equation:

$$q^2 = 1 - \frac{\sum (Y_{\text{predicted}} - Y_{\text{observed}})}{\sum (Y_{\text{observed}} - Y_{\text{mean}})} \quad (1)$$

The optimum number of component (ONC) was used to calculate the value of r_{ncv}^2 (non-cross validated correlation coefficient). Further, the robustness of the developed model was checked by bootstrap analysis. In this method, from the entire dataset random compounds were selected. This method was undertaken many times (minimum 100 times for more accuracy). In each run, some compounds were elided from the dataset while some compounds were included in the dataset. The value of r_{bs}^2 from bootstrap analysis and r_{cv}^2 from cross-validation were determined (Lu *et al.*, 2010).

Predictive correlation coefficient (r_{pred}^2)

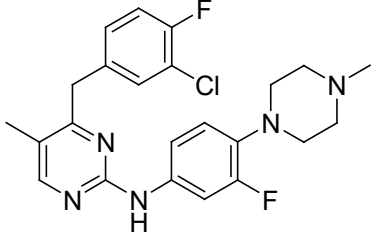
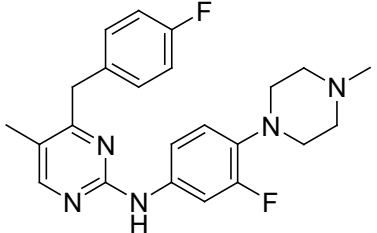
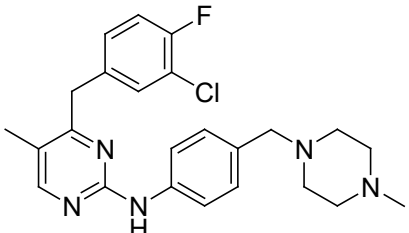
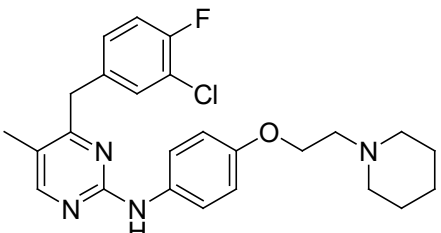
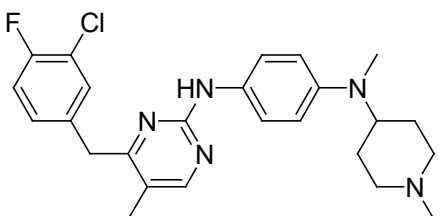
The compounds, which were not incorporated in the training set (7 test set compounds), were used for the calculation of predictability of generated QSAR model. Following equation was used to calculate the value of r_{pred}^2 :

$$r_{\text{pred}}^2 = \frac{(SD - PRESS)}{SD} \quad (2)$$

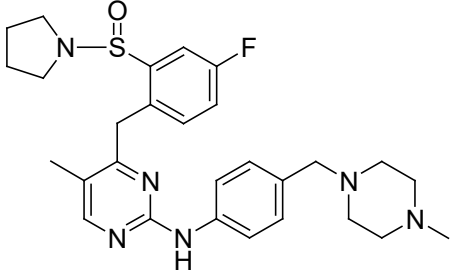
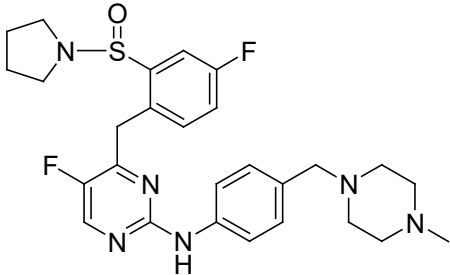
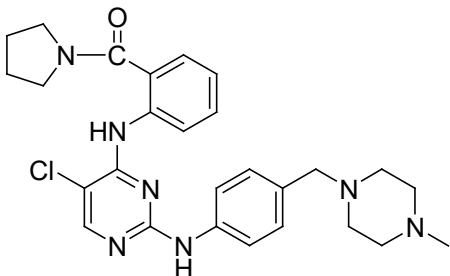
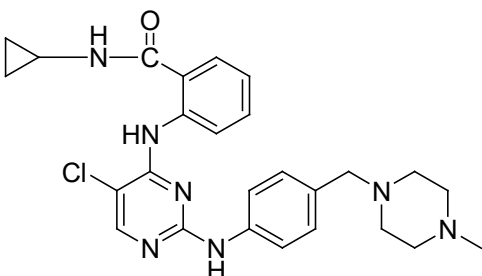
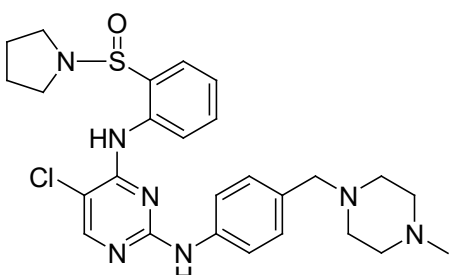
where SD, is the sum of the squared deviations between the biological activity of the test set and mean activity of the training

