

# Effect of estradiol replacement in ovariectomized NMRI mice in response to acute and chronic stress

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## ABSTRACT

The effect of intraperitoneal (IP) and intraventricular (ICV) injection of 17- $\beta$ -estradiol on hormonal and metabolic changes induced by acute and chronic stress in ovariectomized female NMRI mice was evaluated. NMRI female mice ( $30 \pm 5$  g) were used. Animals' ovaries were surgically removed and one cannula was inserted into their skull under deep anesthesia. One week later, animals were received 17- $\beta$ -estradiol either IP (0.05, 0.01, 0.1 mg/kg) 30 min before or ICV (0.05, 0.01, 0.1  $\mu$ g/mouse) 5 min before the stress. Results showed that acute and chronic stress, increased plasma cortisol concentration in ovariectomized mice, but IP and ICV injection of 17- $\beta$ -estradiol after stress induction reduced plasma cortisol concentration. Stress decreased delay to eating time, food and water intake and weight of ovariectomized animal significantly decreased. ICV and IP administration of estradiol in acute stress reduced delay to eating time significantly, but did not any change in water intake. In the acute stress IP administration of estradiol significantly, decreased food intake. In chronic stress both intraperitoneal and intraventricular administration of estradiol caused significant reduction in food intake in ovariectomized mice. Moreover, Stress reduces the brain to the adrenal volume ratio, which, IP and ICV administration of estradiol could inhibit this effect. It can be concluded that 17- $\beta$ -estradiol has different effects when administered IP or ICV on stress-induced metabolic changes in the ovariectomized mice.

## INTRODUCTION

Stress is a response to anything that causes a disruption of the internal balance (homeostasis) and commonly relate to experiences that cause feelings of frustration and anxiety (McEwen, 2006; Rezaei *et al.*, 2016). Stress responses include changes in behavior, autonomic function, hormone secretion, metabolic and tissue changes and also changes in protein expression levels (McEwen & Gianaros, 2011). If the stress is chronic and uncontrollable, it causes emotional disorders, anxiety and depression (Mah *et al.*, 2016). Repeated and chronic stress can lead to hypothalamic-pituitary-adrenal (HPA) axis dysfunction (Asalgoo *et al.*, 2015; Szabo *et al.*, 2012). One of the

the important effects of stress is changes in appetite and nutritional systems (Nishiyama *et al.*, 2008). Metabolic disorders which are one of the most important consequences of social life in the last century are caused by stress (Erfani *et al.*, 2016; Tamashiro *et al.*, 2011). Also, a lot of diseases such as diabetes, cardiovascular diseases, chronic fatigue, depression, anxiety and autoimmune diseases are directly related to stressful life (McEwen, 2006; Hatef *et al.*, 2015). There are gender differences in the incidence and onset of stress-related disorders and other mental disorders that these differences can be attributed to the effects of sex hormones ((McEwen, 2007; Verma *et al.*, 2011; Herzog *et al.*, 2009). Animal models for psychiatric disorders are mainly based on male animals, which the main reason for not using female animal is their estrous cycle and the variation of sex hormones per phase. Female sex hormones such as progesterone and estrogen affect in emotion and cognition behaviors which this leads to sex differences in behavior (ter Horst *et al.*, 2012).

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On the other hand, women respond differently to stress than men, which might be related to the phases of the estrous cycle (ter Horst *et al.*, 2012; Bowman *et al.*, 2009; Chalabi-Yani *et al.*, 2015). In animals, it has been shown that stress-induced corticosterone and adrenocorticotropin levels are higher in the female than in males (Kudielka and Kirschbaum, 2005). Memory and cognition functions are mediated by the hippocampus (Eslamizade *et al.*, 2015; Meftahi *et al.*, 2014; Meftahi *et al.*, 2015). It is well accepted that chronic stress produces deficiency in hippocampal-dependent memory in male rats, but increases it in females (Bowman, 2005; Luine, 2007). Also, Galea *et al.* (1997) showed that apical dendritic atrophy in CA3 pyramidal neurons is seen in male, but not female brains in stress. If estrogen signaling in the brains of females was blocked, stress displayed destructive effects on them and when estrogen signaling was activated in males, the destructive effects of stress were inhibited. Also, aromatase enzyme (which is responsible for producing estrogen) levels, which producing estrogen is significantly higher in the prefrontal cortex of female rats that causes resilience against stress (Wei *et al.*, 2014).

Thus the relationship between HPA and behavioral responses to stress are not the same in male and female animals and explain that there are sex differences in the pathways regulating the stress response. Despite the exact mechanisms are unknown, estradiol plays a crucial role in generating these sex differences effects of stress on central processes.

