# Journal of Applied Pharmaceutical Science

JAPS

Journal of Applied
Pharmaceutical Science

Available online at www.japsonline.com

ISSN: 2231-3354 Received on: 20-09-2011 Revised on: 25-09-2011 Accepted on: 30-09-2011

# Biochemical liver function with aqueous fruit extract of *Solanum macrocarpum* linn in albino rats acutely administered triton-x to induce hyperlipidaemia

O. A. Sodipo, F.I. Abdulrahman, U.K. Sandabe and J.A. Akinniyi

#### O.A. Sodipo

Department of Clinical Pharmacology and Therapeutics, College of Medical Sciences, University of Maiduguri, P.M.B. 1069 Maiduguri.

F.I. Abdulrahman, J.A. Akinniyi Department of Chemistry, Faculty of Science, University of Maiduguri, P.M.B. 1069 Maiduguri.

# U.K. Sandabe

Department of Veterinary Physiology, Pharmacology and Biochemistry, Faculty of Veterinary Medicine, University of Maiduguri, P.M.B 1069, Maiduguri.

# **ABSTRACT**

The effect of the aqueous fruit extract, *Solanum macrocarpum* Linn on some biochemical indices of liver function was studied in triton-induced hyperlipidaemic wistar rats. Thirty rats (160-200g) were used in the study and assigned to 5 groups of 6 rats each. Group I hyperlipidaemic control rats received distilled water only, whereas groups II, III, IV and V, the experimental hyperlipidaemic rats, were administered graded doses of the plant extract (25mg/kg, 50mg/kg, 100mg/kg and 200kg/mg) per body weight intraperitoneally after which blood samples were taken from the rats 24hrs, 48hrs and 72hrs, respectively after extract administration. Serum aspartate amino transferase (AST) dose dependently and significantly decreased (P<0.05) at 48hrs and 72hrs. The values of alanine amino transferase (ALT) decreased significantly (P<0.05) at 72hrs when compared to the control. The decrease in alkaline phosphatase (ALP) activity was not significant (P>0.05) when compared to the control. Serum protein and albumin decreased significantly (P<0.05) while bilirubin increased significantly (P<0.05) at 72hrs of study. In conclusion, *Solanum macrocarpum* probably has hepatoprotective effects.

**Key words:** Hyperlipidaemic, *Solanum macrocarpum*, liver function, rats.

#### INTRODUCTION

Natural substances of botanical origin have been used throughout the world for human and animal health care (Enzo, 2006). About half of the world's medicinal compounds are probably derived or obtained from plants (Ahmadu *et al.*, 2006). The use of medicinal plants in West Africa is probably as old as the duration of human settlement (Abdulrahman *et al.*, 2010). *Solanum macrocarpum* ("Gorongo" in Kanuri) is one of the agents used for folklore medicinal purposes. Although the unripe fruit of the plant is used by traditional healers for the treatment of various ailments (Grubben and Denton, 2004), information on the hepatotoxicity of the extract in man and animals is not readily available except that by Sodipo *et al.*, 2009 that investigated the effect of the aqueous fruit extract of the plant on the liver function of diet-induced hypercholesterolaemic rats. The present study investigated the effect of the fruit of *S. macrocarpum* on triton-induced hyperlipidaemic rats in an attempt to find an alternative hypolipademic agent that is both therapeutically and cost effective, but with fewer side effects than the existing ones which are expensive and at the same time have numerous side effects (Hardman and Limbird, 2001).

For Correspondence: Mrs. O.A. Sodipo. Ph. D.

Department of Clinical Pharmacology and Therapeutics, College of Medical Sciences, University of Maiduguri, P.M.B. 1069 Maiduguri. Telephone: +234(0)8034107098

#### MATERIALS AND METHOD

# Plant collection and identification

The plant material (*Solanum macrocarpum* Linn.) used in this study was obtained from Alau in Konduga Local Government, Borno State, Nigeria, between October and November, 2007. The plant was identified and authenticated by Prof. S.S. Sanusi of the Department of Biological Sciences, University of Maiduguri, Maiduguri, Nigeria. Specimen voucher No. 548 was deposited at the Research Laboratory of the Department of Chemistry, University of Maiduguri.

#### **Extraction**

The fruit of *S. macrocarpum* with the calyx removed was air dried and pulverized by grinding using pestle and mortar. The 2.2kg of the ground fruit was subjected to exhaustive Soxhlet-extraction in distilled water at 100°C to give the extract yield of 15.3% <sup>w</sup>/<sub>w</sub> (Mittal *et al.*, 1981, Fernando *et al.*, 1991; Lin *et al.*, 1999). The resultant solution was concentrated *in vacuo* and it was stored in a specimen bottle at room temperature until when required.