set compounds, and PRESS is the sum of squared deviations between actual and predicted activity values for each compound in the test set (Makhija *et al.*, 2002).

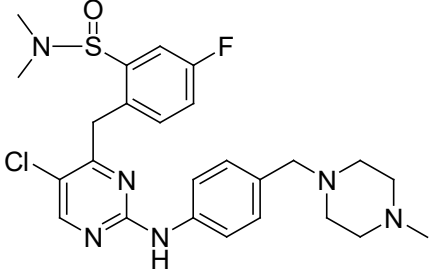
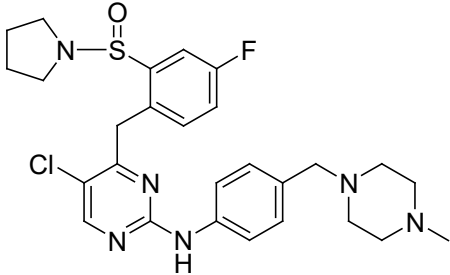
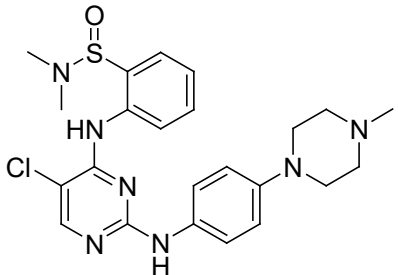
Table 1: Structure of pyrimidine derivatives, experimental activity and predicted inhibitory activity of training set and test set compounds using 3D-QSAR (CoMFA and CoMSIA) method.

Compound No.	Compound	IC ₅₀ ⁱ (μM)	pIC ₅₀ ⁱⁱ	CoMFA		CoMSIA	
				Pred ⁱⁱⁱ	Res ^{iv}	Pred ^v	Res ^{vi}
3#		2.80	5.552	5.623	-0.071	5.627	-0.075
4#		6.10	5.214	5.067	0.147	5.250	-0.036
5		0.73	6.136	6.233	-0.097	6.005	0.131
6		1.63	5.787	5.857	-0.07	5.973	-0.186
7		0.42	6.376	6.407	-0.031	6.589	-0.213

Compound No.	Compound	IC ₅₀ ⁱ (μM)	pIC ₅₀ ⁱⁱ	CoMFA		CoMSIA	
				Pred ⁱⁱⁱ	Res ^{iv}	Pred ^v	Res ^{vi}
8#		0.44	6.356	6.498	-0.142	6.438	-0.082
9		0.29	6.537	6.505	0.032	6.608	-0.071
10		1.40	5.853	5.883	-0.03	5.719	0.134
12#		3.40	5.468	5.372	0.096	5.250	0.218
13		0.027	7.568	7.521	0.047	7.426	0.142
14		0.088	7.055	7.164	-0.109	7.214	-0.159