Although studies investigated the effects of estrogen during stress, but, there is a lack of studies on the effect of estrogen on metabolic changes during stress. Therefore, the aim of this study was to evaluate the protective effect of intraperitoneal and intraventricular injection of 17 beta-estradiol on plasma cortisol concentration and metabolic changes such as delay to eating (anorexia), water and food intake, weight change, an additional aim was to clarify changes in the ratio of brain volume to adrenal gland volume after acute and chronic stress in ovariectomized female NMRI mice.

## MATERIALS AND METHODS

### Animals

Female NMRI mice ( $30 \pm 5$ g, 6 mice for each experiment) were used throughout the study.

The animals were housed in groups of six per cage under a 12 h/12 h light/cycle and controlled temperature 24-22 °C, with *ad libitum* food and water available. The animals were housed in the standard animal room for two weeks before starting the study to adaptation. Amount of food and water in each cage were calculated as water and food intake.

Vaginal Smear was taken from all the animals and their sexual cycle was examined before tests and all animals were in a proestrus phase. All Experiments were done in accordance with standard ethical guidelines and approved by the local ethical committee (The Baqiyatallah University of Medical Committee on the Use and Care, 81/021, July 10, 2014).

### Drugs

The following drugs were used throughout the experiments: estradiol (Abu-Rayhan- Iran), ketamine hydrochloride and diazepam (Sigma, St. Louis, MO, USA). Estradiol was dissolved in oil and administered in intraventricular (0.1, 0.1, 0.05 µg/Mouse) five minutes and intraperitoneal (0.1, 0.1, 0.05 mg/kg) 30 minutes before the stress induction. Ketamine hydrochloride (Sigma, US) and diazepam hydrochloride (Sigma, US) were dissolved in sterile saline. The control group received oil.

### Experimental procedure

Ovariectomy surgical procedure was performed according to the procedure described by Waynforth (Waynforth, 1980). Anesthetizing was performed by intraperitoneal injection of ketamine hydrochloride (50-75 mg / kg) and diazepam (5-7 mg / kg). In order to ovariectomy, a small middle dorsal incision (2-1 cm) was made to the 13th ribs, fallopian tubes along with blood vessels were closed and both ovaries were removed (bilateral ovariectomy). Finally, the incision was sutured. In order to sterilize the sides, betadine solution and penicillin powder were used. In the control group, surgical procedures were performed like ovariectomy group, but the ovaries were not removed. After surgery, the animals were housed individually in cages for a period of one week to allow recovery.

### Cannulation

First, the animals were anesthetized by Ketamine hydrochloride (75-50 mg / kg) and diazepam (7-5 mg / kg). Then, a steel guide cannula was implanted in the animal's head by stereotaxic device according to Paxinos and Franklin atlas Coordinates (Paxinos and Franklin, 2001) for the lateral ventricles, (AP = + 0.9 mm, ML =  $\pm$  2 mm DV = 3 mm) and fixed in place using a lens screw and dental cement. Then, a thin wire with same size as the steel guide cannula was placed inside it to prevent the closure of guide cannula.

### Stress induction

The animals were randomly allocated to experimental and control groups. In the experimental group animals were ovariectomized and were divided into two acute and chronic stress groups. Estradiol was injected intraperitoneal and intraventricular (one-side; left) to the experimental group. Estradiol injected intraperitoneal (0.01, 0.1, 0.5 mg/kg) 30 minutes before induction of chronic stress and injected intraventricular (0.01, 0.1, 0.5 µg / rat), five minutes before the induction of acute stress and then, to stress induction the animals were placed into a Communication Box. The device consists of nine separate sections (50 × 16 × 16 cm) made of plexiglass with tiny holes that allow olfactory and auditory communication. Steel bars (with a 4 mm diameter) are placed on the floor of the instrument at 1.3 cm distance apart, through which electric shock is transmitted to the animal's soles.

The duration and intensity of the induced shock were controlled by a computer connected to the communication box (60

mV, 10Hz, for 60 s). The electric shock was induced randomly between the 9-13 hours. In order to adapt animals to the environment, they were transferred to the test room 60 minutes before the induction of the stress, and remained there 30 minutes before and 30 minutes after the induction of the stress. The control animals were placed in the device for 60 minutes without receiving any shock.

In chronic stress electric shock induction continued for seven consecutive days and in acute stress electric shock was induced one day. After the stress, the animals were returned to the cage.

After completing the tests, animals were anesthetized with high doses of ketamine, then their brain and adrenal glands were removed and kept in 4% formalin solution for fixation. Sixty days later, brain and adrenal glands were removed from formalin. The weight and volume of the brain and adrenal glands were measured by mercury immersion. Weight, food and water intake, delay to eating (the elapsed time between mice replacement in the home cage and beginning food intake was calculated as a delay to eating or anorexia) and plasma cortisol levels were measured as metabolic parameters in both stress and control groups.