#### **Animals**

Thirty male albino rats of Wistar strain weighing 160-200g were used in this study. The animals were obtained from the Animal House Unit of the Department of Veterinary Physiology and Pharmacology, University of Maiduguri. The animals were housed under standard laboratory condition in plastic cages. They were fed commercial growers' mash feed (ECWA Feeds, Jos, Nigeria) and water was provided *ad libitum*. All the animals were handled according to the International Guiding Principles for Biomedical Research Involving Animals (CIOMS, 1985) as certified by the Animal Ethics Committee of the Faculty of Veterinary Medicine, University of Maiduguri.

# Administration of triton and extract

Thirty (30) albino rats were made hyperlipidaemic by feeding them orally (p.o) for 1 week with normal feed diet and triton-X (Sigma Chemical Co. St. Louis, M.O. USA) at a dose of 400mg/kg in saline suspension from the stock concentration of 535g/ml. The rats were divided into 5 groups of 6 animals each. After seven (7) days, the rats were administered with graded doses of the fruit extract. Group I was the control and it was given distilled water only. Groups II, III, IV and V were administered with geometrical doses (25mg/kg, 50mg/kg, 100 mg/kg and 200mg/kg) of the fruit extract intraperitoneally (i.p.) from a stock concentration of 200mg/ml. After 24hrs, 48hrs, and 72hrs, respectively of the effect of the extract on the hyperlipidaemic rats, (adapted from Williamson, et al., 1996), two rats from each group were humanely sacrificed by cutting the throat with a sterile blade and blood was collected from the vena cava into clean, labelled centrifuge tubes without an anticoagulant. The blood was centrifuged at a rate of 12,000 revolutions per minute (rpm) for 10

minutes. The clear, yellow serum was then separated from settled cellular elements. Before the rats were fed with triton-X, their weight was taken. The weights were taken before and after administration of triton-X for 7 days.

# **Biochemical liver function tests**

The liver function parameters estimated from the serum were protein, albumin, total bilirubin and liver enzymes which included aspartate amino transferase (AST), alanine amino transferase (ALT) and alkaline phosphatase (ALP). AST and ALT were assayed using commercial Randox kits (UK) and by Quinica Clinical Applicanda, JA Kits (Moss *et al.*, 1986). The total protein in the serum was estimated using direct Biuret method (Peters *et al.*, 1982; Afonja, 1997). Serum albumin and bilirubin were determined by the dye bromocresol-green method by Doumas *et al.*, (1971); Spencer and Price (1977); Teitz (1994).

# Statistical analysis

Data ware expressed as the mean  $\pm$  SD. The results obtained were subjected to Analysis of Variance (ANOVA) using Graph Pad Software (1998).

# **RESULTS**

# Effect of extract on body weight of rats

The effect of triton-X on mean body weight of albino rats is shown in Table 1. The increase in body weight observed in the rats was not statistically significant (P>0.05) when compared with the zero in all the groups except in group 1 where the increase was significant (P<0.05).

**Table 1:** Change in body weight of male albino rats after being administered Triton-X (400 mg/kg) orally for 7 days.

Group	Mean Body Weight ± S.D. (g)			
	Days of Treatment			
	0	7		
One Two	$114.33 \pm 11.76^{a}$ $98.83 \pm 11.55^{a}$	$129.00 \pm 9.22^b \\ 111.83 \pm 11.18^a$		
Three	$140.83 \pm 37.57^{a}$	$151.67 \pm 36.82^a$		
Four	$137.33 \pm 30.89^a$	$147.00 \pm 30.44^{a}$		
Five	$175.33 \pm 31.10^{a}$	$194.83 \pm 37.12^{a}$		

Within rows, means with the same superscript are not statistically significant (p > 0.05) when compared with day 0 using student t-test.

n = 6 rats

# Effect of extract on liver function

The effect of the aqueous fruit extract of *Solanum macrocarpum* on protein, albumin and total bilirubin are shown in Table 2 whilst those of the liver enzymes are shown in Table 3. Serum protein and albumin decreased significantly (P<0.05) whilst bilirubin increased significantly (P<0.05) at 72hrs of study with increase in extract dose when compared to the control. Aspartate amino transferase (AST) dose-dependently and significantly decreased (P<0.05) at 48hrs and 72hrs. The values of alanine

<sup>0</sup> day = Before triton-X administration

<sup>7</sup> days = After oral administration of triton-X

amino transferase (ALT) decreased significantly (P<0.05) at 72hrs when compared to the control. The decrease in alkaline phosphatase (ALP) activity was not significant (P>0.05) when compared to the the control.

