

Statistical Optimization of Olanzapine Ternary Solid Dispersions with Pvp K 30 and Peg 20,000 by Response Surface Methodology

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ABSTRACT

The aim of present study was to improve dissolution rate of olanzapine by means of solid dispersion using combination of hydrophilic polymer (PEG & PVP) by using response surface design. Solid dispersion containing olanzapine were prepared using PEG 20000 & PVP K 30 by melted fusion method. Response surface method was used for the optimization olanzapine solid dispersions. Amount of PEG 20000 and Amount of PVP K 30 were selected as the critical process parameters (Independent variable) whereas amount dissolved in 10 minute (Q_{10}) and amount dissolved in 45 minute (Q_{45}) were selected as critical quality attributes (dependent variables). Optimized solid dispersion batch was characterized using infrared spectroscopy, differential scanning calorimetry (DSC) and X-ray diffractometry (XRD). Dissolution studies indicated a significant improvement in dissolution of olanzapine when dispersed in PEG 20000 and PVP k 30. XRD and DSC study indicated amorphous form of prepared solid dispersions. On the basis of numerical optimization technique, PEG 20000(X_1) and amount of PVP K 30(X_2) were 11.20 % and 14.53 % in optimized solid dispersion. The observed responses were closed well with the predicted values. The response surface method is found to be robust and accurate for optimization of solid dispersion for increase in solubility and dissolution rate of olanzapine, coherent with the needs of poorly water soluble drugs.

INTRODUCTION

Olanzapine is an atypical antipsychotic. It is approved for the treatment of schizophrenia and bipolar disorder. Olanzapine is a BCS II drug (Poor solubility and high permeability)(Dixit, *et al.*, 2011). The bioavailability of a BCS class II drug is rate-limited by its dissolution, so that even a small increase in dissolution rate sometimes results in a large increase in bioavailability (Lobenberg and Amidon, 2000). Therefore, an enhancement of the dissolution rate of the drug is thought to be a key factor for improving the bioavailability of BCS class II drugs. Crystal modification particle size reduction, self-emulsification, pH modification, and amorphization are considered to be effective for improving the dissolution behaviour of BCS class II drugs (Kaushal, *et al.*, 2004) (Xia, *et al.*, 2010, Tran, *et al.*, 2010).

Solid dispersions are one of the most successful strategies to improve drug release of poorly soluble drugs. These can be defined as molecular mixtures of poorly water soluble drugs in hydrophilic carriers, which present a drug release profile that is driven by the polymer properties (Chiou and Riegelman, 1971a, Das, *et al.*). The first drug whose rate and extent of absorption was significantly enhanced using the solid dispersion technique was sulfathiazole by Sekiguchi and Obi (Sekiguchi and Obi, 1961). Over the years, a variety of carriers have been used to prepare solid dispersion of olanzapine to improve dissolution rate such as; Croscarmellose sodium (Krishnamoorthy *et al.*, 2011), Mannitol (Krishnamoorthy, *et al.*, 2012), pregelatinised starch (PGS) and sodium starch glycollate (Krishnamoorthya, *et al.*, 2011). Polyethylene glycols (PEG), polymers of ethylene oxide are widely used as vehicles for solid dispersions because of their low melting point, rapid solidification rate, capability of forming solid drug solutions, low toxicity and low costs (Betageri and Makarla, 1995, Chiou and Riegelman, 1971b).

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However, at higher drug concentrations, the drug is often present in the crystalline form within the PEG dispersion or it recrystallizes over time, resulting in unstable formulations with lower dissolution rates.

The combination of two hydrophilic polymeric carriers like PEG and PVP could improve the solubility as well as dissolution profiles of various poorly aqueous soluble drugs due to inhibition of drug recrystallization as well as a rapid solidification rate (Hasnain and Nayak, 2012, Bley, *et al.*, 2010, Guedes, *et al.*, 2011). Design of experiment could be used for the statistical optimization of pharmaceutical dosage forms. Central composite design is a type of response surface methodology that requires smaller number of experimental runs and is less time consuming than conventional formulation methods (Elbary, *et al.*, December 2011, Akhgari, *et al.*, 2005). The aim of this study is to improve dissolution rate of olanzapine by means of solid dispersion using combination of hydrophilic polymer (PEG & PVP). The physical-chemical properties of olanzapine in solid dispersion was studied by differential scanning calorimeter (DSC), X-ray diffraction (XRD) and Fourier transform infrared (FTIR). Dissolution profile of solid dispersion also studied. In order to accomplish this aim, computer-aided optimization techniques using two-factor, three-level central composite design was used to determine the effect of two formulation factors, namely, the contents of PEG 20000 and PVP K 30 on the dissolution of drug and to statistically optimize the levels of these factors using multivariate analysis and response surface plots in order to incur the targeted dissolution rate for olanzapine.