#### Measuring the concentration of cortisol hormone

One day before the experiment and seven days following the induction of the stress, blood samples were taken from all the animals through their retro-orbital sinus (0.5 ml blood in 0.5 ml of EDTA 3%) and were centrifuged at 3000 rpm for five minutes at 4°C. Then, the animals' supernatant plasma was collected for

measuring cortisol levels using a cortisol measurement kit (Cortisol ELISA kit; 4164; DRG Instruments GmbH, Germany) at 450 nm.

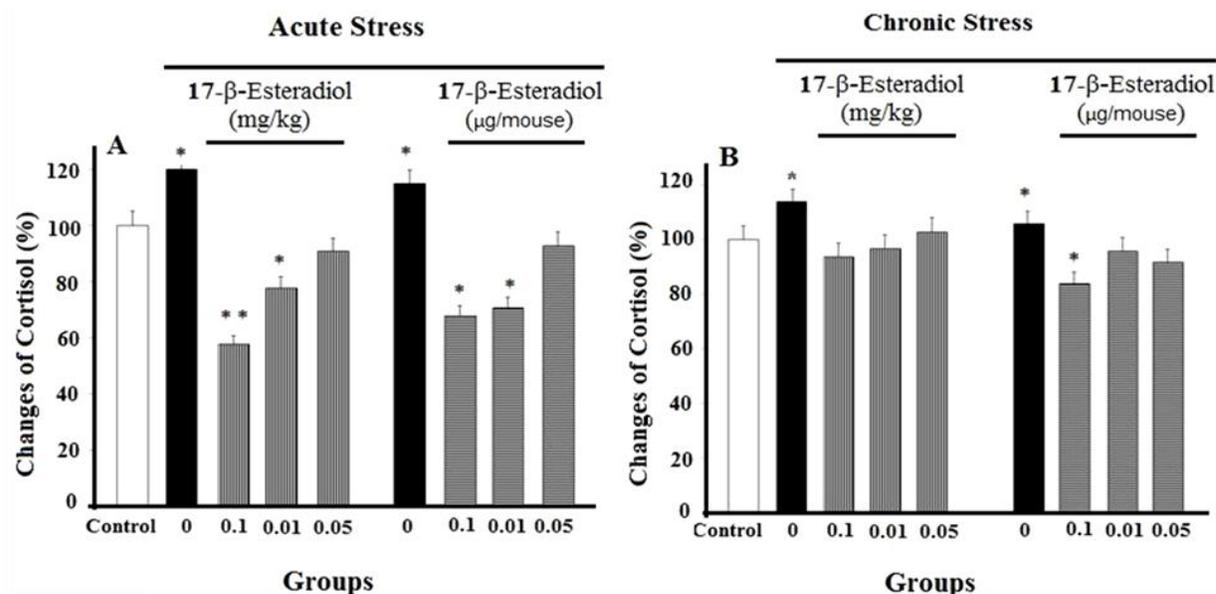
#### Data analysis

The data were expressed as mean  $\pm$  S.E.M. In order to analyze the data one-way and two-way analysis of variance (ANOVA), followed by Tukey test were used.  $P < 0.05$  was considered significant differences.

## RESULTS

### Effect of intraperitoneal and intraventricular injection of estradiol on plasma cortisol level after induction of acute and chronic stress

The results were considered 100, in the first day, and for the other days it is evaluated according to first day (percentage). The results showed that acute and chronic stress significantly increased plasma cortisol compared to the control group (Figure 1A). IP administration of estradiol could inhibit the effects of acute stress and significant decrease of cortisol concentrations, compared to the stress group. Also, ICV administration of estradiol also caused a significant reduction in cortisol concentration compared to stress (Figure 1A). Moreover, the results showed that chronic stress was also an increase in plasma cortisol. On the other hand only the ICV administration of estradiol and only in doses of 0.1  $\mu\text{g}/\text{mouse}$  could lead to a significant reduction in plasma cortisol levels, compared to the stress group (Figure 1B).



**Fig. 1:** Effect of intraperitoneal and intraventricular injection of estradiol on plasma cortisol during induction of acute (A) and chronic stress (B). Mean  $\pm$  S.E.M. for 6 animals. \*  $P < 0.05$  and \*\*  $P < 0.01$  is a significant difference compared to the control group.