Compound No.	Compound	IC ₅₀ ⁱ (μM)	pIC ₅₀ ⁱⁱ	CoMFA		CoMSIA	
				Pred ⁱⁱⁱ	Res ^{iv}	Pred ^v	Res ^{vi}
15		2.93	5.533	5.685	-0.152	5.607	-0.074
16		1.15	5.939	5.887	0.052	5.848	0.091
18		0.061	7.214	7.530	-0.316	7.622	-0.408
19#		0.032	8.494	8.653	-0.159	8.560	-0.066
20		0.019	8.721	8.138	0.583	7.779	0.942

Compound No.	Compound	IC ₅₀ ⁱ (μM)	pIC ₅₀ ⁱⁱ	CoMFA		CoMSIA	
				Pred ⁱⁱⁱ	Res ^{iv}	Pred ^v	Res ^{vi}
21		0.240	6.619	6.429	0.19	6.650	-0.031
22#		0.037	7.431	6.707	0.724	6.985	0.446
23		1.08	5.966	6.574	-0.608	6.993	-1.027
24		1.32	5.879	6.396	-0.517	5.939	-0.06
25		3.91	5.407	5.268	0.139	5.275	0.132

Compound No.	Compound	IC ₅₀ ⁱ (μM)	pIC ₅₀ ⁱⁱ	CoMFA		CoMSIA	
				Pred ⁱⁱⁱ	Res ^{iv}	Pred ^v	Res ^{vi}
26		0.082	7.086	7.248	-0.162	7.180	-0.094
27		0.091	7.040	6.984	0.056	7.021	0.019
28#		0.056	7.251	7.058	0.193	6.984	0.267
29		1.73	5.761	5.569	0.192	5.716	0.045

#Test set compounds, ⁱAXL kinase inhibitory activity IC₅₀ (μM), ⁱⁱpIC₅₀ values (-logIC₅₀), ⁱⁱⁱPredicted values based on CoMFA analysis, ^{iv}Residual values based on CoMFA analysis (predicted value- experimental value), ^vPredicted values based on CoMSIA analysis, ^{vi}Residual values based on CoMSIA analysis (predicted value-experimental value).

RESULTS AND DISCUSSION

Results of CoMFA model

CoMFA model was developed using steric and electrostatic fields. As the first step, the partial least square analysis was undertaken using “leave-one-out (LOO)”. The value of q² was found to be 0.700 with 5 optimum numbers of components. The same 5 components along with column filtering set to 2.0 were used to calculate the value of the conventional correlation coefficient (r²_{ncv}). The value of r²_{ncv} was found to be 0.911; the value of F was found to

be 38.856; the value of SEE (Standard Error of Estimate) was found to be 0.316. The steric and electrostatic field values were found to be 2.605 and 3.030 respectively. Table 2 depicts the statistical parameters obtained by development of the CoMFA model. The results of cross-validation coefficient r²_{cv} (0.641) and bootstrap analysis r²_{bs} (0.957) supported the reliability of the developed QSAR model. Experimental and predicted activities of training set compounds and test set compounds are already represented in Table 1. Figure 4 (A, B) depicts the correlation of experimental and predicted activities of training and test set compounds.

