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## Neuropharmacological study of *Argemone mexicana* Linn.

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## ABSTRACT

In the present study methanolic and ethyl acetate extracts of *Argemone mexicana* whole plant (Papaveraceae) were tested orally in swiss albino mice at doses of 100 mg/kg, 200 mg/kg and 400 mg/kg b.w. for CNS related activities. Papaveraceae family is known to have CNS depressant activity, so *A. mexicana* was evaluated for CNS activities. Significant central and peripheral nociceptive activity was observed for both extracts. Methanolic and ethyl acetate extract have also showed significant decrease in motor activity and fall off time of animals on rotating rod, along with sedative effect by potentiating phenobarbitone-induced sleeping time. In the acute toxicity study, both extracts was found to be safe upto 2500 mg/kg b.w. These results suggested that methanolic and ethyl acetate extracts of *Argemone mexicana* show analgesic, anxiolytic and sedative effects.

Key words: Argemone mexicana, hot plate test, writhing test, locomotor activity, muscle relaxant activity

## INTRODUCTION

Argemone mexicana (Papaveraceae) is a prickly glabrous herb with yellow juice and flashy yellow flowers, height of this plant varies between 0.3 to 0.12 m, Leaves are thistle like. Flowering time is all round the year in Indian circumstances and the flowers are scented and hermaphrodite. The plant prefers light sandy soils, requires well-drained soil and can grow in nutritionally poor soil and also prefers acid, neutral and basic (alkaline) soils (Rastogi R. P., 1979). A. mexicana is used by traditional healers in Mali to treat malaria (Willcox ML et al., 2007), externally in the treatment of cataracts and internally in the treatment of dropsy and jaundice. The root is used for the treatment of chronic skin diseases and alterative (Chevallier A., 1996, Chopra. R. N. et al 1986). The seeds are used as demulcent, emetic, expectorant, laxative and also used as an antidote in snake poisoning. The seed oil is purgative and used in the treatment of skin problems (Chevallier A., 1996 and Chopra. R. N. et al., 1986). Argemone maxicana has been investigated in terms of modern pharmacology for its anti-malarial activity (Merlin L. et al., 2007, Bertrand Graz et al., 2010, M. Sakthivadivel et al., 2003.), Molluscicidal and nematocidal activity (Sushma Singh, 1999, S. Singh 2000), Anticancer activity (Chang, Y.-C, 2003), Anti-bacterial, antifungal and antimicrobial activity (Bhattacharrjee I., 2006, Satish S. 2007, Siddiqui I. A., 2002, A. Osho, 2010.), Hepatoprotective activity (Das P. K. et al., 2009), Antistress and antiallergic activity (Bhalke et al., 2009), Anti-cataleptic activity (Bhalke et al., 2009)., Anti-HIV activity (Chang et al., 2003), Neuropharmcological study (Capasso A. et al., 2002). In the present study A. mexicana was evaluated for its analgesic, anxiolytic and sedative activities.

### MATERIAL AND METHOD

#### Plant material and preparation of extracts

The whole *Argemone mexicana* plant was collected in June 2009 from Ahmednagar district, Maharashtra (India). The plant specimen was authenticated from Botanical Survey of India, Pune (Voucher specimen no. YMA1). Plant material was dried under shade and coarsely powdered for extraction. The coarsely powdered whole plant (500g) of *Argemone mexicana* was separately subjected to extraction using ethyl acetate and methanol for 10 days by cold maceration. The methanolic and ethyl acetate extracts were concentrated by rotary vacuum evaporator under reduced pressure and then dried in open air.

#### Animals

Healthy wistar albino mice of either sex and of approximately the same age, weighing about 30-40 g were used for study. All the mice were housed in polypropylene cages maintained under standard condition (12 h light/12 h dark cycle). All the animals were carefully monitored and maintained in accordance with CPCSEA guidelines on control and supervision of experimental animals. The ethical clearance was obtained from the Institutional Animal Ethics Committee before the experiment.

#### ANALGESIC ACTIVITY

#### Hot plate test

Central analgesic activity for *A. mexicana* whole plant was evaluated using hot plate method as per described by Woolfe and MacDonald. Albino male mice were divided into eight groups containing six animals each.