MATERIAL & METHODS

Materials

Olanzapine was kindly gifted by Micro labs Bangalore. PEG 20000 and PVP K 30 were obtained from SD Fine Chemicals Ltd. (Mumbai, India). All other materials and reagents were of analytical grade of purity.

Preparation of solid dispersion

Solid dispersion containing 12.9 % olanzapine were prepared using PEG 20000 & PVP K 30 by melted fusion method. The drug and polymer were heated until the polymer melt. The molten mixture was stirred until the drug was dissolved completely in the melt and a homogeneous solution was obtained. The solution was brought to solidification by quick cooling. It was kept in desiccators under vacuum for 24 hr. Then solid dispersion formulation was pulverized using mortar and pestle. The pulverized powder was classified using the sieve # 60.

Experimental Design

A central composite response surface design (**DESIGN EXPERT 8.0.1 demo version software**) was used for the optimization olanzapine solid dispersions. Amount of PEG 20000 (A) and Amount of PVP K 30 were selected as the independent variables whereas Q₁₀ (amount dissolved in 10 minute) and Q₄₅

(amount dissolved in 45 minute) were selected as dependent variables. Levels for two factors are presented in Table 1.

Table 1: Central composite response surface design layout.

Formulation	Variables in coded Form		Q10	Q45
	X1(%)	X2(%)		
SD 1	-1.000	-1.000	14.72	54.464
SD 2	1.00	-1.000	16.14	62.464
SD 3	-1.000	1.000	31.69	88.69
SD 4	1.000	1.000	36.21	89.42
SD 5	-1.414	0.000	24.56	76.23
SD 6	1.414	0.000	29.07	82.49
SD 7	0.000	-1.414	9.21	45.76
SD 8	0.000	1.414	34.76	89.32
SD 9	0.000	0.000	25.998	79.162
SD 10	11.20	14.53	24.70	76.13
Coded Value	Actual Value (%)			
	X1	X2		
-1.414	7.93	7.93		
-1.000	10.00	10.00		
0.000	15.00	15.00		
1.000	20.00	20.00		
1.414	22.07	22.07		

X1 indicates amount of PEG 20000 (%); X2, amount of PVP K 30 (%); Q₁₀, amount dissolved in 10 minute; and Q₄₅, amount dissolved in 45 minute. SD 10 used as checks point and optimized batch.

Different trial formulations of olanzapine solid dispersions were prepared according to the trial proposal of central composite response surface design. The prepared solid dispersions of Olanzapine were evaluated for dissolution study. The responses were analyzed using ANOVA and the individual response parameters were evaluated using F test and polynomial equation, generated for each response using multiple linear regression analysis (MLRA). The study design including investigated factors and responses is shown in Table 1. The optimized formulation was prepared which have the Q₁₀ in range 20-25 % and Q₄₅ in range 75-80 %. Constraints for responses and factors are shown in Table 2. By utilizing the software, we got one solution for optimized formulation. The optimized formulation was prepared and evaluated for Q₁₀ and Q₄₅. Observe response value of the optimized formulation was compared with predicted value. The optimised batch(s) was further investigated by DSC, XRD, and FTIR.

Table 2: Optimization of olanzapine solid dispersion.

Name	Goal	Constraints		
		Lower limit	Upper Limit	
PEG 20000	In range	-1.414	1.414	
PVP K 30	In range	-1.414	1.414	
Q ₁₀ (%)	In range	20	25	
Q ₄₅ (%)	In range	75	80	
SOLUTION (SD 10)				
PEG 20000	PVP K 30	Q ₁₀ (%)	Q ₄₅ (%)	Desirability
11.20	14.53	24.70	76.13	1.000

Q₁₀ indicates amount dissolved in 10 minute; and Q₄₅, amount dissolved in 45 minute. SD 10 used as checks point and optimized batch.

In vitro dissolution study

Drug dissolution studies was carried out using USP dissolution apparatus 2 using a paddle at a speed of 100 rpm with 900 mL of Phosphate buffer pH 7.4. as dissolution medium at

37°C. Solid dispersion powders containing 50 mg of olanzapine were dispersed on the surface of the dissolution medium and the time was recorded. At intervals, 5 mL samples were withdrawn through a filter.