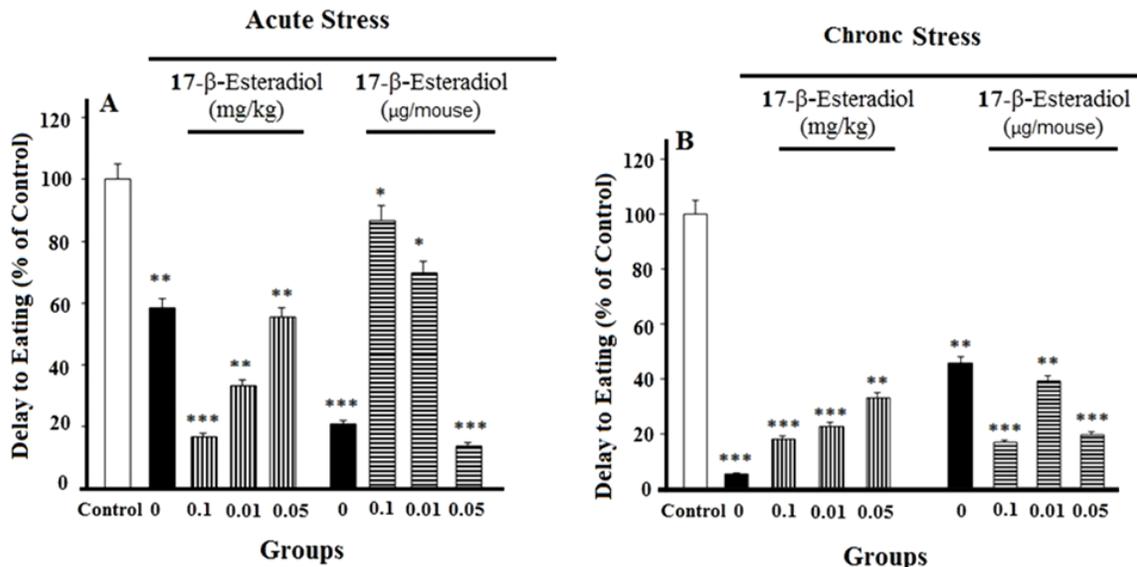
**Effects of intraperitoneal and intraventricular injection of estradiol on delay to eating after induction of acute and chronic stress**

The results showed that acute stress decreased delay to eating time in gonadectomized animal significantly, compared to controls. On the other hand, IP administration of estradiol at doses of 0.1 and 0.01 mg/kg in acute stress reduced delay to eating time significantly compared to acute stress groups. However, ICV injection of estradiol had different results than IP injection, which by reducing the estradiol dosage, the delay to eating time decreased (Fig. 2 A). Also, results showed that chronic stress reduces the delay to eating time in the ovariectomized animals, compared to the control group. IP administration of estradiol significantly increased delay to eating than chronic stress group and gradually reduce of the estradiol doses increased delay to eating, compared to the stress group. ICV injection of

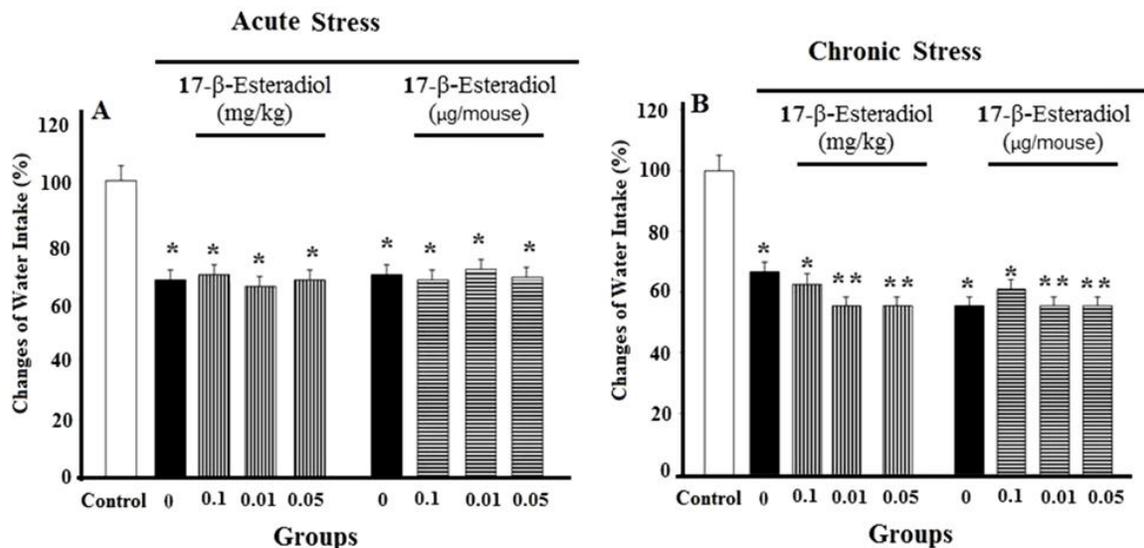
estradiol in 0.1 and 0.05 mg/kg reduced delay to eating compared to chronic stress group (Fig. 2 B).