Group-I- control (received only vehicle) Group-II -standard drug pentazocine (50 mg/kg, i.p.) Group-III- 100 mg/kg b.w. methanolic extract Group IV-200 mg/kg b.w. methanolic extract Group V- 400 mg/kg b.w. methanolic extract Group VI-100 mg/kg b.w. ethyl acetate extract Group VII- 200 mg/kg b.w. ethyl acetate extract Group VIII- 400 mg/kg b.w. ethyl acetate extract

Individual mice was placed on the hot plate which was maintained at 55  $^{0}$ C  $\pm$  1  $^{0}$ C, and latency of nociceptive response such as jumping, flicking of a hind limb and licking was noted. And after administration of extracts, the readings were recorded at 0, 30, 60, 90, 120, 150 and 180 minutes time interval. The experiment was terminated by 20 second after their placement on the hot plate to avoid damage to the paws of animals.

#### Writhing test

Peripheral analgesic activity for *A. mexicana* was evaluated using writhing test (Koster R et al., 1959). Albino male mice (30-40 g) were grouped into eight groups of six animals each.

Group-I- control (received only vehicle) Group-II -standard drug paracetamol (50 mg/kg, i.p.) Group-III- 100 mg/kg b.w. methanolic extract Group IV-200 mg/kg b.w. methanolic extract Group V- 400 mg/kg b.w. methanolic extract Group VI-100 mg/kg b.w. ethyl acetate extract Group VII- 200 mg/kg b.w. ethyl acetate extract Group VIII- 400 mg/kg b.w. ethyl acetate extract

Before administration of acetic acid, extracts and standard drug, each mice was placed in the glass beakers. 0.1 ml of 0.6 % of acetic acid solution was injected intraperitoneally to each animal. Then the animals were allowed to elapse for 5 minutes, after administration of acetic acid and number of writhes was recorded individually to each animal for the period of 30 minutes.

#### Locomotor activity

The healthy adult albino mice (30-40 g) were firstly divided into eight groups containing six animals each.

Group-I- control (received only vehicle) Group-II -standard drug diazepam (4 mg/kg i.p.) Group-III- 100 mg/kg b.w. methanolic extract Group-IV-200 mg/kg b.w. methanolic extract Group-V- 400 mg/kg b.w. methanolic extract Group-VI-100 mg/kg b.w. ethyl acetate extract Group-VII- 200 mg/kg b.w. ethyl acetate extract Group-VIII- 400 mg/kg b.w. ethyl acetate extract

After thirty minutes of standard drug and extract administration, each animal was placed in photoactometer individually and then the locomotor activity was counted for 10 minutes (Dewan S, 2000, Amos s, 2001).

#### Motor coordination (Muscle relaxant activity)

For the assessment of the motor co-ordination test, albino mice (30-40 g) were divided into eight groups containing six animals each group.

Group-I- control (received only vehicle) Group-II -standard drug diazepam (2 mg/kg, i.p.) Group-III- 100 mg/kg b.w. methanolic extract Group-IV-200 mg/kg b.w. methanolic extract Group-V- 400 mg/kg b.w. methanolic extract Group-VI-100 mg/kg b.w. ethyl acetate extract Group-VII- 200 mg/kg b.w. ethyl acetate extract Group-VIII- 400 mg/kg b.w. ethyl acetate extract

Rota-rod device was used for the assessment of the experiment. The mice were positioned on rota-rod which was set at a rate of 16 revolutions per minutes. The fall off time for each animal was noted. The difference in the fall off time from the rotating rod was noted at the interval of 30 minutes for 3h (Ozturk et al., 1996; Perez et al., 1998).

#### Effect on Phenobarbitone sodium sleep

For the evaluation of phenobarbitone sodium induced sleeping test, albino mice (30-40 g) were divided into eight groups containing six animals each group.

Groun	Treatment						
Group	Treatment	0min	30min	60min	120min	150min	180min
Ι	Control (Normal	5.29±	$5.3 \pm 0.0208$	$4.96 \pm 0.0409$	$5.09 \pm 0.1217$	$5.35 \pm 0.1590$	$5.48 \pm 0.1350$
	saline)	0.0288					
II	Pentazocin (50	$5.42 \pm$	$9.45 \pm 0.1198$	13.96±	$21.29 \pm 0.0466$	$19.42 \pm 0.0556$	16.01±
	mg/kg)	0.0321		0.0288			0.0176
III	ME-100mg/kg	$5.04 \pm$	6.21±	$8.22\pm$	11.06±	$10.42 \pm 0.0829^{*a}$	9.44±
		0.0240	$0.0832^{*^{a}}$	$0.0721^{*a}$	$0.0176^{*a}$		$0.0240^{*a}$
IV	ME-200mg/kg	4.91±	7.30±	9.36±	13.29±	$12.34{\pm}0.0819^{*^a}$	10.31±
		0.0371	$0.1167^{*^{a}}$	$0.0480^{*a}$	$0.0864^{*^{a}}$		$0.0887^{*^{a}}$
V	ME- 400mg/kg	$5.2 \pm 0.0208$	7.76±	$10.34 \pm$	$15.44 \pm$	$13.46 \pm 0.0866^{*a}$	11.57±
			$0.1490^{*a}$	$0.0744^{*^{a}}$	$0.1365^{*a}$		$0.0929^{*^{a}}$
VI	EA-100mg/kg	$4.55\pm$	$5.94\pm$	$7.35\pm$	$10.55 \pm$	$9.08 \pm 0.0296^{*a}$	$8.25\pm$
		0.1286	0.0230* <sup>a</sup>	$0.0768^{*a}$	$0.1322^{*a}$		$0.0584^{*a}$
VII	EA-200mg/kg	4.91±	$6.24 \pm$	$8.44\pm$	12.43±	$10.17 \pm 0.1200^{*a}$	$9.48 \pm$
		0.0115	$0.0693^{*a}$	$0.0763^{*a}$	$0.0448^{*^{a}}$		$0.1110^{*a}$
VIII	EA-400mg/kg	$5.07\pm$	7.31±	9.61±	13.42±	$11.32 \pm 0.0545^{*a}$	$9.08\pm$
		0.0176	$0.1185^{*a}$	$0.1015^{*a}$	0.0731* <sup>a</sup>		$0.0384^{*a}$

Table no. 1 Analgesic activity of methanolic and ethyl acetate extracts of A. mexicana by Eddy's hot plate method: Mean reaction time in minutes (min +SEM)

Results are expressed as  $\pm$  SEM (n=6). Data processed by one way ANOVA followed by Dunnett's test, \* p<0.01 significant when compared to control group. <sup>a</sup>p <0.01 significant when compared to standard group.

Group-I- control (received only vehicle) Group-II -standard drug diazepam (1 mg/kg, i.p.) Group-III- 100 mg/kg b.w. methanolic extract Group-IV-200 mg/kg b.w. methanolic extract Group-V- 400 mg/kg b.w. methanolic extract Group-VI-100 mg/kg b.w. ethyl acetate extract Group-VII- 200 mg/kg b.w. ethyl acetate extract Group-VIII- 400 mg/kg b.w. ethyl acetate extract

Phenobarbitone sodium (40 mg/kg b.w.) was administered to individual animal later than 30 minutes of extract and standard drug administration. And all the animals were observed for onset of sleep and extent of sleep with condition being loss of righting reflex (Wambebe, 1985; Rolland et al., 1991).

#### Statistical analysis

Statistical analysis was performed using Graph Pad prism 3. All the results were expressed as mean  $\pm$  standard error of mean (S.E.M.) and analyzed for ANOVA and post hoc Dunnet's *t*-test (Multiple). Differences between groups were considered significant at p<0.05, p<0.01 levels.

#### **RESULT AND DISCUSSION**

Acute toxicity studies for methanolic and ethyl acetate extract of *A. mexicana* showed no mortality till doses of 2.5 g/kg b.w. even after 72 h. All mice were compared to normal animals for their awareness, vocalization, tremors, righting reflex, pain, salivation and lacrimation.

In present study methanolic and ethyl acetate extract of *A. mexicana* whole plant was studied for CNS depressant activity at doses of 100 mg/kg, 200 mg/kg and 400 mg/kg b.w. with various animal models such as analgesic activity by hot plate method and acetic acid induced writhing test, locomotor activity, motor coordination (muscle relaxant activity) and phenobarbitone induced sleeping time. In the assessment of analgesic activity, methanolic extract and ethyl acetate extracts of *A. mexicana* showed significant central analgesic activity (Table 1) at dose of 200 mg/kg as compare to control (p<0.01) and standard pentazocine (p<0.01). For peripheral analgesic activity percentage inhibition for ethyl acetate extract and methanol extract at dose 200 mg/kg b.w. was found to be 53.34 % and 52.84 %. And ethyl acetate extract was found to be more potent as compare to methanol extract. But both extracts showed significant peripheral activity when results were compared to control (p<0.01) and standard paracetamol (p<0.01). As methanolic and ethyl acetate extracts of *A. mexicana* also showed remarkably locomotor activity on photoactometer (Table 3), amongs both extracts methanolic extract showed significant reduction in motor activity at doses of 200 mg/kg, when results are compared to control (p<0.01) and standard diazepam (p<0.01).