**Table 2:** Effect of the aqueous fruit extract of *S. macrocarpum* on protein albumin and total bilirubin of hyperlipidaemic rats administered orally with Triton-X for 7 days.

Hours after extract administration	Extract dose mg/kg	Protein (g/L)	Albumin (g/L)	Total Bilirubin (µmol/L)
	Control	$71.00 \pm 1.41^{a}$	$39.00 \pm 1.41^{a}$	$2.50 \pm 0.71^{a}$
	25.00	$65.50 \pm 5.00^{a}$	$37.5 \pm 0.71^{a}$	$2.50 \pm 0.71^{a}$
24	50.00	$65.00 \pm 4.24^{a}$	$37.00 \pm 1.41^{a}$	$3.50 \pm 0.71^{a}$
	100.00	$64.50 \pm 0.71^{a}$	$36.00 \pm 0.00^{a}$	$4.00 \pm 0.00^{a}$
	200.00	$64.50 \pm 5.00^{a}$	$35.50 \pm 0.71^{a}$	$4.00 \pm 1.41^{a}$
	Control	$72.00 \pm 2.83^{a}$	$39.00 \pm 1.41^{a}$	$3.50 \pm 0.71^{a}$
48	25.00	$69.50 \pm 0.71^{a}$	$37.50 \pm 0.71^{a}$	$4.00\pm0.50^a$
	50.00	$68.50 \pm 2.12^{a}$	$37.00 \pm 1.41^{a}$	$4.00 \pm 1.41^{a}$
	100.00	$66.50 \pm 2.12^{a}$	$36.50 \pm 0.71^{a}$	$4.50 \pm 0.71^{a}$
	200.00	$66.00 \pm 1.41^{a}$	$35.50 \pm 0.71^{a}$	$5.00 \pm 0.71^{a}$
	Control	67.00 ± 1.41 <sup>a</sup>	$36.50 \pm 0.71^{a}$	$2.50 \pm 0.71^{a}$
72	25.00	$64.00 \pm 2.83^{b}$	$34.00 \pm 1.41^{b}$	$3.50 \pm 0.71^{b}$
	50.00	$61.50 \pm 0.71^{b}$	$33.51 \pm 0.71^{b}$	$4.00 \pm 1.41^{b}$
	100.00	$60.50 \pm 0.71^{b}$	$33.50 \pm 0.71^{b}$	$4.50 \pm 0.71^{b}$
	200.00	$59.50 \pm 0.71^{b}$	$32.50 \pm 0.71^{b}$	$6.50 \pm 0.71^{b}$

Means with different superscripts are statistically significant (p < 0.05) among the groups. Control = Distilled water.

**Table 3:** Effect of the aqueous fruit extract of *S. macrocarpum* on serum enzymes of hyperlipidaemic rats administered orally with Triton-X for 7 days.

Hours after	Extract	Serum enzymes (U/L)				
extract	dose	AST	ALT	ALP		
administration	mg/kg	M CD				
	C 1	00.50 - 0.718	Mean ± S.I			
	Control	$89.50 \pm 0.71^{a}$	$32.00 \pm 9.90^{a}$	$174.00 \pm 15.56^{a}$		
	25.00	$89.00 \pm 0.00^{a}$	$31.50 \pm 3.54^{a}$	$171.50 \pm 6.36^{a}$		
24	50.00	$78.00 \pm 5.56^a$	$27.00 \pm 2.83^{a}$	$170.50 \pm 6.36^a$		
	100.00	59.50 ±	$23.00 \pm 2.83^{a}$	$167.50 \pm 16.26^{a}$		
	200.00	$10.61^{a}$ $58.50 \pm 0.71^{a}$	$21.00 \pm 0.00^{a}$	$131.00 \pm 15.56^{a}$		
48	Control	$59.00 \pm 0.00^{a}$	$36.50 \pm 3.54^a$	$144.50 \pm 0.71^{a}$		
	25.00	$52.50 \pm 0.71^{b}$	$36.50 \pm 3.54^a$	$143.00 \pm 2.83^a$		
	50.00	$38.50 \pm 3.54^{b}$	$34.00 \pm 0.71^{a}$	$141.00 \pm 1.41^{a}$		
	100.00	$36.00 \pm 7.07^{b}$	$25.00 \pm 0.00^a$	$137.00 \pm 7.07^{a}$		
	200.00	$31.50 \pm 0.71^b$	$25.00 \pm 0.00^a$	$126.50 \pm 8.49^a$		
72	Control	$71.50 \pm 6.36^{a}$	$45.50 \pm 3.54^{a}$	110.00 ± 2.83°		
	25.00	$67.00 \pm 0.00^{b}$	$43.50 \pm 0.71^{b}$	$108.00 \pm 2.83^{a}$		
	50.00	$52.50 \pm 0.71^b$	$41.00 \pm 2.83^b$	$105.00 \pm 0.71^{a}$		
	100.00	$44.00 \pm 4.24^b$	$39.00 \pm 0.00^b$	$104.50 \pm 0.71^a$		
	200.00	$44.00 \pm 4.24^b$	$31.50 \pm 3.64^b$	$102.50 \pm 0.71^a$		