**Effects of intraperitoneal and intraventricular estradiol injection in water intake after induction of acute and chronic stress**

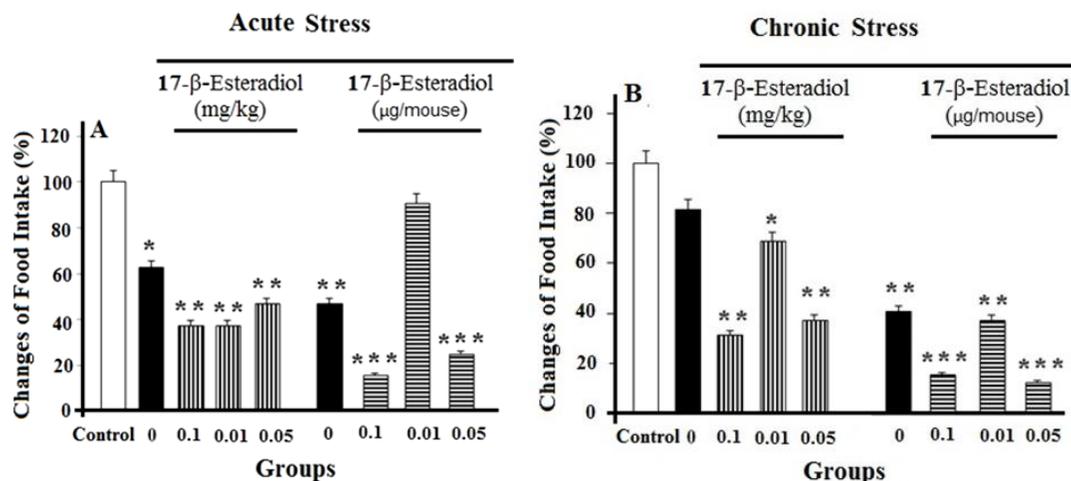
As shown in Figure 3 A, acute stress caused a significant reduction in water intake, compared to the control group. However, intraperitoneal and intraventricular injection of estradiol in ovariectomized mice did not cause any change in water intake, compared to stress group. Also, as shown in Figure 3 B, chronic stress also causes a significant reduction in water intake in ovariectomized mice, compared to the control group. Intraventricular and intraperitoneal injections of estradiol in ovariectomized mice didn't change water intake, compared to the chronic stress group.



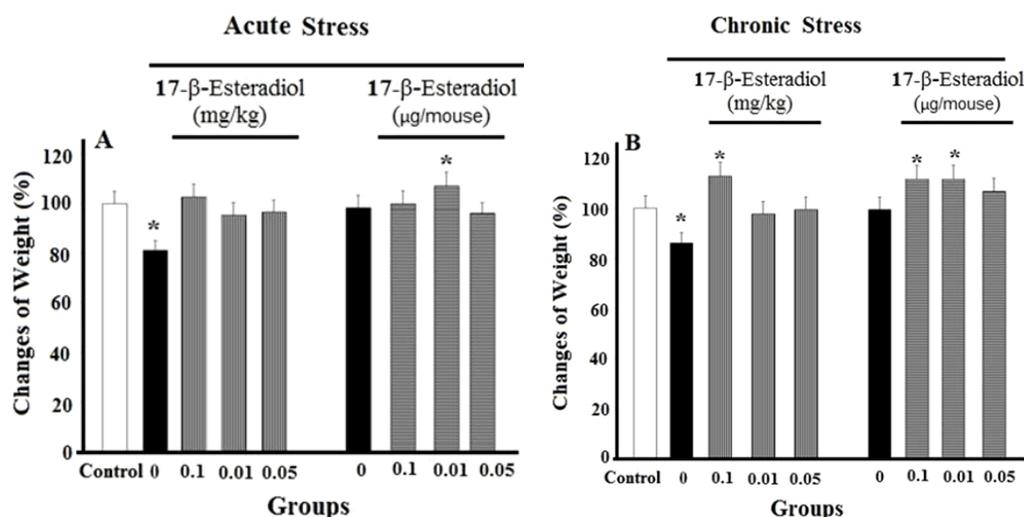
**Fig. 2:** Effect of intraperitoneal and intraventricular injection of estradiol on delay to eating during induction of acute (A) and chronic stresses (B). Mean ± S.E.M. for 6 animals. \* P<0.05, \*\* P<0.01, \*\*\* P<0.001 is a significant difference compared to the control group.



**Fig. 3:** Effect of intraperitoneal and intraventricular injection of estradiol in water intake during induction of acute (A) and chronic stress (B). Mean ± S.E.M. for 6 animals. \* P<0.05, \*\* P<0.01 is a significant difference compared to the control group.



**Fig. 4:** Effect of intraperitoneal and intraventricular injection of estradiol on food intake during induction of acute (A) and chronic stress (B). Mean  $\pm$  S.E.M. for 6 animals. \*  $P < 0.05$ , \*\*  $P < 0.01$ , \*\*\*  $P < 0.001$  is a significant difference compared to the control group.



**Fig. 5:** Effect of intraperitoneal and intraventricular injection of estradiol on change of weight during induction of acute (A) and chronic stress (B). Mean  $\pm$  S.E.M. for 6 animals. \*  $P < 0.05$  is a significant difference compared to the control group.