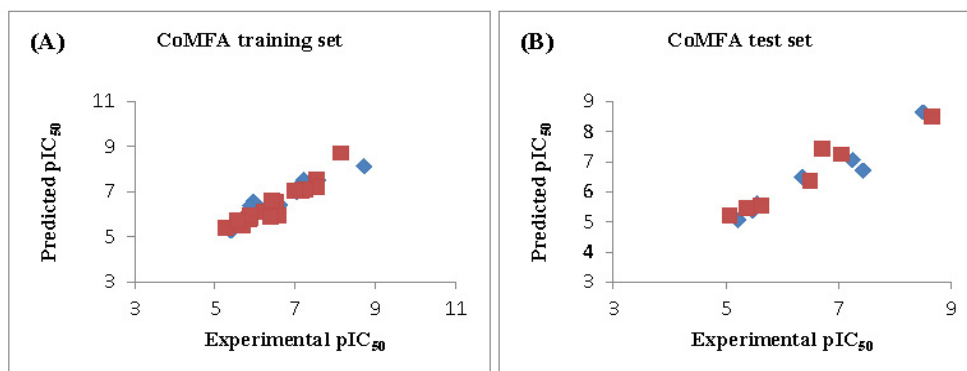


Fig. 4: Plot of experimental and predicted activities of the training and the test set compounds based on CoMFA model.

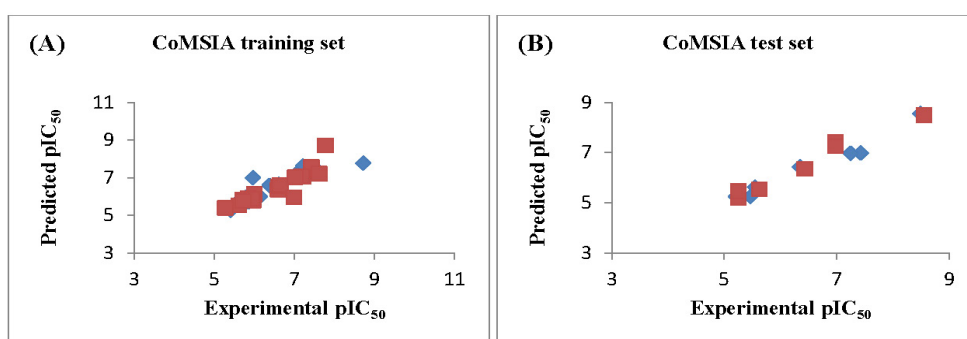


Fig. 5: Plot of experimental and predicted activities of the training and the test set compounds based on CoMSIA model.

Results of CoMSIA model

CoMSIA provides information of steric, electrostatic, hydrophobic, H-bond donor and H-bond acceptor fields. Comparative molecular field similarity indices analysis was undertaken by similar training and test set compounds as that of CoMFA following the successful results of CoMFA. The value of q^2 was found to be 0.622 with 6 optimum numbers of components. To calculate the conventional correlation coefficient, same 6 components and column filtering as 2.0 were used. The value of r^2_{ncv} was found to be 0.875. The contribution of steric, electrostatic, hydrophobic, H-bond donor and H-bond acceptor were found to be 0.666, 1.571, 0.985, 1.015 and 0.884 respectively. Further, bootstrap analysis ($r^2_{bs} = 0.948$) supported the quality of the developed CoMSIA model. To check internal reliability within the dataset, cross-validation by two groups (leave half out method) was undertaken. The r^2_{cv} was found to be 0.595. The values of r^2_{bs} and SEE_{bs} were found to be 0.948 and 0.256 respectively. Statistical parameters based on CoMFA and CoMSIA model were depicted in Table 2. Experimental and predicted activities of training and test set compounds are represented in Table 1. Figure 5(A, B) depicts the correlation of experimental and predicted activities of the training and the test set compounds.

QSAR visualization

3D contour maps are the important features of the comparative molecular field analysis (CoMFA) and comparative molecular similarity indices analysis (CoMSIA). These contour

maps are derived when there are changes in molecular fields. The 3D space contour maps surrounding the compounds were derived for CoMFA as well as CoMSIA model. Based on the contour maps study, changes can be carried out to improve the biological activity and to optimize pyrimidine derivatives as AXL kinase inhibitors. Most active compound 20 was bifurcated into three equal parts to understand the effect of various fields on biological activity, (Figure 6A-C).

Table 2: Statistical parameters based on CoMFA and CoMSIA model.