Means with different superscripts are statistically significant (p < 0.05) among the groups. Control = Distilled water

# DISCUSSION

The observed decrease in serum protein which was significant at 72hrs (P<0.05) in the present study may be associated with liver damage, nutritional deficiency and renal failure (Mukherjee, 1980; Odutola, 1992; Sood, 2006). Since proteins are constituents of muscle, enzymes, hormones and several other key

factors, invariably these factors will be affected. Thus, the effect of the extract on the hyperlipidaemic rats is probably that of liver toxicity or excessive protein catabolism. The reduction in total protein in this study agrees with the reports of Rabo et al., (2003) working on Butyrospermum paradoxum extracts in rats; Iweala and Okeke (2005) on the aqueous leaf, flowers and tender stem extract of Catharanthus roseus Linn. (Madagascar periwinkle-Apocynaceae) on alloxan induced diabetic rats and Sodipo et al., (2009) on the aqueous fruit extract of diet-induced hypercholesterolaemic rats. This decrease in serum protein was attributed to increased binding of the plant components to serum albumin. The decease in total protein probably portrays hepatocellular damage. The decrease in albumin level which was significant (P<0.05) at 72hrs with increase in extract dose on the hyperlipidaemic rats probably potrays liver damage at this contact time. The observed decrease in albumin levels is in conformity with the results of Uhegbu and Ogbuechi (2004) who reported a decrease in albumin levels. This decrease in albumin in this study contrasts the report by Atangwho et al., (2007) that the more protected the heaptocytes become, the more the boost to their synthetic function.

The increase in bilirubin levels which was significant (P<0.05) at 72hrs, was caused by increasing doses of the extract. Increase in bilirubin values may be cuased by liver damage, excessive haemolytic destruction of erythrocytes, obstruction of the biliary tract (obstructive jaundice) and in drug-induced reactions (Mukherjee, 1988, Odutola, 1992; Sood, 2006). However, if the AST and ALT values are normal, the diagnosis of hepatocellular damage cannot be confirmed (Odutola, 1992). In the present study, the effect of the extract on AST and ALT was that of reduction, significant for AST at 48hrs and 72hrs and for ALT at 72hrs of study respectively, thus confirming the extract's protective ability on the liver cells. Thus, the aqueous fruit extract of *Solanum macrocarpum* under the condition of the present study was probably not toxic just like in the hypercholestolaemic rats (Sodipo *et al.*, 2009).

The result of the liver enzymes showed that the extract had a significant decrease (P<0.05) on AST of the hyperlipidaemic rats at 48hrs and 72hrs and the decrease in ALT was significant (P<0.05) at 72hrs. However, the decrease in ALP for all the times of study was not significant (P>0.05). This observation is in line with the recent findings of Kim et al., (2006); Atangwho et al., (2007) who reported a decrease in elevated liver enzymes upon treatment of alloxan-induced diabetic rats with ethanol leaf extract of Veronia anygdalina Del. The value of the liver function tests depends on the specificity for damage as well as their sensitivity (Cutler, 1974; Okonkwo et al., 1997; Sodipo et al., 2009). Although serum levels of both AST and ALT become elevated when disease processes affect the liver integrity, ALT is the more liver specific enzyme and therefore generally more sensitive to changes in activity levels than AST (Kachmar and Moss, 1976; Sodipo et al., 2009).