#### The Effect of intraventricular and intraperitoneal administrations of estradiol on food intake after induction of acute and chronic stress

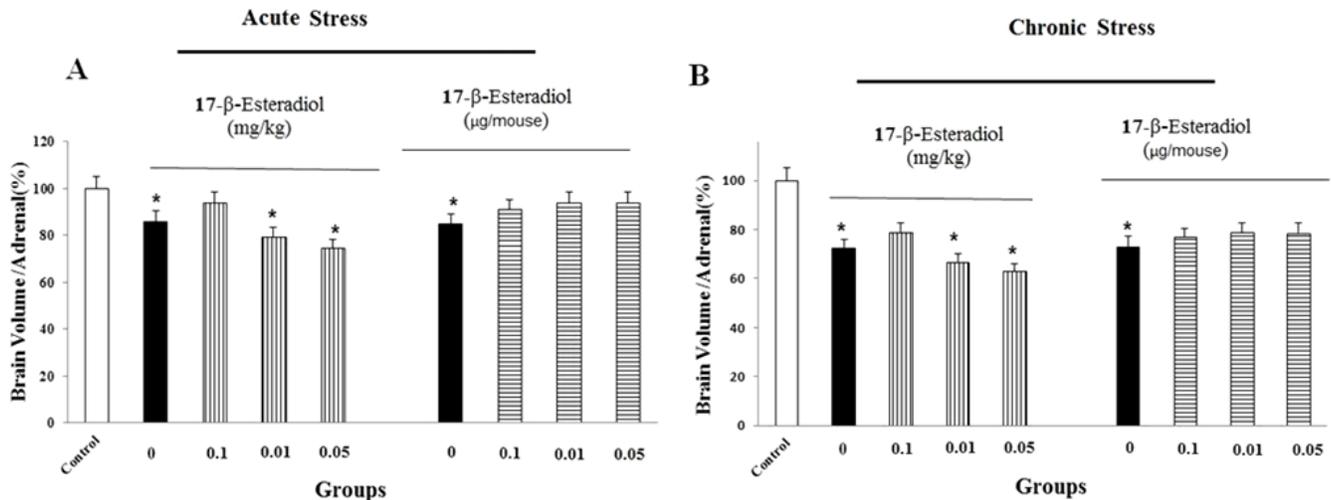
The results showed that acute stress caused a significant reduction in food intake in ovariectomized mice, compared to the control group. On the other hand, in the acute stress intraperitoneal administration of estradiol significantly decreased food intake compared to the stress. ICV injection of estradiol at a 0.01  $\mu\text{g}/\text{mouse}$  significantly increased food intake compared to the stress group.

However, the ICV injection of estradiol at 0.1 and 0.05  $\mu\text{g}/\text{mouse}$  caused a significant reduction in food intake in ovariectomized animals, compared to stress group (Figure 4 A). The present results also showed that chronic stress led to a significant reduction in food intake in ovariectomized mice. Both IP and ICV administration of estradiol caused significant reduction in food intake in ovariectomized mice, compared to

stress group (Figure 4 B).

#### Effect of intraventricular and intraperitoneal injection of estradiol on weight changes after induction of acute and chronic stress

Figure 5 A shows the effect of different doses of intraperitoneal and intraventricular estradiol on weight changes following acute stress. Acute stress causes weight loss in ovariectomized animals, compared to the control group. Both intraventricular and intraperitoneal administration of estradiol significantly increased the weight in ovariectomized animals, compared to the stress group ( $P < 0.05$ ). As shown in Figure 5 B chronic stress also causes weight loss in ovariectomized animals, compared to the control group and intraventricular and intraperitoneal administration of estradiol significantly increased the weight in ovariectomized animals, compared to stress group ( $P < 0.05$ ).



**Fig. 6:** Effects of acute (A) and chronic stress (B) of changes in the ratio of brain volume/adrenal gland. Animals received different doses of estradiol intraventricular (0.1, 0.1 and 0.05  $\mu\text{g}/\text{mouse}$ ) 5 minutes and intraperitoneal (0.1, 0.01, 0.05  $\text{mg}/\text{kg}$ ) 30 minutes before stress induction. Mean  $\pm$  S.E.M. for 6 animals. \* $P < 0.05$  compared to the control group.

### The effect of intraperitoneal and intraventricular administration of estradiol on brain volume/adrenal gland volume after induction of acute and chronic stress

The results showed that acute stress can decrease the ratio of brain to adrenal gland volume in animals. Intraperitoneal administration of estradiol only at a dose of 0.1  $\text{mg}/\text{kg}$  could inhibit this effect. Intraventricular administration of estradiol in all three doses (0.1, 0.01 and 0.05  $\mu\text{g}/\text{mouse}$ ) inhibited stress effect in ovariectomized mice (Figure 6 A). [Two-Way ANOVA; estradiol effect:  $F(1, 60) = 2.11, P < 0.05$ , side effect:  $F(2, 60) = 1.24, P < 0.05$ , side  $\times$  estradiol effects:  $F(10, 60) = 3.22, P < 0.05$ ]. The results showed that chronic stress also decreased brain/adrenal gland volume in stressed animals. Intraperitoneal injection of estradiol only at a dose of 0.1  $\text{mg}/\text{kg}$  could inhibit this effect. However, intraventricular injection of estradiol in all three doses of estradiol was able to increase the ratio of brain/adrenal gland volume in chronic stress animals (Figure b -6). [Two-Way ANOVA; estradiol effect:  $F(1, 60) = 1.98, P < 0.05$ , side effect:  $F(2, 60) = 1.343, P < 0.05$ , side  $\times$  estradiol effects:  $F(10, 60) = 1.87, P < 0.05$ ; left side].

