Pharmacognostic, Physicochemical Standardization and Phytochemical Analysis of leaves of Cultivated Crotalaria lachnosema Stapf.

Jemilat A. Ibrahim1*, Opeoluwa Makinde1 and Nneka N. Ibekwe2
1Department of Medicinal Plant Research and Traditional Medicine, National Institute for Pharmaceutical Research and Development (NIPRD), PMB 21, Garki, Abuja.
2Department of Medicinal Chemistry and Quality Control, National Institute for Pharmaceutical Research and Development (NIPRD), PMB 21, Garki, Abuja.

ARTICLE INFO
Article history:
Received on: 22/08/2012
Revised on: 08/09/2012
Accepted on: 15/09/2012
Available online: 28/09/2012

Key words:
Authentication, chromatograph, Macroscopic, Microscopic

ABSTRACT
Towards authentication and quality assurance of medicinal plants, pharmacognostic, physicochemical and preliminary phytochemical studies of the leaves of Crotalaria lachnosema Stapf. were carried out. The macroscopic and microscopic evaluation revealed characters that are of diagnostic value and useful in authentication of the plant. The Physicochemical analyses reveals values for moisture content, alcohol extractive, water extractive and total ash which are within the World Health Organisation (WHO) standards for crude drug from medicinal plants. Phyto-screening for secondary metabolites revealed the presence of saponins, terpenes/steroids, flavonoids, resins and balsams, while alkaloids, glycosides and tannins were absent. Information obtained from these studies can be used as markers in the identification and standardization of this plant as a herbal remedy and also towards monograph development on the plant.

INTRODUCTION
The plant Crotalaria lachnosema Stapf. belongs to the family Fabaceae (Leguminosae), subfamily Papilionoideae. It is a woody plant with a height of about 2m high. It is found along forest margin in damp sites (Burkill, 1995). The plant is known as ‘Fara bi – rana’ in Hausa, ‘Kompo’ in Yoruba, ‘Ake dinwo’ in Igbo and ‘Birijibei’ in Fulani (Nuhu et al, 2009). In a study carried out on the traditional medicinal uses of Crotalaria species, Crotalaria lachnosema was found to be important in the treatment of scabies. It was also found to be highly sorted after in love matters and acceptance (Nuhu et al., 2009). In the same study, it is said that the whole plant grounded and mix with water fed to animals to treat liver diseases (Nuhu et al., 2009). Pyrrolizidine alkaloids, a poisonous compound has been found to be present in Crotalaria spp and according to WHO report, many species of the genus have been reported to be toxic with epidemic outbreaks in some part of the world (WHO, 1988; Arzt and Mount, 1999). Bras et al. (1961) stated that only few species of the genus Crotalaria have been studied and also that WHO have labelled the Pyrrolizidine alkaloids as a very important toxicant and have made a call that all species of plants that might contain the alkaloid should be studied ethnomedicinally, phytochemically and taxonomically, etc (WHO,1988). Despite the medicinal importance of this species and its likely danger because of the presence of the Pyrrolizidine alkaloids, information on the pharmacognostic parameters for identification of this species in whole and powdered form are unavailable. The present study aimed at investigating the
macromorphology, pharmacognostic evaluation and phytochemical screening of the leaves of *Crotalaria lachnosema* towards standardization and monograph development.

**MATERIALS AND METHODS**

**Sample collection and processing**

The plant was collected from Jos in Plateau state of Nigeria in the year 2009 and the seeds were planted in the botanical garden of National Institute for Pharmaceutical Research and Development (NIPRD) Idu, Abuja. The plant material used for this study was collected in the year 2010 from the botanical garden of the Institute. The plant was air dried indoor and powdered using mortar and pestle. The powdered sample was stored in air tight container for the phytochemical analysis.

**Pharmacognostic analysis**

**Macromorphology**

Macromorphological characters of the leaves like leaf shape, size, colour, texture, margin type, apex, base and petiolo size, flower colour and length etc were observed. Measurements were carried out using line ruler.

**Microscopic analysis**

Microscopic analysis was carried out on epidermal layers and transverse sections of leaf.

**Epidermal layers of leaves**

Leaves of the specimens were cut at the median portions. These were soaked in concentrated Nitric acid for about 24hrs. The appearance of air bubbles indicated the readiness of the epidermises to be separated. The samples were then transferred to Petri dishes containing water and with the use of fine forceps and dissecting needle; the upper and lower epidermises were separated. One set was stained with saffranin and another one with Sudan IV and later mounted on a slide with glycerol. The edges of the cover slip were sealed with nail vanish to prevent dehydration.