All the readings were blanked with same media as was used in the dissolution study. The olanzapine content was measured by HPLC method and the percentage of drug released was calculated using calibration curves.

Drug release kinetic

The release from the different SD formulations was determined by curve fitting method. Data obtained from in vitro release studies were fitted to various kinetic equations.

Zero order Model: $Q_t = Q_0 + k_0t$, where Q_t is the amount of drug dissolved in time t , Q_0 is the initial amount of drug in the solution and K_0 is the zero order release constant.

First order Model: $\ln(Q_\infty - Q_t) = \ln Q_0 + kt$, where Q_t is the amount of drug dissolved in time t , Q_0 is the initial amount of drug in the solution, Q_∞ is the amount release in time ∞ (100 % drug release) and K is the first order release constant.

Korsmeyer–Peppas Model (power law): $\frac{Q_t}{Q_\infty} = k_k t^n$, or $\log Q_t = \log k_k + n \log t$ where Q_t is the amount of drug dissolved in time t , Q_∞ is the amount release in time ∞ , k_k is the rate constant and n is the diffusional exponent, this indicates the drug release mechanism.

X-ray diffraction

Powder X-ray diffraction patterns were obtained with a diffractometer (Geigerflex, RAD-IB, Rigaku, Tokyo). The operating conditions were as follows: target, Cu; filter, Ni; voltage, 40kV; current, 20 mA and scanning speed, $2\theta = 4^\circ/\text{min}$. Physical mixtures, as control of the solid dispersion, were prepared by simply mixing the powdered olanzapine and polymers at the same composition ratios as those of the solid dispersions.

Differential scanning calorimetry (DSC)

DSC studies of Olanzapine and optimized solid dispersion batch were conducted using a Perkin Elmer DSC-4 differential scanning calorimeter using aluminium sample pans for volatiles. Samples (about 5 mg) were heated at $10^\circ\text{C}/\text{min}$ using nitrogen as the purging gas.

Fourier transform infrared spectroscopy (FTIR)

An approximately minimum quantity (about 1mg) of sample was thoroughly blended with adequate quantity of IR grade KBR (about 5mg) in a mortar. The mix was then made into thin films on a sample plate using a hand operated compression lever. The samples were then analyzed in a Perkin Elmer Model 1330 double beam IR spectrometer using KBr film as negative control (blank).

RESULT AND DISCUSSION

A total of 9 trial formulations of olanzapine solid dispersion were proposed by central composite response surface design for two independent variables: Amount of PEG 20000 and amount of PVP K 30 which were varied at three different levels. All the batches of solid dispersion were evaluated for its amount dissolved in 10 minute (Q_{10}) and amount dissolved in 45 minute (Q_{45}). The dependent variables obtained at three levels of the 2 independent variables (X_1 and X_2) was subjected to multiple regression to yield a interactive and polynomial equation. The polynomial equation generated by this experimental design was as follows:

$$Y_i = b_0 + b_1X_1 + b_2X_2 + b_{12}X_1X_2 + b_{11}X_1^2 + b_{22}X_2^2$$

Where y_i is the dependent variable, b_0 is the arithmetic mean response of the 9 runs; and b_1 and b_2 are the estimated coefficients for the independent factors X_1 and X_2 , respectively. The main effects (X_1 and X_2) represent the average result of changing one factor at a time from its low to high value. The interaction term (X_1X_2) shows how the response changes when 2 factors are simultaneously changed. The polynomial terms (X_1^2 and X_2^2) are including investigating nonlinearity.

The Q_{10} and Q_{45} for the 9 trial batches (SD 1 to SD 9) showed a wide variation from 9.21 to 36.21% and from 45.76 to 89.42 % respectively (Table 1). The observation show that the Q_{10} and Q_{45} strongly depends upon the selected independent variables. The model (full and reduced) relating the responses, Q_{10} and Q_{45} to the transformed factor are shown in table 4. The result of the analysis of variance (ANOVA) for the model simplification by eliminating non significant terms ($P > 0.05$) were shown in table 5. For both Q_{10} and Q_{45} term X_1^2 was found to be insignificant (p value > 0.05), hence this was omitted from the full model to generate the reduced models. The F value in the ANOVA table was the ratio of model mean square (MS) to the appropriate error (i.e. residual) mean square. The larger the F value and the more likely that the variance contributed by the model was significantly larger than random error. The model F -value and high R square values suggested that these models were significant.

The results of multiple linear regression analysis reveal that both the coefficient b_1 and b_2 bear a positive sign for both Q_{10} and Q_{45} . Therefore, increasing the amount of either PEG 20000 or PVP K 30 is expected to increase the Q_{10} and Q_{45} . Three-dimensional response surface plots and corresponding contour plots to study the effects of the independent variables (factors) on each dependent variable (response) were presented in Figure 1 and 2.

A numerical optimization technique based on the desirability approaches was adopted to achieve new optimized solid dispersion (SD 10) which was also used as the check point. For evaluation the optimization capability of response surface factorial design. The variable settling used for the formulation of optimized solid dispersion were amount of PEG 20000 (X_1) and amount of PVP K 30 (X_2) were 11.20 % and 14.53 % (Fig.3 and

table 2). The optimized formulations were prepared with the optimized amount of independent variables. Observed values were found similar to predicted values in SD 10. Thus, we can conclude that the statistical model was mathematically valid.

In vitro dissolution study

Drug release from the different solid dispersion formulation and pure powdered drug is shown in figure 4. Compared with the pure olanzapine, the dissolution of olanzapine increases by PEG-PVP solid dispersion. The increase in dissolution of olanzapine with PEG-PVP could be attributed by the reduction in crystallinity of olanzapine. All the release profile shows two different phases of drug release. An initial rapid release phase followed by a slower one. Same result was reported by Omaira A et al (2006) for rofecoxib solid dispersion (Omaira A Sammour, *et al.*, 2006).

Drug release kinetic

The drug release data were analyzed using the zero-order and first-order equations to determine the drug release kinetics from the solid dispersion formulation. The regression coefficient (r^2) data based on kinetic analysis using various release models are listed in Table 5. To find out the mechanism of drug release first 60% drug release data was fitted in Korsmeyer–Peppas model, in which log cumulative percentage of drug release was plotted against time.

The regression coefficients (table 5) obtained for zero order kinetics were found to be higher ($r^2 = 0.9862$ to 0.9994) when compared with those of first order kinetics ($r^2 = 0.9387$ to $0.0.998$), indicating that drug release independent of concentration from all formulation. All the formulation shows good linearity ($r^2 = 0.9890$ to 0.9986), with the slope (n) values 0.5896 to 1.014 , indicating that release mechanism was anomalous non-Fickian or anomalous release ($0.5 < n < 1.0$), which indicates that the drug release occurred through diffusion in the hydrated matrix and polymer relaxation.

Powder X-Ray diffraction study.

The x-ray powder diffraction (XRD) study of olanzapine and optimized solid dispersion is done in the following manners. The angular range is 5 to 50° , 2θ counts are accumulated for 1 sec at each step. A typical x ray diffraction of olanzapine & solid dispersion are shown in figure 5. The pure olanzapine XRD showed numerous sharp narrow and intense peaks indicating its high crystallinity. Solid dispersion of olanzapine did not show the characteristic peak, indicates reduction in crystalline and phase transition from crystalline to amorphous form in the solid dispersion sample.

Differential scanning calorimetry (DSC)

DSC curve obtained for pure olanzapine and solid dispersion prepared by melting method. Pure olanzapine showed a melting endotherm 197.5°C . The DSC of solid dispersion showed melting point for the PEG around 59°C with no endothermic peak corresponding to the Olanzapine (figure 6). The absence of peak at temp corresponding to the melting of the drug could potentially be assigned to the solubility and distribution of the drug within the polymer matrix resulting in the conversion of crystalline drug form into amorphous form.

Fourier transforms infrared spectroscopy (FTIR)

The IR spectra for olanzapine was characterised by sharp transition occurring at 1033 , 2225 , 1559 and 745 cm^{-1} corresponding to the bond stretching associated with O-H bending, N-H stretching, C = N stretching and C-S bending respectively. Analysis of the spectra for the solid dispersion of olanzapine (figure 7) did not reveal any changes for the specific absorption bands for the drug suggesting a lack of interaction with carrier moieties. It was also noticed that the significant peaks of pure drug were found to be less sharpness and more broadness in the solid dispersion sample. These findings clearly prove the reduction of crystallinity in the drug molecule present in samples (AYALA, *et al.*, 2006, AYALA, 2007, Krishnamoorthy, *et al.*, 2012).