### DISCUSSION

In the present study, the effects of both intraperitoneal and intraventricular administration of estradiol after acute and chronic uncontrolled stress (or inevitable), on plasma cortisol, anorexia, water and food intake, body weight and brain/adrenal volume in ovariectomized female mice were studied. The results showed that acute and chronic stress, increased plasma cortisol concentration which, injection of estradiol in ovariectomized mice could inhibit the effects of stress and a slight decrease plasma cortisol levels. Stress, particularly emotional stress has plenty of biomarkers but, increased plasma levels of glucocorticoid hormones such as cortisol and corticosterone are known as a

potential marker for the diagnosis of stress. Studies have shown that stress can increase HPA axis activity and therefore increase glucocorticoid hormones such as corticosterone and cortisol as one of the most important indicators of stress (Cohen *et al.*, 2012). However, Studies showed that corticosterone is the main glucocorticoid in rodents and especially mice, but also cortisol increase in mice during the stress (Gong *et al.*, 2015) and in this study, we measured plasma cortisol concentration which stress caused significant increased in plasma cortisol concentration.

Estradiol is steroid and can cross the blood-brain barrier. Also, estrogen is produced locally in the brain. The estrogen produced in the brain may play an important role in keeping the brain's autonomic tone (McEwen *et al.*, 2012). Estrogen could change neurotransmitter system function in the brain, like GABAergic, glutamatergic, dopaminergic and serotonergic system and alter the behavioral and electrophysiological function of brain (Barth *et al.*, 2015). Estrogen receptors exist in many brain regions and estrogen's signaling could prevent the harmful effects of stress (Toran-Allerand, 2005). Estrogen receptors via activation of the MAPK pathways, participates in mediating neuroprotection (Fiocchetti *et al.*, 2012). Estrogen via multiple mechanisms, such as increasing the inhibitory effects of glucocorticoids on the HPA axis, reducing corticosterone and cortisol metabolism, increases the synthesis of corticosterone or cortisol and increased sensitivity or reduces the degradation of glucocorticoid receptors, affects the stress response (Bagheri Nikoo *et al.*, 2014). In the present study, perhaps decreasing effect of estradiol administration on plasma cortisol concentrations after acute and chronic stress is through these mechanisms.

In another part of this study the results showed that acute and chronic stress in ovariectomized mice decreased weight, food and water intake. In acute and chronic stress intraperitoneal and intraventricular administration of estradiol at 0.1 and 0.01 doses cause weight gain in animals.

The results also showed that intraperitoneal administration of estradiol decreases anorexia in acute stress (especially in 0.1 mg/kg), but in 0.05 mg/kg anorexia increased. In acute stress intraventricular injection of estradiol at a dose 0.05 µg/mouse decreased anorexia significantly, but in 0.1 and 0.01 µg/mouse anorexia increased than the acute stress group. It is believed that overeating due to stress, may lead to metabolic diseases such as obesity and diabetes (Mikolajczyk *et al.*, 2009). High concentrations of cortisol in plasma and brain cause brain reward system to get highly sensitive and this sensitivity increased tendency to use fat and feeding activity. Cortisol may affect the reward value of food via peptide mediators such as ghrelin (Yau and Potenza, 2013).

Ghrelin is a peptide which increases appetite that is produced mainly in the stomach and is seen in the blood circulation of healthy human, but its concentration increased in people under stress. It has been shown that ghrelin also play a similar role to increases of food intake, weight gain and obesity in rodents as in humans (Patterson *et al.*, 2011). Since chronic stress can lead to exposure of the brain and body to high levels of cortisol and this have direct and indirect effects on reward systems. In the acute stress the HPA axis modulation of food intake allows the stressful phenomenon to be reduced with and the energy used to be replaced afterward, but, in chronic stress glucocorticoids can lead to chronically stimulate eating behavior and excessive weight gain. In addition, stress can increase palatable food via its interaction with central reward pathways. Activation of this system can also interface with the HPA axis to inhibit its more activation, which means not only can stress, enhance eating behavior, but eating can inhibit the HPA axis and stress (Yau and Potenza, 2013; Sominsky and pencer, 2014).