PLS analysis parameters	CoMFA	CoMSIA
$r^2_{LOO}(q^2)$	0.700	0.622
r^2_{ncv}	0.911	0.875
SEE	0.316	0.384
ONC	5	6
F value	38.865	21.067
Steric field contribution	2.605	0.066
Electrostatic field contribution	3.030	1.571
Hydrophobic field contribution	-	0.985
H-bond donor field contribution	-	1.015
H-bond acceptor field contribution	-	0.884
r^2_{bs}	0.957	0.948
SEE_{bs}	0.200	0.256
r^2_{cv}	0.641	0.595
Test set $r^2(r^2_{pred})$	0.709	0.668

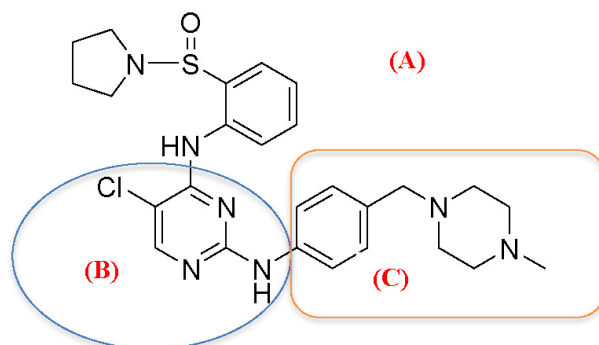


Fig. 6: Compound 20 bifurcated into three parts (A), (B) and (C).

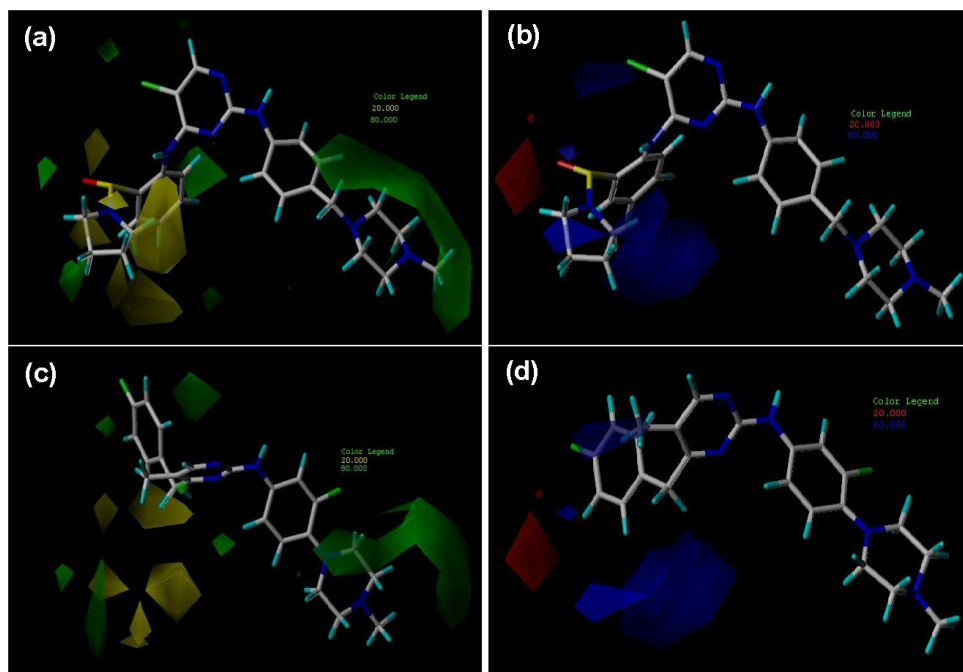


Fig. 7: CoMFA (StDev*Coeff) Contour maps of steric field (a, c) and electrostatic fields (b, d) for compound 20 (a, b) and the compound 4 (c, d) respectively.

CoMFA contour maps

Most active compound 20 ($IC_{50} = 0.019 \mu M$) was selected from the series for the examination of CoMFA contour maps. Contour maps were generated surrounding the molecule. Here, style of contour map was set as transparent for visualization of contours nearby the molecule. Green and yellow contour maps (80% and 20% contributions) represent the steric field. The green area indicates the favorable steric groups, and the inhibitory activity can be improved against AXL kinase receptor by incorporating the bulky groups. While, a yellow contour indicates the non-favorable steric groups for the activity, so the presence of bulky groups near this area can reduce biological activity. The electrostatic field with blue contours depicts positively charged groups. The electrostatic field with red contours depicts negatively charged groups.

Green contour near "C" region, in Figure 7(a), at piperazine ring, suggests that the bulky groups are favorable for the activity. Therefore, N-substituted piperazine ring may increase the biological activity. Green contour near "C" region at the C-4 place of phenyl ring indicates that bulky groups

are favored for inhibitory effect. This is apparent from actual biological activity values of compound 3 ($IC_{50} = 2.80 \mu M$), 4 ($IC_{50} = 6.10 \mu M$) and 29 ($IC_{50} = 1.73 \mu M$). These compounds have 1-methylpiperazine ring directly attached to C-4 atom of phenyl ring as compared to other compounds such as 8 ($IC_{50} = 0.44 \mu M$), 14 ($IC_{50} = 2.80 \mu M$), 18 ($IC_{50} = 0.061 \mu M$), 19 ($IC_{50} = 0.032 \mu M$), and 28 ($IC_{50} = 0.056 \mu M$) in which 1,4-dimethylpiperazine ring was present. This leads to the difference in inhibitory activity values of compounds. This is also evident from the comparison of compound 28's ($IC_{50} = 0.056 \mu M$) and 29's ($IC_{50} = 1.73 \mu M$) activities. Additionally, green contour at phenyl ring (C-5) in "A" region indicates that the bulky groups at this position results in increased inhibition. As seen in Figure 7(a), yellow contour are seen at the C-4 position in phenyl ring at "A" region, which suggests that incorporation of bulky groups are not favored. The bulky groups at C-5 position indicate increased inhibitory effect. This is apparent from the actual biological activity values of compounds 13, 14, 18, 19, 20, 28 in which no substitution was present at the C-5 position so, they have very potent activity,

while those compound with -F as substituent has minimal activity.

The electrostatic field represents blue and red contour maps. Blue contour favors positively charged groups and red contour map favors negatively charged groups. Most potent compound 20 was chosen as reference compound from the series.

The blue contour around phenyl ring at C-4 and C-5 position in "A" region suggests that addition of positively charged substituent results in an increase in the biological effect. (Figure 7(b)) This is apparent from relatively more active un-substituted phenyl ring in compounds 13, 14, 18, 19, 20 as compared with the presence of negatively charged substituent in compounds 15, 16, 21, and 25.

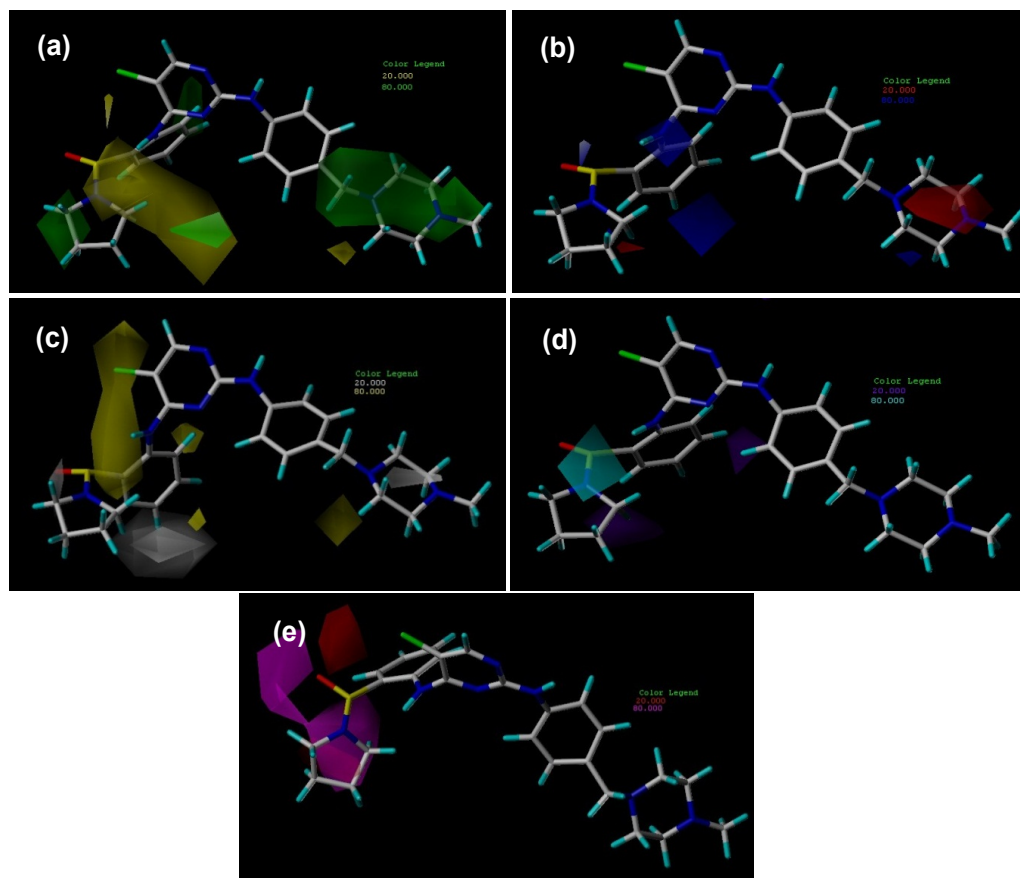


Fig. 8: CoMSIA (stDev*Coeff) contour maps of (a) steric, (b) Electrostatic (c) Hydrophobic (d) Hydrogen bond donor and (e) Hydrogen bond acceptor fields for compound 20.

CoMSIA contour maps

Steric field and electrostatic field contour maps observed in the comparative molecular field similarity indices analysis (CoMSIA) are homogenous to those observed in CoMFA. In CoMSIA steric maps, the yellow contour at the C-2 position in phenyl ring and at pyrrolidine ring in "A" region suggests that incorporation of bulky groups are not favored. This is apparent from the comparison of the biological effect of moderately active compound 21 ($IC_{50} = 0.240 \mu\text{M}$) and compound 26 ($IC_{50} = 0.082 \mu\text{M}$).

As it appears in Figure 8(b), red contour near piperazine ring in the C region, suggests that substituent with negative charge leads to enhancement of the inhibitory effect of compounds. So, an introduction of -F, -Cl or $-\text{CF}_3$ groups may lead to a significant change in the inhibitory activity of compounds.

In Figure 8(c), white contour at the C-4 of phenyl scaffold in "A" region suggests that the hydrophilic substitution in this region results in increased biological activity. This is clear

from the contour maps of the molecule, where bulky substitution such as -F and -Cl are present, leading to a reduction in activity. Further, yellow contours near the pyrimidine ring indicates the hydrophobic substitution, which leads to an increase in activity.

Cyan and purple contour maps represent the CoMSIA donor fields. H-bond donor substituents are favored in the region where the cyan contour map is present. While purple contour suggests that H-bond donor reduces the biological activity. Small cyan contour map in Figure 8(d), was observed at the C-2 position in phenyl ring which suggests that the presence of H-bond donor in this region results in increased biological activity. Acceptor field represents magenta and red contour maps. H-bond acceptor substituents are favored in a region where the magenta contour is present. While H-bond acceptor substituents are not favored where the red contour is present. In Figure 8(e), the pyrrolidine ring in "A" region contains magenta contour map which suggests that the H-bond acceptor are favored for the activity. Important finding of QSAR study are depicted in Figure 9.