Table no. 2 Analgesic activity of methanolic and ethyl acetate extracts of A. mexicana by acetic acid induced method

Group	Treatment	Number of writhing	Percent inhibition
Ι	Control	$67.80 \pm 0.1590$	-
	(Normal saline)		
II	Paracetamol	$20.04 \pm 0.0763$	70.44
	(50 mg/kg)		
III	ME-100mg/kg	$43.14 \pm 0.0550$ * <sup>a</sup>	36.37
IV	ME-200mg/kg	$31.97 \pm 0.0437 * a$	52.84
1.4	WIE-200Ing/Kg	51.97 ± 0.0457	52.04
V	ME- 400mg/kg	$30.10\pm 0.0176^{\asta}$	55.60
VI	EA 100	$41.21 + 0.0560 \pm a$	20.07
VI	EA-100mg/kg	$41.31 \pm 0.0369$ *	39.07
VII	EA-200mg/kg	31.63 ± 0.1212* <sup>a</sup>	53.34
	5 6		
VIII	EA-400mg/kg	$29.27 \pm 0.0841$ * <sup>a</sup>	56.82

Results are expressed as  $\pm$  SEM (n=6). Data processed by one way ANOVA followed by Dunnett's test, \* p<0.01 significant when compared to standard group. <sup>a</sup>p <0.01 significant when compared to control group.

In the assessment of muscle relaxant test, methanolic and ethyl acetate extracts of *A. mexicana* showed significant reduction in fall off time of animals (sec) on rota rod (Table no. 4) when results were compared to control group (p<0.01) and standard diazepam (p<0.01) and in case of phenobarbitone induced sleeping test, methanolic extract at dose of 200 mg/kg b.w. significantly potentiate phenobarbitone induced sleeping time (Table 5) as compared to control (p<0.01) and standard (p<0.01).

Table no. 3 Effect of methanolic and ethyl acetate extracts of A. mexicana on locomotor activity

~		Locomotor activity observed for 10 min			
Group	Treatment	Before dosing	After dosing		
Ι	Control (Normal saline)	$178.10 \pm 0.0726$	$178.37 {\pm} 0.3327$		
II	Diazepam (4 mg/kg)	$177.54 \pm 0.1374$	$83.97 \pm 0.1332$		
III	ME-100mg/kg	$178.24 {\pm} 0.1593$	$128.43 \pm 0.0266 *$		
IV	ME-200mg/kg	$177.65 \pm 0.1202$	$110.32\pm 0.0523^{\ast a}$		
V	ME- 400mg/kg	$177.76 \pm 0.0700$	$108.46 \pm 0.0796^{\ast^a}$		
VI	EA-100mg/kg	$177.65 {\pm} 0.0491$	$132.39 \pm 0.1313*$		
VII	EA-200mg/kg	$178.3 \pm 0.06506$	$113.50\pm 0.0617 \ast^a$		
VIII	EA-400mg/kg	$178.54 {\pm} 0.1378$	$110.45 \pm 0.2245 \ast^{\rm a}$		

Results are expressed as  $\pm$  SEM (n=6). Data processed by one way ANOVA followed by Dunnett's test, \* p<0.01 significant when compare control group. <sup>a</sup>p <0.01 significant when compared to standard group.

As remarkable analgesic activity of methanolic extract at dose 200 mg/kg b.w. was seen in Eddy's hot plate method and acetic acid induced writhings. The hot plate test is widely used for assessment of central antinociceptive activities, having tendency to respond to the pain stimuli through neuronal pathways (Chapman et al., 1985; Morales et al., 2001). In general, acetic acid writhing test is used for evaluation of peripheral antinociceptive activities, as writhing test is useful to differentiate between central and peripheral nociception (Le Bars et al., 2001) and acetic acid injection induces peritoneal inflammation, which may triggers a response characterized by writhing (Koster et al., 1959). This study demonstrated that acetic acid indirectly induces endogenous release of pain mediators (such as prostaglandins, kinins, histamin, etc.) that stimulate the nociceptive neurons, which are responsive to non- steroidal anti-inflammatory drugs and opioids (Derardt et al., 1980; Sánchez-Mateo et al., 2006; Sulaiman et al., 2008).