The results of this study showed that acute and chronic stress reduced water intake in female mice. However, it has been shown that stress could increase water intake by stimulating the secretion of CRF and vasopressin, which inconsistent with our results (Elman *et al.*, 2003). However, previous studies showed that the effects of stress in male mice are different from female. For instance, Ranjbaran *et al.* (2013) and Bagheri Nikoo *et al.* (2014) have been shown that electro foot shock stress in male mice increased water intake. Estrogen receptors contain two major types (ER $\alpha$  and ER $\beta$ ) which, describing ER $\beta$  express at high levels by neurons within the paraventricular nucleus (PVN) and a large percent of ER $\beta$  cells in the PVN are arginine vasopressin (AVP) positive, but ER $\beta$  is also found in some CRH containing neurons of the PVN. ER $\alpha$  is found at low levels and just in the periventricular PVN and barely in CRH, AVP or oxytocin neurons, which prevent a direct action of estradiol as mediated through ER $\alpha$  on PVN responses to stress. Nonetheless, ER $\alpha$  is colocalised with GAD67 (the enzyme which involve in the synthesis of GABA) in neurons surrounding the PVN. One explanation from this localization would be that ER $\alpha$  can modulate inhibitory input to the PVN, hence affecting HPA axis function through trans-synaptic mechanisms (Handa *et al.*, 2009; Miller *et al.*, 2004; Bahari *et al.*, 2015). This may be able to explain, which

water intake decreased during stress in female mice. In other part of this study, the effect of acute and chronic stress on the changes in the ratio of brain volume/adrenal gland volume was measured. Results showed that stress can cause a decrease in the ratio of brain volume to adrenal gland volume. In the present study, estradiol inhibited the damaging effects of stress, so that intraperitoneal injection of estrogen (0.1 mg/kg) and ICV injection (0.1, 0.01 and 0.05 µg/mouse) increased brain volume/ adrenal gland volume.

The brain is a target of stress, which showing stress-induced dendrite atrophy and shrinkage, loss of dendrite spines in neuronal populations, heightened susceptibility to cell death and decreased size and weight of some parts of the brain such as the hippocampus (Lee *et al.*, 2009; Liston *et al.*, 2006). Thus, it seems that stress could damage the neurons, but estradiol by binding to its receptor activates neurite growth, such as nerve growth factor (NGF) and brain-derived nerve growth factor (BDNF), increase synaptogenesis, modulating immediate-early-gene expression, kinase activity and calcium handling and prevents neuronal damages (McCarthy, 2008). Also, there is indicating that in older women, due to lower estrogen levels during menopause, brain volume began to decrease. This atrophy occurs particularly in the hippocampus and the parietal lobe that these areas associated with memory and cognition. Thus, lack of estrogen affected this area of the brain and can cause atrophy and impairment in learning and memory (Shepherd, 2001). Glutamatergic system is another mechanism that can be changed during stress. Acute exposure to stress rapidly increases glutamate release in several limbic and cortical areas, including the prefrontal cortex, hippocampus and amygdala (Bahari *et al.*, 2014; Hussein *et al.*, 2016; Venero and Borrell, 1999). It has been shown that repeated unpredictable stress causes a loss of surface AMPAR and NMDAR glutamate subunits in prefrontal neurons and the level of total NR1 and GluR1 subunits in the prefrontal is also greatly reduced by repeated exposure to stress (Yuen *et al.*, 2012).

Thus, in chronic stress animals' disrupted membrane trafficking or synthesis of glutamate receptors may contribute to the loss of glutamatergic transmission and could damage neurons. Yokomaku *et al.* showed that estrogen exhibit facilitating effect on glutamate transmission (Yokomaku *et al.*, 2003). The mechanisms which estrogen effects on cognition are related to NMDA glutamate receptors. For instance, it has been shown that estrogen promotes an increase in NMDA receptor subunit expression and binding sites and improves neuronal function and prevents neuronal damage (Adams *et al.*, 2004).

Ulrich-Lai *et al.* displayed that chronic stress, increase weight of the adrenal gland by hyperplasia of the outer zona fasciculata and hypertrophy of the inner zona fasciculata and medulla. Thus, these findings are consistent with the present results demonstrating brain volume/adrenal volume decrease during stress. Stress also increased adrenal medullary size and/or catecholamine content is frequently observed after other types of stress. Furthermore, after stress there is a generalized raise in medullary function, which suggesting that medullary hypertrophy

may be a general consequence of chronic stress (Hosseini *et al.*, 2015; Miyamoto *et al.*, 1999; Rezaeian Dalooei *et al.*, 2016; Ghodrat *et al.*, 2014).

## CONCLUSION

Therefore, these findings suggest that administration of intraperitoneal (IP) and intraventricular (ICV) injection of estradiol in female adult ovariectomized mice can protect against the hormonal and metabolic changes of acute and chronic stress. Among the indications of these results are those understandings the mechanisms of estrogen neuroprotection will allow for the development of more effective estrogenic agents useful for the treatment of stress.

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**Conflict of Interests:** There are no conflicts of interest.

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