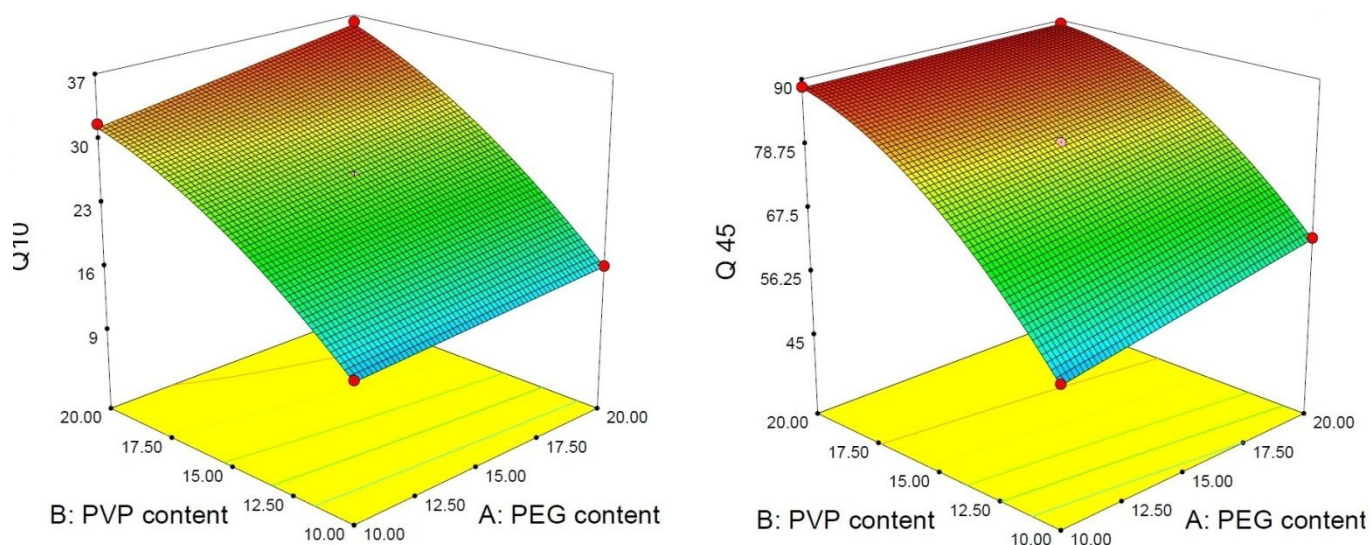


Fig. 1: 3 D response surface plot showed the effect of amount of PEG 20000 (X1) and amount of PVP K 30 (X2) on Q 10 (amount release at 10 minute) and Q 45 (amount release at 45 minute).

Table 3: Summary of results of regression analysis.

Response	For Q ₁₀					
	b ₀	b ₁	b ₂	b ₁₂	b ₁₁	b ₂₂
FM	26.0	1.54	9.15	0.78	0.48	- 1.93
P value	-	0.0008	< 0.0001	0.0159	0.0793	0.0018
RM	26.70	1.54	9.15	0.78	-	- 2.24
P value	-	0.0009	< 0.0001	0.0345	-	0.0005
Response	For Q ₄₅					
	b ₀	b ₁	b ₂	b ₁₂	b ₁₁	b ₂₂
FM	79.16	2.20	15.35	-0.1.82	0.18	-5.73
P value	-	0.0002	< 0.0001	0.0009	0.3435	< 0.0001
RM	79.42	2.20	15.35	-0.1.82	-	-5.85
P value	-	< 0.0001	< 0.0001	0.0002	-	< 0.0001

FM indicates Full model and RM, Reduce Model.

Table 4: Result of Analysis of variance (ANOVA).

Regression	For Q ₁₀					R ²
	Df	SS	MS	F		
FM	5	716	143	1456.23		0.9996
RM	4	715.19	178.80	738.80		0.9986
Residual						
FM	3	0.29	0.098			
RM	4	0.97	0.24			
Regression	For Q ₄₅					R ²
	Df	SS	MS	F		
FM	5	2103.55	420.71	5852.91		0.9999
RM	4	2103.46	525.86	6870.95		0.9999
Residual						
FM	3	0.22	0.072			
RM	4	0.31	0.077			

DF indicates: degrees of freedom; SS, sum of squares; MS, mean of squares; F, Fischer's ratio; R², regression coefficient

Table 5: Regression coefficient for different drug release kinetic model.

Formulation	Zero Order Kinetic	First order Kinetic	Korsmeyer–Peppas Model	
	r ²	r ²	r ²	n
SD 1	0.998	0.983	0.9968	0.94
SD 2	0.9973	0.993	0.9979	0.97
SD 3	0.9971	0.9431	0.9986	0.65
SD 4	0.9958	0.9425	0.9969	0.55
SD 5	0.9969	0.9708	0.9980	0.76
SD 6	0.9943	0.9666	0.9964	0.73
SD 7	0.9994	0.9908	0.9982	1.01
SD 8	0.9868	0.9387	0.9890	0.71
SD 9	0.9942	0.9711	0.9957	0.80

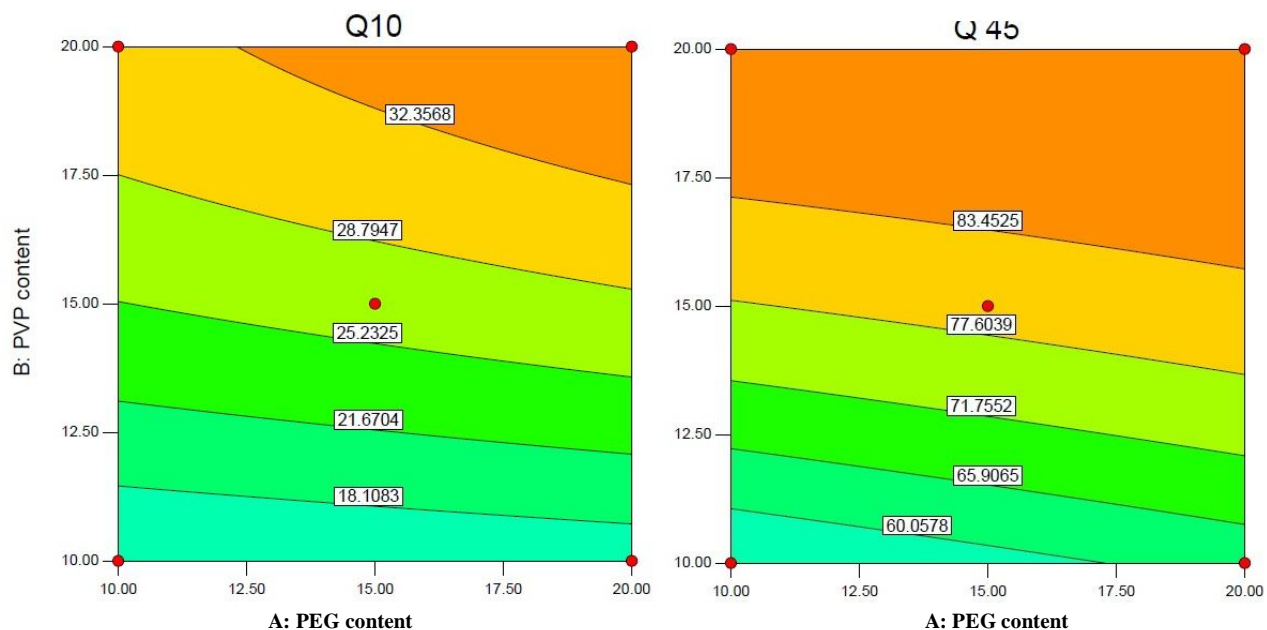


Fig. 2: Contour surface plot showed the effect of amount of PEG 20000 (X₁) and amount of PVP K 30 (X₂) on Q₁₀ (amount release at 10 minute) and Q₄₅ (amount release at 45 minute).