The locomotor activity was also assessed for methanolic and ethyl acetate extract by actophotometer but methanolic extract was found to be more potent. As it showed decrease in locomotion and grip strength on rota rod was also decreased (Leewanich et al., 1996) which reveals the CNS depressant activity. The CNS depression may be due to the increase in the GABA concentration in brain (Nagarjun et al., 2003). The pentobarbitone-induced sleep test, also significantly showed sedative effect for methanolic and ethyl acetate extracts but methanolic extract was found to be potent.

As preliminary Phytochemical investigation showed the presence of phenolics, flavonoids, steroids, alkaloids and tannins, which may be responsible for the CNS depressant activity. The above studies indicate that the methanolic extract of *A. mexicana* whole plant possesses analgesic, sedative, and anxiolytic activity.

## REFERENCE

Amos S., Kolawole E., Akah P., Wambebe C., Gamaniel K., Behavioural effects of the aqueous extract of *Guiera senegalensis* in mice and rats. Phytomed, 2001; 8: 356–361.

Bertrand Graz, Merlin L. Willcox, Chiaka Diakite, Jacques Falquet, Florent Dackuo, Oumar Sidibe, Sergio Giani, Drissa Diallo. *Argemone mexicana* decoction versus artesunateamodiaquine for the management of malaria in Mali: policy and public-health implications. Transactions of the Royal Society of Tropical Medicine and Hygiene, 2010; 104(1): 33-41.

Bhattacharjee I., Chatterjee S. K., Chatterjee S., Chandra G. Antibacterial potentiality of Argemone mexicana solvent extracts against some pathogenic bacteria. Mem Inst Oswaldo Cruz., 2006; 101(6): 645-8.

Table no. 4 Effect of methanolic and ethyl acetate extracts of A. mexicana on muscle relaxant activity.

~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	Treatment	Fall of time in seconds (sec ±SEM)					
Group		0min	30min	60min	120min	150min	180min
Ι	Control (Normal saline)	35.52± 0.0470	35.71± 0.1595	$35.76 \pm 0.2084$	$35.4{\pm}~0.1595$	$35.51 \pm 0.0560$	$35.72 \pm 0.1757$
II	Diazepam (2 mg/kg)	36.17± 0.0578	$28.21 \pm 0.0993$	$19.15 {\pm} 0.0458$	$14.06 \pm 0.0264$	$20.16 \pm 0.0260$	$25.25 \pm 0.1115$
III	ME-100mg/kg	35.56± 0.1081	$33.14 \pm 0.0352^{*a}$	$31.35\pm 0.1058*^a$	$28 \pm 0.0592 *^{a}$	$29.33 {\pm}~ 0.0393 {*}^a$	$27.42\pm \ 0.0602^{*^a}$
IV	ME-200mg/kg	35.71± 0.0832	$32.13 \pm 0.0688^{*a}$	$27.32{\pm}0.0635{*}^a$	$23.25{\pm}0.0416{*^a}$	$25.45{\pm}0.1868{*}^a$	$26.80 \pm 0.1073^{*a}$
V	ME- 400mg/kg	35.99± 0.0554	$31.29{\pm}0.0617{*^a}$	$25.58 {\pm} 0.1418^{*^a}$	$21.35{\pm}0.0433{*}^a$	$23.58{\pm}0.2098{*}^a$	$26.24{\pm}0.0819^{*^a}$
VI	EA-100mg/kg	36.36± 0.0480	$33.98 \pm 0.0173^{*a}$	$32.13{\pm}0.0208{*^a}$	$29.38 \pm 0.0360 *$	$30.31{\pm}0.0845{*}^a$	$31.35 \pm 0.0674 *$
VII	EA-200mg/kg	$35.31\pm 0.0536$	$33.03 {\pm} 0.0321 {*}^a$	$28.33 {\pm} 0.1475{*}^a$	$24.46{\pm}0.0786{*^a}$	$25.08{\pm}0.0556{*}^a$	$27.31{\pm}0.2488{*}^a$
VIII	EA-400mg/kg	35.18± 0.1012	$32.08 \pm 0.0425^{*a}$	$27.32 \pm 0.1281^{*a}$	$23.02 \pm 0.0409^{*a}$	24.15±s 0.0328* <sup>a</sup>	$27.32 \pm 0.2136^{*a}$

Results are expressed as  $\pm$  SEM (n=6). Data processed by one way ANOVA followed by Dunnett's test, \* p<0.01 significant when compared to standard group. <sup>a</sup>p <0.01 significant when compared to control group.

Table no. 5 Effect of methanolic and ethyl acetate extracts of *A. mexicana* on phenobarbitone induced sleeping time.

Group	Treatment	Mean sleeping time in min.
I	Control (Normal saline)	$23.36 \pm 0.05000$
II	Phenobarbitone (40 mg/kg)	$74.53 \pm 0.2661$
III	ME-100mg/kg	$35.22 \pm 0.08988*$
IV	ME-200mg/kg	$43.41 \pm 0.1368 {*}^a$
V	ME- 400mg/kg	$48.58 \pm 0.0696 \ast^a$
VI	EA-100mg/kg	$37.25 \pm 0.1146*$
VII	EA-200mg/kg	$45.46 \pm 0.1375^{*a}$
VIII	EA-400mg/kg	$47.33 \pm 0.0783^{*^a}$

Results are expressed as  $\pm$  SEM (n=6). Data processed by one way ANOVA followed by Dunnett's test, \* p<0.01 significant when compared to control group. <sup>a</sup>p <0.01 significant when compared to standard group.

Capasso A., Aquino R., Tommasi N. De, Piacente S., Rastrelli L., Pizza C. Neuropharmacology Activity of Alkaloids from South American Medicinal Plants. Curr. Medi. Chem.- CNS agents, 2002; 2(1): 1-15.

Chang Yuh-Chwen, Hsieh Pei-Wen, Chang Fang-Rong, Wu Ru-Rong, Liaw Chih-Chuang, Lee Kuo-Hsiung, Wu Yang-Chang. Two new protopines argemexicaines A and B and the anti-HIV alkaloid 6-acetonyldihydrochelerythrine from Formosan *Argemone Mexicana*. Planta medica, 2003; 69(2):148-152.

Chang, Y.-C., Chang, F-R., Khalil A. T., Hsieh, P.-W. and Wu, Y.-C., "Cytotoxic benzophenanthridine and benzylisoquinoline alkaloids from *Argemone mexicana*". *Z.* Naturforsch, 2003; 58c: 521-526.

Chapman C. R., Casey K. L., Dubner R., Foley K. M., Gracely R. H., Reading, A.E. Pain measurement: an overview. Pain, 1985; 22: 1–31.

Chevallier A. The Encyclopedia of Medicinal Plants Dorling Kindersley. London An excellent guide to over 500 of the more well known medicinal herbs from around the world. (1996).

Chopra R. N., Nayar. S. L. and Chopra. I. C. Glossary of Indian Medicinal Plants (Including the Supplement). Council of Scientific and Industrial Research, New Delhi. (1986).

Das P. K., Prasanna Panda, Somya Ranjan Pani, Ranjan Sethi. Hepatoprotective activity of plant *Moringa pterygosperma* against carbon tetrachloride (ccl4) induced hepatopathy in rats. I J P R D, (2009); 1(8): 002.

Derardt R., Jougney S., Benzoni J., Peterfalvi M. Release of prostaglandins E and F in an algogenic reaction and its inhibition. Euro. J. Pharmacol., 1980; 61: 17–24.

Dewan S., Sangraula H., Kumar V. L., Preliminary studies on the analgesic activity of latex of *Calotropis procera*. J. Ethnopharmacol., 2000; 73 (1-2): 307–311.

Koster R., Anderson M., De Beer E. J. Acetic acid for analgesic screening. Fed. Proc., 1959; 18: 412.

Le Bars D., Gozariu M., Cadden S. Animal models of nociception. Pharmacol. Rev., 2001; 53: 628–651.

Leewanich P., Tohda M., Matsumoto K., Subhadhirsakul S., Takayama H., Watanbe H. Behavioural studies on alkaloids extracted from leaves of *Hunteria zeylanica*. Biol. Pharm. Bull., 1996; 19: 394–399.