**Transverse section of leaves**

Sections were manually obtained by sectioning with razor blade. The sections were cleared for some minutes in sodium hypochlorite solution. It was washed in water and then stained with Sudan IV and later mounted on a slide with glycerol. The edges of the cover slip were sealed with nail vanish to prevent dehydration.

**Physicochemical Analysis**

Physicochemical analyses were carried out on the powdered sample following standard methods (Sofowora, 2008; Evans, 2002; African Pharmacopeia, 1986). Moisture content, alcohol extractive value, water extractive value and total ash value were tested for.

**Phytochemical screening**

Preliminary phytochemical investigations for secondary metabolites were carried out on the powdered sample of the leaves of *C. lachnosema* using standard procedures (Sofowora, 2008; Evans, 2002). The metabolites tested for were carbohydrates, balsams, resins, saponins, sterols, terpenes, tannins, flavonoids, cardiac glycosides, glycosides, alkaloids and phlobatannins.

**Thin layer chromatography**

Thin-layer chromatography (TLC) was employed in the qualitative analysis of organic extracts of the powdered leaves. The plant material was extracted successively with hexane (Hex), ethyl acetate (EtOAc) and methanol (Meth) between intervals of 24 hours. The extracts were spotted on activated silica gel plates. Two solvent systems of Hex-EtOAc (9-1) and Hex-EtOAc-Meth (7-2-1) were used for development of the plates. Spots were detected on TLC plates by spraying with sulphuric acid, followed by charring at 110 °C for 10 minutes in an oven. The retention factor (Rf) for each spot was calculated using the formula:

$$R_f = \frac{\text{Distance moved by solute}}{\text{Distance moved by solvent}}$$

**RESULTS**

**Pharmacognostic analysis**

**Macroscopical evaluation**

*C. lachenosema* is a shrub, whole plant hairy. Stem densely brownish hairy and round in shape. Leaves thick and densely hairy on both surfaces, greenish in colour. Leaf venation more prominent on lower surface with submarginal nerves. Leaf is compound and alternate in arrangement with stipules. Petiole about 0.6 – 1 cm long. Leaflets are trifoliate and oblanceolate to oblong in shape, 3.7 – 6.7cm x 1.5 – 2.5cm. Petiolule about 0.3cm long. Middle leaflet is largest. Leaf apex slightly mucronate, base cuneate with entire margine. Flowers orange in colour. Fruit a legume, densely hairy, brownish in colour, about 1.5cm to 2.5cm long, almost oblong in shape (Figure 1).

**Fig. 1: Pictures of Crotalaria lachenosema Stapf. a: whole plant; b: matured flower; c: matured fruits.**

---

068 Ibrahim et al. / Journal of Applied Pharmaceutical Science 2 (09); 2012: 067-070
**Microscopic evaluation of leaf**

*Upper epidermis*

The upper epidermis shows abundant long unicellular unbranched trichomes. Trichomes bases are also seen abundant on the surface. Epidermal cells are irregular to polygonal in shape with abundant oil globules on the surface (Plate 1).

**Plate 1:** Adaxial epidermal layer of *Crotalaria lachenosema* Stapf. a: abundant trichomes on the surface; b: polygonal to irregular cell shape and abundant trichome bases; c: abundant oil globules (arrowed).

*Lower epidermis*

The lower epidermal surface also shows abundant long unicellular unbranched trichomes and also abundant trichome bases. Epidermal cells are irregular to polygonal in shape. Anisocytic stomata type abundant while few anomocytic stomata type was observed. Epidermal cells were shortly and randomly striated (Plate 2).

**Plate 2:** Abaxial (lower) epidermal surface of *Crotalaria lachenosema* Stapf. a: irregular cell shape and abundant stomata; b: abundant anisocytic and anomocytic stomata types, abundant oil globules.

**Transverse section of leaf**

Transverse section shows isobilateral leaf type with one layer of palisade cells on one surface and two layers of palisade cell on the other surface (Plate 3b & c). Thick Midrib protruding abaxially (lower surface) while a slight groove is observed adaxially (Upper surface) (Plate 3a). Trichomes abundant on both layers of the sections (Plate 3a). Abundant oil globules are seen on the palisade, mesophyll and epidermal cells. Spiral xylem vessels also observed.

**Physicochemical analyses**

The physicochemical parameters are presented in Table 1.

**Table 1:** Physicochemical parameters of the leaves of *Crotolaria lachenosema* Stapf.

<table>
<thead>
<tr>
<th>Test</th>
<th>Results %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Moisture content</td>
<td>13.93</td>
</tr>
<tr>
<td>Alcohol extractive value</td>
<td>2.16</td>
</tr>
<tr>
<td>Water extractive value</td>
<td>16.73</td>
</tr>
<tr>
<td>Total ash value</td>
<td>6.40</td>
</tr>
</tbody>
</table>

**Phytochemical analyses**

The preliminary phytochemical screening revealed the presence of saponins, terpenes, sterols, flavonoids, resins and balsams while tannins, cardiac glycoside, alkaloids, phlobatannins and glycosides were absent (Table 2).