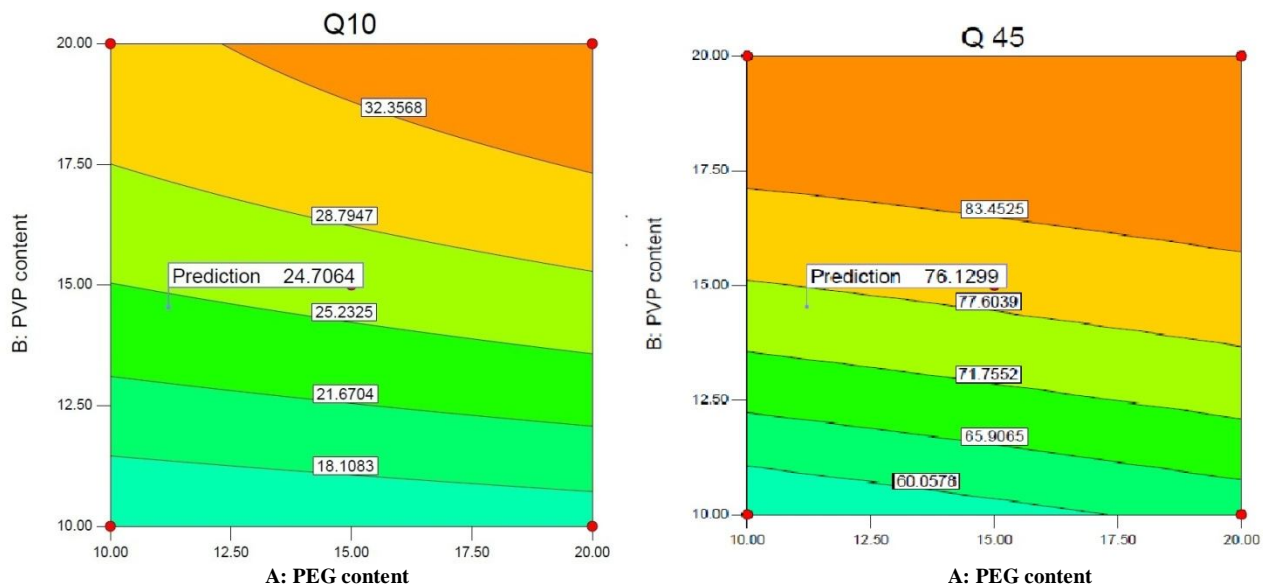


Fig. 3: Contour response surface Q 10 (amount release at 10 minute) and Q 45 (amount release at 45 minute) prediction plot.

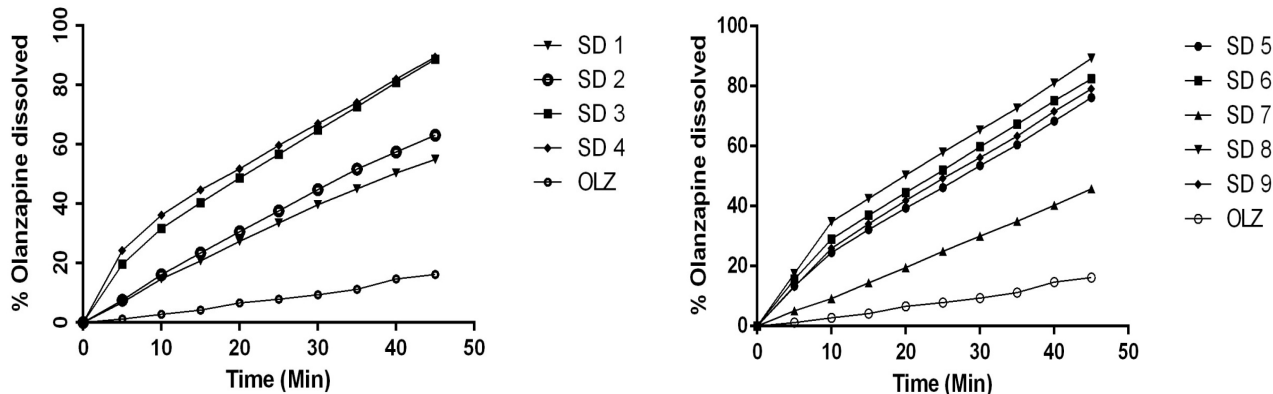


Fig. 4: Drug release from the different solid dispersion formulation and pure powdered drug.