The results of the present study in which ALT was significantly reduced at 72hrs and AST was significantly reduced

at 48hrs and 72hrs of study respectively, therefore, suggest that the extract had no significant influence on the liver function. Also, AST is highly concentrated in several tissues including the heart, muscle, liver, skeletal muscle and kidney while ALT has its highest concentration in the liver (Kaneko and Cornelius, 1971; Wilkinson, 1976; Okonkwo *et al.*, 1997; Nduka, 1997; Mayne, 1998; Atangwho *et al.*, 2007; Sodipo *et al.*, 2009), therefore, measure of ALT in serum is of greater diagnostic specificity in confirming or excluding liver damage. Since the decrease in ALT in the present study was significant at 72hrs, then there is no likelihood of liver damage by the aqueous fruit extract of *Solanum macrocarpum*.

A non-statistically significant (P>0.05) decrease in ALP value as obtained on extract administration in the present study is not of much clinical significance (Atangwho *et al.*,2007, Sodipo *et al.*, 2009). Even if there had been an elevation in the ALP upon extract administration, it could still not have confirmed liver damage because according to Odutola (1992), ALP and AST originate from different tissues such as the liver, bones, intestine and placenta.

In the present study, upon extract administration, both the liver enzymes AST and ALT decreased significantly whilst ALP did not show any significant change. All these may show that the effect of the extract on the rats was not that of toxicity. Therefore, the aqueous fruit extract *Solanum macrocarpum* appears to be safe.

# CONCLUSION

The present study shows that the aqueous fruit extract of *Solanum macrocarpum* may probably have a hepatoprotective effect. However, the results of the histopathological studies on hepatic architecture are required in order to confirm the findings of the biochemical indices of liver function.

# ACKNOWLEDGEMENTS

The authors gratefully acknowledge the technical assistance of Rebecca Gali (Mrs), Bitrus Wampana and Fine Akawo.

# REFERENCES

Abdulrahman FI, Akan JC, Sodipo OA and Onyeyili PA. Effect of aqeous root bark extract of *Vitex doniana* sweet on haematological parameters in rats. *J. Am. Sci.* 2010. **6** (12): 8-12.

Afonja OA. Basic Clinical Biochemistry Practice Manual; 85. 1997.

Ahmadu AA, Akpulu IN, Hassan HS, Sule MJ, Pateh UU. Preliminary phytochemical screening and antibacterial activity of *Cirsampelos mucronata* A. Rich. (Menispermacea) extracts. *J. Pharm. Biores* 2006. **3** (2): 103-106.

Atangwho IJ, Ebona PE, Egbung GE, Eteng MUand Eyong EU. Effect of *Veronia amygdalina* Del. on liver function in alloxan-induced hyperglycaemic rats. *J. Pharmacy Biores.* 2007. **4** (1): 25-31.

CIOMS. Council for International Organizations of Medical Sessions. International Guiding Principles for Biomedical Research Involving Animals. WHO 1211, Geneva 27, Switzerland. 1985.

Cutler MG. The sensitivity of function tests in detecting liver damage in the rat. *Toxicol. Appl. Pharmacol.* 1974. **28**: 349-357.

Doumas BT, Watson WA and Biggs HG. Albumin standards and the measurement of scrum albumin with bromocresol green. *Clin, Chem. Acta.* 1971. **3**:87-96.

Enzo AP. Phytochemicals from traditional medicinal plants used in the treatment of diarrheoa. Modes of action and effects on intestinal function. *Phytotherapy Res.* 2000. **20**:717-724.

Fernando MR, Wickramanshingbe SMD, Nalinle I, Thabrew MI, Ariyanando PL and Karunanyake EK. Effects of *Artocarpus heterophyllus* and *Asteracanthus longifolia* on glucose tolerance in normal human subjects and in maturity-onset diabetic patients. *J. Ethnopharmacol.* 1991. **31**:277-283.

Graph Pad Software. Graph Pad Software, Inc., San Diego, California U.S.A. www.graphpad.com. 1998.

Grubben GJH and Denton OA PROTA 2. *Plant Resources of Tropical Africa 2. Vegetables.* Ponen and Looijen hv, Wagening en, Netherlands. (2004) 484-487.

Hardman JG, Limbird LE. *Goodman and Gilman's The pharmacological Basis of Therapeutics* 10<sup>th</sup> ed. McGraw Hill Co. U.S.A. (2001) 971-1001.

Iweala EEJ and Okeke CU. Comparative study of the hypoglycaemic and biochemical effect of *Catharanthus roseus* Linn; family Apocynaceae (Madagascar Periwinkle) and chlorpropamide (diabenese) on alloxan-induced diabetic rats. *Biokemistri*: 2005. **17** (2):149-156.