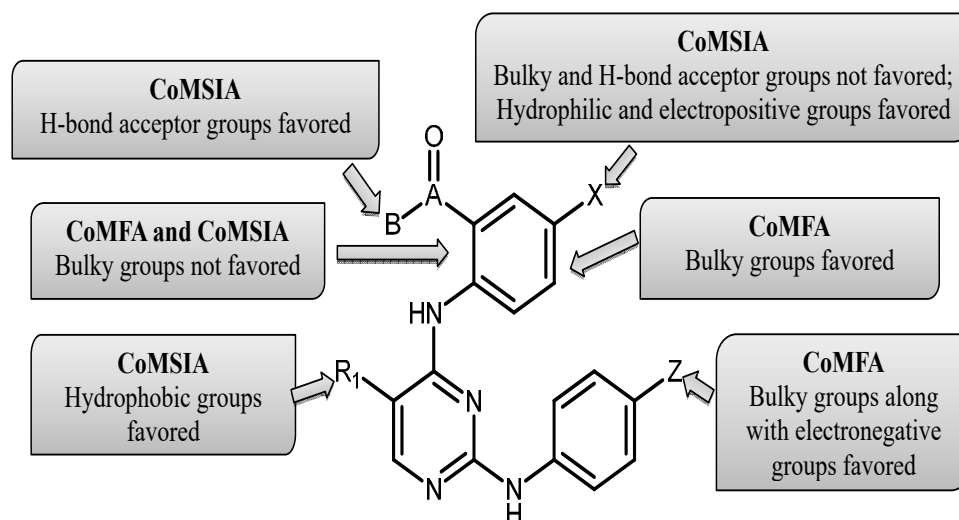


Fig. 9: Important structure finding and SAR from QSAR study.

CONCLUSION

Currently, AXL tyrosine kinase receptor is an emerging target for anti-cancer drug development. We undertook the quantitative structure-activity relationship analysis using CoMFA and CoMSIA to build the correlation between pyrimidine derivatives and their biological activity as AXL kinase inhibitors. To achieve CoMFA and CoMSIA, PLS analysis was undertaken to correlate the descriptor with biological activity. The cross-validation coefficient (q^2) value of 0.700 and 0.622 for CoMFA and CoMSIA model along with all the statistical parameters and generated contour maps strongly reveal that the developed models are satisfactory. Contour maps indicate the important structural features such as incorporation of bulky groups along with negatively charged substituents at “C” region in phenyl ring and the positively charged substituent at “A” region. These features lead to the significant enhancement of the biological activity. CoMFA and CoMSIA analysis strongly suggest that steric, electrostatic and hydrophobic fields play a crucial role in biological activity. 3D-QSAR analysis discussed in this study can be used further for the discovery of novel pyrimidine analogs as AXL kinase inhibitors as anti-cancer agents with a high degree of selectivity and specificity.

ABBREVIATIONS

RTK	Receptor Tyrosine Kinase
QSAR	Quantitative Structure Activity Relationships
NSCLC	Non-Small Cell Lung Cancer
CoMFA	Comparative Molecular Field Analysis
CoMSIA	Comparative Molecular Similarity Indices Analysis
PLS	Partial Least Square Analysis
LOO	Leave One Out
ONC	Optimum Number of Components
GH	Gasteiger Huckel
r^2_{ncv}	Non-cross validation correlation coefficient

r^2_{pred}	Predictive correlation coefficient
q^2	Cross-validation correlation coefficient
SEE	Standard Error of Estimate
F value	Fisher's value

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AUTHOR CONTRIBUTIONS

Both the authors, Siddharth J. Modi and Vithal M. Kulkarni, have approved the final version of the manuscript.

CONFLICT OF INTEREST

The authors declare no conflict of interest pertaining to this manuscript.

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