M. Sakthivadivel, D. Thilagavathy. Larvicidal and chemosterilant activity of the acetone fraction of petroleum ether extract from *Argemone mexicana* L. seed. Biores. Tech, 2003; 89(2): 213-216.

Merlin L. Willcox, Bertrand Graz, Jacques Falquet, Oumar Sidibé, Mathieu Forster, Drissa Diallo. *Argemone mexicana* decoction for the treatment of uncomplicated falciparum malaria. Transactions of the Royal Society of Tropical Medicine and Hygiene, 2007; 101(12): 1190-1198.

Morales L., Perez-Garcia C., Alguacil, L. F. Effects of yohimbine on the antinoci- ceptive and place conditioning effects of opioid agonists in rodents. Brit. J. Pharmacol., 2001; 133: 172–178.

Nagarjun N. S., Soundari P. G., Kumaresan P.T. CNS depressant activity of *Dalbergia malaberica*. Ind. Drugs 2003; 40: 716–717.

Osho and T. Adetunji. Antimicrobial activity of the essential oil of *Argemone mexicana* Linn. J. of Med. Plants Res, 2010; 4 (1): 019–022.

Ozturk Y., Aydine S., Baser K. H. C., Berberoglu H. Effects of *Hypericum perforatum* L. and *Hypericum calycinum* L. Extracts on the central nervous system in mice. Phytomed, 1996; 3: 139–146.

Perez G. R. M., Perez L. J. A., Garcia D. L. M., Sossa M. H. Neuropharmacological activity of *Solanum nigrum* fruit. J. of Ethnopharmacol, 1998; 62: 43–48.

R. D. Bhalke, S. A. Gosavi. Anti-Stress and Antiallergic Effect of *Argemone Mexicana* Stems in Asthma. Arch Pharm Sci & Res 2009; 1 (2): 127-129.

Rastogi R. P. and Mehotra B. N. Compendium of Indian Medicinal Plants, Vol. II, CDRI, Lucknow, 1979; 446.

Rolland A., Fleurentain J., Lanhers M., Younos C., Misslin R., Morier F. Behavioural effects of American traditional plant *Eschscholzia califormica*; sedative and anxiolytic properties. Planta Medica, 1991; 57: 212–216.

S. Satish, D. C. Mohana, M. P. Raghavendra and K. A. Raveesha. Antifungal activity of some plant extracts against important seed borne pathogens of *Aspergillus* sp. J. Agri. Tech, 2007: 109-119.

S. Singh and D. K. Singh. Effect of Active Molluscicidal Componenets Of Abrus Precatorius, Argemone Mexicana And Nerium Indicum On Certain Enzymes In The Nervous Tissue Of Lymnaea Acuminata. J. Sci. I. R. Iran., 2000; 11(3): 187-194.

Sánchez-Mateo, C. C., Bonkanka, C. X., Hernández-Pérez, M., Rabanal R. M. Evaluation of the analgesic and topical anti-inflammatory effects of Hypericum reflexum L. fil. J. of Ethnopharmacol., 2006; 107: 1–6.

Siddiqui I. A., Shaukat S. S., Khan G. H., Zaki M. J. Evaluation of *Argemone mexicana* for control of root-infecting fungi in potato. J. Phytopathol. 2002; 150: 321-329.

Sulaiman M. R., Zakaria Z. ., Bujarimin A. S., Somchit M. N., Israf D. A., Moin S., 2008. Evaluation of Moringa oleifera aqueous extract for antinociceptive and anti- inflammatory activities in animal models. Pharma. Biol., 2008; 46: 838–845.

Sushma Singh, D. K. Singh. Molluscicidal activity of Abrus precatorius linn. And Argemone maxicana linn. Chemosphere, 1999; 38(14): 3319-3328.

Wambebe C. Influence of some agents that affect 5-HT metabolism and receptors and nitrazepam-induced sleep in mice. British J. of Pharmacol, 1985; 84:185–191.

Willcox M. L., Graz B., Falquet J., *et al.* "Argemone mexicana decoction for the treatment of uncomplicated falciparum malaria". Trans R Soc Trop Med Hyg, 2007; 101: 1190–1198.

Woolfe G. and MacDonald A. D. The evaluation of the analgesic action of pethidine hydrochloride (Dermol). J. Pharmacol. Exp. Ther., 1944; 80: 300-330.