Kachmar JF and Moss DW. Enzymes (Transaminases) In: Fundamental of Clinical Chemistry W. E. Nobert and N.W. Teitz, eds, W.B. (1976).

Kaneko JJ and Cornelius CE. Clinical Biochemistry of Domestic Animals 2<sup>nd</sup> ed. Academic Press New York, U.S.A.; 20-25.

Kim SJ, Ju, JB, Choi CW and Kim MC. Hypoglycaemic and antihyperglycaemic effect of four Korean medicinal plants in alloxan-induced diabetic rats. *Ann. J. Biochem. Biotech.* 2006 **2** (4):154-160.

Lin J, Opuku AR, Geheeb-Keller M, Hutchings AD, Terblanche SE, Jager AK, Van-Standen J. Preliminary screening of some traditional Zulu medicinal plants for anti-inflamatory and antibacterial activities. *J. Ethnopharmacol.*, 1999. **68**: 267-274.

Mayne, PD. Clinical Chemistry, Diagnosis and Treatment 6<sup>th</sup> ed. London: Arnold International; 119-204. (1998).

Mitall GC, Aguwa CN, Ezeinu BU and Akubue PI: Preliminary pharmacological studies on antivenom action of *Diodia scandens* leaves. *Nig. J. Pharm* 1981. **12**:432-436.

Moss DW, Henderson RA, and Kacher JF. Enzymes In: Textbook of Clinical Chemistry. N.W. Tietz, ed. W.B. Saunders, Philadelphia, U.S.A. (1986) 674-678; 708-709.

Mukherjee KL. Medical Laboratory Technology: A Procedure Manual for Routine Diagnostic Tets. Vol. III. Tata McGraw Hill Pub. Co. Ltd, New Delhi. (1988) 1,282.

Nduka N. Clinical Biochemistry for Students of Chemical Pathology, 1<sup>st</sup> Ed. Longman Lagos: Nigeria Plc. (1997) 122-123.

Odutola A.A. *Rapid Interpretation of Routine Clinical Laboratory Tests* S. Asekome and Company, Zaria, Nigeria. (1992) 112..

Okonkwo CA, Iyaniwura TT and Ukoha AO. Biochemical liver function test with cypermethrin in rats *West Afr. J. Pharmacol. Drug Res.* 1997. **13** (182):18-22.

Peters TT Biamonte GT and Doumas BT. Protein (total protein) in serum, urine and cerebrospinal fluid, albumin in serum. In: Selected Methods of Clinical Chemistry, W.R. Faulkner and S. Meiters, eds, Vol. 9 American Association for Clinical Chemistry, Washington D.C. U.S.A. (1982) 30-35.

Rabo JS, Onyeyili PA and Khalil MI. Chronic toxicity of aqueous extract of *Butryospernum paradoxum* stem bark in rabbit. Haematology and ezymology. *Stud. Res. Vet. Med. Bucharest.* 2003. **1:**37-46.

Sodipo OA, Abdulrahman FI, Sandabe UK and Akinniyi FI. Effect of *Solanum macrocarpum* Linn. on biochemical liver function in diet-induced hypercholesterolaemic rats. *Nig. Vet. J.* 2009. **30** (1): 1-8.

Sood, R. Textbook of Medical Laboratory Technology. 1st ed Jaypee Brothers Medical Publishers (p) New Delhi, India, (2006) 609-672.

Spencer K and Price CP. The determination of serum albumin using bromocresol green. In: Practical Clinical Biochemistry. Vol. II 5<sup>th</sup> ed. Heinemann Medical Books Ltd, London. (1977) 553-554.

Tietz, NW. Fundamentals of Clinical Chemistry with Clinical Correlation. Bailliere Tindall, London. (2006) 2334.

Uhegbu FO and Ogbuechi KU. Effect of aqueous extract (crude) of leaves of *Veronia amygdalina* Del. on blood glucose, serum albumin and cholesterol levels in diabetic albino rats. *Global J. Pure and Applied Sci.* 2004. **10** (10):189-194.

Wilkinson JH. The principles and practice of Diagnostic Enzymology. Edward Arnold Press, London, UK; 87-95-129-138; 303. (1976).

Williamson EM, Okpako DT and Evans FI. *Pharmacological Methods in Phytotherapy Research*. Vol. 1, Selection, Preparation and *Pharmacological Evaluation of Plant Material*. Wiley and Sons, England. (1996) 228