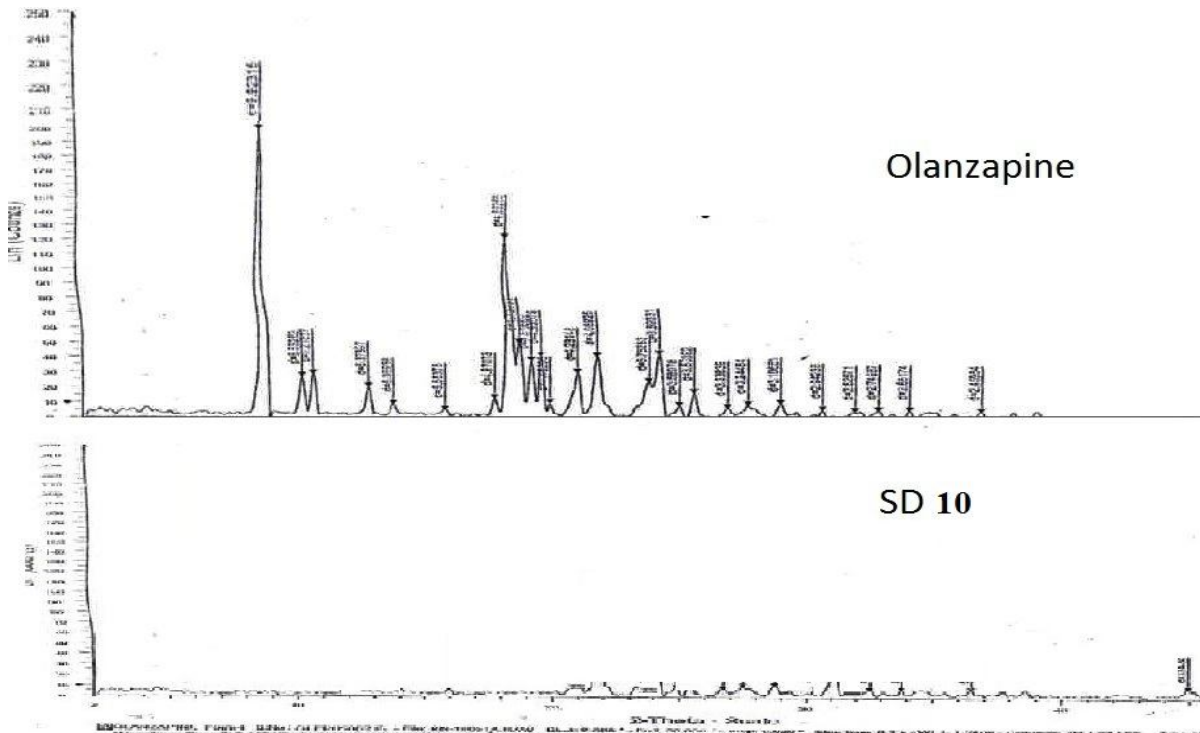


Fig. 5: X-ray powder differtometry (XRD) study of olanzapine and optimized solid dispersion (SD 10).

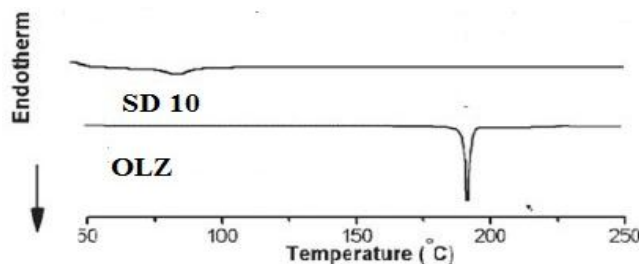


Fig. 6: DSC study of olanzapine and optimized solid dispersion (SD 10).

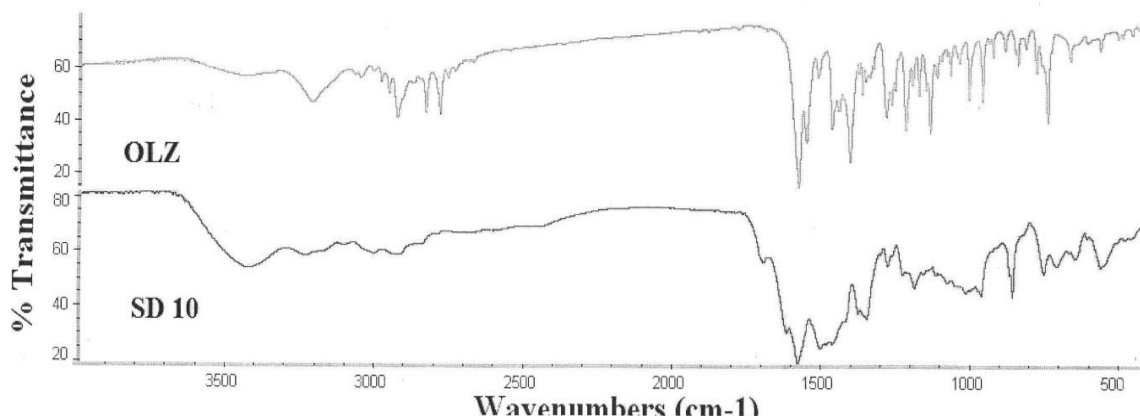


Fig. 7: FTIR study of olanzapine and optimized solid dispersion (SD 10).

CONCLUSION

The dissolution of olanzapine may be improved by the formation of solid dispersions with PEG 20000 and PVP K30 blends as carrier by the melting method. The central composite design was used to optimize the formulation. The three-dimensional response surface plots and corresponding contour plots relating Y_1 (Q_{10}) and Y_2 (Q_{45}) indicated the synergistic effect on both responses with the increase in X_1 (amount of PEG 20000) value and X_2 (amount of PVP K30) value. The optimized formulation with PEG 20000 (11.20 %) and PVP K 30 (14.53 %) showed 24.7 % Q_{10} and 76.132 % Q_{45} . Thus the formulation can be considered as one of the promising tool for improvement of dissolution and solubility of olanzapine.

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