**Table 2:** Phytochemical screening of the powdered leaf of *Crotolaria lachenosema* Stapf.

<table>
<thead>
<tr>
<th>TEST</th>
<th>INFERENCE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tannins</td>
<td>-</td>
</tr>
<tr>
<td>Saponins</td>
<td>+</td>
</tr>
<tr>
<td>Cardiac glycosides</td>
<td>-</td>
</tr>
<tr>
<td>Terpenes</td>
<td>+</td>
</tr>
<tr>
<td>Sterols</td>
<td>+</td>
</tr>
<tr>
<td>Flavonoids</td>
<td>+</td>
</tr>
<tr>
<td>Resins</td>
<td>+</td>
</tr>
<tr>
<td>Balsams</td>
<td>+</td>
</tr>
<tr>
<td>Alkaloids</td>
<td>-</td>
</tr>
<tr>
<td>Phlobatannins</td>
<td>-</td>
</tr>
<tr>
<td>Glycosides</td>
<td>-</td>
</tr>
</tbody>
</table>

Key:  
+ = positive  
- = negative

**Thin layer chromatography**

The *Rf* values of spots detected on TLC analysis of the three extracts; hexane, ethylacetate and methanol developed in the solvent system of Hex: EtOAc (9:1) is presented in Table 3, while the *Rf* values of the spots of the extracts developed in the solvent system of Hex: EtOAc: Meth (7:2:1) is presented in Table 4.

**Table 3:** Retension factors (*Rf*) of components of organic extracts of the leaves of *Crotalarialachenosema* in solvent system hex- EtOAc (9:1).

<table>
<thead>
<tr>
<th>Hexane extract (cm)</th>
<th>Colour</th>
<th>Ethylacetate extract (cm)</th>
<th>Colour</th>
<th>Methanol extract (cm)</th>
<th>Colour</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.65</td>
<td>Brown</td>
<td>0.64</td>
<td>Purple</td>
<td>0.40</td>
<td>Brown</td>
</tr>
<tr>
<td>0.40</td>
<td>Purple</td>
<td>0.40</td>
<td>Brown</td>
<td>0.40</td>
<td>Brown</td>
</tr>
<tr>
<td>0.26</td>
<td>Purple</td>
<td>0.27</td>
<td>Purple</td>
<td>0.44</td>
<td>Brown</td>
</tr>
<tr>
<td>0.25</td>
<td>purple</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Table 4:** Retension factors (*Rf*) of components of organic extracts of the leaves of *Crotalarialachenosema* in solvent system hex- EtOAc: Meth (7:2:1).

<table>
<thead>
<tr>
<th>Hexane extract (cm)</th>
<th>Colour</th>
<th>Ethylacetate extract (cm)</th>
<th>Colour</th>
<th>Methanol extract (cm)</th>
<th>Colour</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.89</td>
<td>Brown</td>
<td>0.46</td>
<td>Brown</td>
<td>0.69</td>
<td>Brown</td>
</tr>
<tr>
<td>0.68</td>
<td>Purple</td>
<td>0.46</td>
<td>Brown</td>
<td>0.44</td>
<td>Brown</td>
</tr>
<tr>
<td>0.64</td>
<td>Purple</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Adaxial palisade cells
Abaxial palisade cells

Plate. 3: Transverse sections of leaves of Crotalaria lachenosema Stapf. a: midrib projecting abaxially; b: one layer of palisade cells on adaxial surface; c: two layers of palisade cells on abaxial surface

Good resolution of components was achieved with the two solvent systems used. Colouration of the spots may also be vital in compound isolation and identification as compound classes such as terpenes and steroids are known to exhibit distinct colorations.

The pharmacognostic evaluation which comprises of macromorphology and microscopic characters, the estimation of physicochemical parameters and the phytochemical and TLC profile are constant features of a plant which are highly essential for raw drugs or plant parts used for preparation of phytomedicine. Therefore, the result generated from this study would be useful in identification and standardization of the plant material towards quality assurance and also for preparation of a monograph on the plant. Detailed phytochemical studies of C. lachenosema are ongoing in our laboratory, to establish the presence or absence of this class of compounds.

ACKNOWLEDGEMENTS

The authors wish to acknowledge the contribution of Mr. John Atogwe of the forestry Unit of National Institute for Pharmaceutical Research and Development, Idu, Abuja for cultivating the plant in our garden.

REFERENCES


How to cite this article: