Journal of Applied Pharmaceutical Science Vol. 10(07), pp 059-063, July, 2020 Available online at http://www.japsonline.com DOI: 10.7324/JAPS.2020.10708 ISSN 2231-3354



In vitro anticancer activity screening of Iridaceae plant extracts

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ARTICLE INFO

ABSTRACT

Received on: 07/11/2019 Accepted on: 17/04/2020 Available online: 04/07/2020

Key words: Anti-cancer activity, *Iridaceae*, dry extracts. The different effects on human tumor cells (MCF-7, HepG2, HCT116, HeLa, HL-60, and HaCaT) for Ukrainian plant extracts have been examined for the first time. The extracts were from *Crocus sativus* (stigma, flowers, leaves, and corms), *Juno bucharica* (leaves), *Gladiolus hybrid Zefir* (leaves), and *Iris hungarica* (rhizomes) from *Iridaceae* family. All the studied extracts exhibited *in vitro* dose- and time-dependent killing capabilities on applied tumor cell lines. Their killing activity was tissue specific toward tumor cells. The HL-60 acute promyelocytic leukemia cell line and HCT116 colon adenocarcinoma cell line were found as the most sensitive toward treatment with studied plant extracts. It was found that the extracts were not active against normal human epidermal keratinocytes of the HaCaT line. *Gladiolus* leaf extract showed particularly strong anticancer properties *in vitro*. The current results for the first time suggest that *Iridaceae* plant water dry extracts are promising anticancer reagents. Subsequently, the results can be used to develop a prevention agent for the treatment of human cancers.

INTRODUCTION

According to the World Health Organization (WHO) (Cancer WHO, 2019; Lang-Schwarz *et al.*, 2019), cancer today is one of the major causes of death among the world's population; in 2018, the number of deaths was 9.6 million people. Recently, the new methods of treating oncological diseases have appeared (nanotherapy, neutron capture, and low-intensity electroresonance therapy), and besides, classical methods are used – chemotherapy, surgery, and radiation therapy. However, all of the above methods are accompanied by a number of side effects, which also negatively affect the patient's health. Therefore, the development of the cancer drugs from the plant materials and their use in the clinical practice are

Olha Mykhailenko, Department of Botany, National University of Pharmacy, Kharkiv, Ukraine. E-mail: mykhailenko.farm @gmail.com still today an urgent task (Desai et al., 2008; Sawadogo et al., 2012; Wang et al., 2019). More than 60% of drugs based on the natural products or herbal raw material are currently used for the treatment of cancer (Majolo et al., 2019; Solowey et al., 2014). The recent scientific studies show that various food plants, medicinal plants, and spices contain the primary and secondary metabolites that help in the treatment of cancer (Asowata-Ayodele et al., 2016; Deng et al., 2002). A study of the pharmacological activity of the plant extracts and their biologically active components, namely, piperine, curcumin, thymoquinone, crocin, capsaicin, oenothein B (Li et al., 2018; Zheng et al., 2016), polysaccharides (Ma et al., 2019), polyphenols (Wang et al., 2019), and other bioactive metabolites (Bobach et al., 2014), has demonstrated their anticancer, antibacterial, anti-inflammatory, antioxidant, and immunomodulatory effects. The presence of the above activities ensures the effectiveness of the prevention and treatment of different cancer types such as lung, stomach, skin, breast, cervix, liver, and prostate cancers. Currently, more than 3,000 plants worldwide have anticancer properties. From the scientific

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viewpoint, the idea of the anticancer herbal raw materials searching is not new, but for the correct user, the detailed phytochemical and pharmacological studies are always necessary. Thus, the present research results undoubtedly provide new knowledge about the anticancer effect of different dry extracts of plants from *Iridaceae* family (*Crocus, Iris, Gladiolus,* and *Juno* genus) of Ukrainian flora.

The diverse study of Iridaceae plants indicates their potential as anticancer agents. The phytochemical research of *Iridaceae* plants has shown the presence of the phenolic compounds (isoflavones, flavones, flavanones, xanthones, and hydroxycinnamic acids) (Mykhailenko et al., 2019; Singab et al., 2016) as well as polysaccharides and amino acids (Mykhailenko et al., 2020), which possess the presence of the anti-proliferative activity (Alireza et al., 2016; Asif et al., 2013; Gohari et al., 2013; Xie et al., 2014). Different plant extracts and biological active compounds of Iridaceae members have shown different pharmacological activities such as diuretic, anti-inflammatory, antiradical (Jadouali et al., 2019), hepatoprotective, antidepressant, and hypolipidemic activities (Basgedik et al., 2014; Munyemana et al., 2013; Shinwari and Rao, 2018; Singab et al., 2016) as well as immunomodulating, antimutagenic (Kuete et al., 2013; Samarghandian and Borji, 2014), antibacterial (Ayoub et al., 2014), and antitumor activities (Mykhailenko et al., 2019; Wollenweber et al., 2003; Xie et al., 2014). To the best of our knowledge, those plants have not been studied for anticancer activity on leukemia and carcinoma cells in vitro. There are a large number of articles on the antitumor potential of the individual components of Crocus sativus, such as crocin, crocetin, safranal, and picrocrocin; besides, publications on the proliferative activity of an extract from C. sativus stigma (saffron) are presented (Samarghandian and Borji, 2014). However, the effect of C. sativus extracts (stigma, leaves, leaves, and flowers) on the presented in vitro cell lines (MCF-7, HepG2, HCT116, HeLa, HL-60, and HaCaT) was not evaluated. A study of the pharmacological potential of Gladiolus and Juno plants has never been done. The present study is aimed to preliminary evaluate the anticancer potential activities against six cancer cell lines of the different tissue origin and against non-tumor cells in vitro.

MATERIALS AND METHODS

Plant material

Juno bucharica (Foster) Vved. leaves (a voucher specimen CWU0056539) and Gladiolus hybrid Zefir leaves (a voucher specimen CWU0056538) as well as Iris hungarica Waldst. et Kit. rhizomes (a voucher specimen CWU0056534) were harvested from the collection of botanical gardens of V.N. Karazin Kharkiv National University (Kharkiv, Ukraine) in May 2017. C. sativus stigmas, flowers, leaves, and corms were collected from the plantation in the village Lyubymivka (Ukraine) in November 2017 (a voucher specimen CWU0056541–CWU005654). All voucher specimens were verified by Dr. Gamulya and saved at the Herbarium of V.M. Karazin Kharkiv National University, Kharkiv, Ukraine. Fresh plant material was air-dried and crushed.

Preparation of extracts

The ground plant material of *Juno* leaves, *Gladiolus* leaves, *Crocus* leaves, flowers, and corms, and *Iris* rhizome (100 g each) was extracted with distilled water (1 l), on a water bath for

2 hours for three times. These extracts were cooled and filtered. The extracts were first concentrated using Rotavapor and then completely dried in drying cabinet and preserved at 4°C until further use. *Crocus* stigma powder (5 g) was poured with hot distilled water (500 ml, 80°C) and infused in a dark place for 24 hours. The resulting extract was drained, and the rest of the raw material was refilled with 500 ml of distilled water and left to maceration for another 24 hours at 4°C. Maceration repeated one more time in the same conditions. The obtained extracts were combined, filtered, and dried in a rotary evaporation apparatus at 80°C. The powders were stored in airtight containers at 4°C for further use.

MTT assay

The antineoplastic activity of plant extracts was established by the MTT test (EZ4U, Biomedica, Austria) (Finiuk et al., 2017) on different tissue origin cell lines. For the experiment, the cells were plated in 96-well plates of 100 µl at the concentrations of 5,000 cells/well (substrate-dependent cells) or 10,000-15,000 cells/well (suspension cells) overnight. Then, aliquots of 100 μ l of experimental extracts (0, 1, 10, and 100 μ g/ ml) as well as the reference compound doxorubicin (0, 1, and 10 μ g/ml) in culture medium were added to the cells and incubated for the next 72 hours. Furthermore, the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT reagent) was appended to the cells following the manufacture recommendations (Biomedica, Austria). The results of the reaction were determined by an Absorbance Reader Bio Tek ELx800 (BioTek Instruments, Inc., Winooski, VT) at 490 and 630 nm. The relative number of living cells (parts per unit) was determined by the ratio of optical densities measured for the wells with tested compounds to the wells with control (non-treated) cells. The half-maximal inhibitory concentration (IC₅₀) of tested extracts was calculated as the extract concentration that reduced the cell viability to 50%. Doxorubicin (Actavis, Romania) was chosen as a reference drug as a positive control. The extracts were added to the cultured cells at the indicated concentrations $(0-100 \mu g/ml)$ and then were treated for 72 hours. The results were analyzed and illustrated using GraphPad Prism 6 software (GraphPad Software., San Jose, CA). The obtained data are presented as the average \pm standard deviation (SD) of three replications in two parallels (n = 6).

Cell culture

Collection sample of HL-60 human myeloid leukemia cells (acute promyelocytic leukemia) was obtained from the Institute of Cancer Research at Vienna Medical University (Vienna, Austria). HepG2 human hepatocarcinoma cells (liver hepatocellular carcinoma), HCT116 human colon carcinoma cells (human colorectal carcinoma), and the collected sample of HaCaT human cell (normal human epidermal keratinocytes) were received from the Institute of Molecular Biology and Genetics, National Academy of Sciences of Ukraine (Kyiv, Ukraine). Collection samples of HeLa human cervix adenocarcinoma cells (cervical adenocarcinoma) and MCF-7 human breast adenocarcinoma cells (Michigan Cancer Foundation-7) were received from the R.E. Kavetsky Institute of Experimental Pathology, Oncology, and Radiobiology (Kyiv, Ukraine). All cells were grown in RPMI-1640 or Dulbecco's Modified Eagle's Medium culture supplemented with 10% fetal bovine serum (all from BioWest, France). Cells were cultivated at 37°C in an atmosphere of 5% CO₂.

RESULTS AND DISCUSSION

An in vitro screening was performed by the MTT assay of the anticancer activity of Iridaceae dry extracts toward to the following cell lines: MCF-7 human breast adenocarcinoma cells, HCT116 human colon adenocarcinoma cells, HeLa cervical adenocarcinoma cells, HepG2 hepatocellular carcinoma cells, HL-60 acute promyelocytic leukemia cells, and HaCaT normal human epidermal keratinocytes cells was performed by the MTT assay. The results of the analysis were expressed as IC₅₀ and shown in Table 1. The Iridaceae extracts have shown different antiproliferative activities. Compared to the doxorubicin, reference drug, the activity of those extracts was lower; however, some extracts had a micromolar value of the IC_{50} (Fig. 1).

The obtained data showed that the human acute promyelocytic leukemia cells of HL-60 line were the most sensitive to the action of all the studied plant extracts. The dry Gladiolus leaf extracts (Table 1 and Fig. 1) had the highest toxicity toward leukemia cells with the IC₅₀ value of 0.5 μ g/ml. Juno leaf extracts and Iris rhizome extract demonstrated similar anticancer activity with the IC_{50} of 3.5 and 3.6 µg/ml, respectively. The IC_{50} value for

Crocus corms extract was 7.4 µg/ml. The extracts of Crocus stigma, flowers, and leaves did not attain the IC₅₀ value even at 100 μ g/ml. Previously, the antitumor effect of crocin from Crocus stigma on human leukemia cells of HL-60 was studied but not for the whole extract and not to myeloid leukemia cell line (Sun et al., 2013). Thus, the potential anticancer activity of Iridaceae extracts on leukemia cells of HL-60 line can be placed in the following row: *Gladiolus* leaves > Juno leaves = Iris rhizome > Crocus corms.

Human colon adenocarcinoma cells of HCT116 line were also sensitive to the activity of plant extracts. The IC₅₀ for Gladiolus leaf extract was 9.4 µg/ml, for Iris rhizome was 42.3 µg/ml, for Crocus corms was 74.6 µg/ml, and for Crocus stigma was 94.8 µg/ml. It was reported about the effect of saffron extract (0.2-1 mg/ml) in HCT116 colorectal cancer cells (Bukhari et al., 2018), but this work includes only Crocus stigma extract. According to the results of the study, the IC₅₀ values for stigma extract ranged between 0.58 and 0.98 mg/ml. It should be noted that the composition of the active components of the raw material has a direct effect on the activity of the extracts (Mykhailenko et al., 2020). Moreover, the component composition may vary

| No | Dry extract | MCF-7 | HCT116 | HeLa | HepG2 | HL-60 | HaCaT |
|----|------------------|-------|--------|------|-------|-------|-------|
| 1 | Gladiolus leaves | >100 | 9.4 | >100 | >100 | 0.50 | >100 |
| 2 | Juno leaves | >100 | >100 | >100 | >100 | 3.5 | >100 |
| 3 | Iris rhizome | >100 | 42.3 | 78.7 | >100 | 3.6 | >100 |
| 4 | Crocus stigma | >100 | 94.8 | >100 | >100 | >100 | >100 |
| 5 | Crocus corms | >100 | 74.6 | >100 | >100 | 7.4 | >100 |
| 6 | Crocus flowers | >100 | >100 | >100 | >100 | >100 | >100 |
| 7 | Crocus leaves | >100 | >100 | >100 | >100 | >100 | >100 |
| | Dox | 0.51 | 0.79 | 0.21 | 0.34 | 0.11 | 0.7 |

Table 1. Antiproliferative activity of tested extracts, IC₅₀, µg/ml.





Figure 1. Cytotoxicity of all tested dry extracts toward different cell lines. After a total experimental time (72 hours), cell vitality was detected by the MTT assay. 1– Gladiolus leaf extract; 2—Juno leaf extract; 3—Iris rhizome extract; 4—Crocus stigma extract; 5—Crocus corm extract; 6—Crocus flower extract; 7—Crocus leaf extract.

depending on the location of the plant cultivation (Ramakrishna and Ravishankar, 2011). This is probably why saffron stigma extract from Ukraine has a slightly higher activity than the extract from Kashmir (Bukhari *et al.*, 2018). The extracts of *Juno* leaves and *Crocus* flowers and leaves did not attain the IC₅₀ value even at 100 μ g/ml (Table 1 and Fig. 1). In this case, the most active sample was also *Gladiolus* leaf extract.

Iris rhizome extract demonstrated toxicity toward human cervical adenocarcinoma of HeLa line with the IC₅₀ of 78.7 µg/ml. Other studied plant extracts were not toxic to HeLa cells (Table 1 and Fig. 1). MCF-7 human breast adenocarcinoma cells and HepG2 human hepatocellular carcinoma cells showed the weakest sensitivity to the extracts of the studied plants. The tested extracts did not have an IC₅₀ value of 100 µg/ml. Doxorubicin showed a higher cytotoxic effect toward all studied cell lines if compared to the activity of plant extracts.

It should be noted that the studied plants' extracts were not toxic to normal human epidermal keratinocytes of HaCaT line. The maximal inhibition of the growth of HaCaT cells equaled 28.6% at the action of *Crocus* corms extract and minimal at the action of *Juno* leaf extract (Table 1 and Fig. 1). The IC₅₀ value of doxorubicin for HaCaT cells was 0.7 µg/ml. Thus, *Gladiolus* leaf and *Iris* rhizome extracts demonstrated the highest cytotoxic effect toward the HL-60 human acute promyelocytic leukemia cells and HCT116 colon adenocarcinoma cells. Previously, for *C. sativus* flower acetone extract (50–200 µg/ml), the concentration-dependent activity on HaCaT cells was established (Verjee *et al.*, 2017). However, the acetone flower extract has a more toxic solvent than the aqueous extract. In the following studies, we need to increase the dose of *Iridaceae* plant extract and repeat the experiment, as other authors also take a pre-high concentration of plant extract.

In the current investigation, several species of Iridaceae genus plants have been tested for their anticancer and cytotoxic activity. Kuete et al. (2013) evaluated the cytotoxicity of the methanol extract of Gladiolus quartinianus (40 g/ml) against CCRF-CEM leukemia cells, which exhibited the lowest IC₅₀ values below 30 µg/ml. Besides, the induction of apoptosis in CCRF-CEM cells with the loss of mitochondrial membrane potential for G. quartinianus extract was found. The methanol extract of Iris kashmiriana rhizome (400 mg/ml) showed a potent cytotoxic effect against A549 human epithelial cancer cell lines and Caco-2 epithelial cell lines with IC50 of 128 and 237 mg/ml, respectively (Asif et al., 2013). Sánchez-Vioque et al. (2016) studied the antiproliferative effect of ethanol extracts from C. sativus corms, flowers, and leaves on human colon adenocarcinoma cells (Caco-2). From their results, the extract of the flower and leaves (ED₅ 0.42 mg/ml) greatly inhibited the activity of Caco-2 cells. In other investigations, C. sativus stigma aqueous extract inhibited cancer progression dose dependently in doses of 100, 150, and 175 mg/kg (Bathaie et al., 2013). Previously, Parizadeh et al. (2011) isolated a glycoconjugate from saffron corms, for which cytotoxic activity in relation to the HepG-2 hepatocellular carcinoma cell line and Hep-2 laryngeal carcinoma cell line was also established. Another study found the cytotoxic effects of crocin on HeLa and MCF-7 cell lines (Mousavi et al., 2011). However, the presented studies relate mainly to the individual compounds from C. sativus and its stigma.

For the first time, the anticancer activity of *J. bucharica* leaves, *G. hybrid Zefir* leaves, *I. hungarica* rhizomes as well as *C.*

sativus raw material extracts was investigated on MCF-7 human breast cancer cell line, HepG2 human hepatocellular carcinoma cell line, HCT116 human colon adenocarcinoma cell line, HeLa cervical adenocarcinoma cell line, HL-60 acute promyelocytic leukemia cells line, and HaCaT normal human epidermal keratinocyte cell line. Thus, the results show the potential of *Iridaceae* plants for the treatment of cancer, but further research is still needed.

CONCLUSION

In folk and clinical medicine of different countries, plants have always played an important role in the treatment and prevention of cancer and other diseases. Moreover, natural components from plants at the level of synthetic compounds are used in the creation of new drugs. Plants of Iridaceae family (J. bucharica, G. hybrid Zefir, I. hungarica, and C. sativus) are herbaceous plants traditionally used in folk medicines of European countries as antitumor, anti-inflammatory, anticonvulsant, analgesic, antioxidative activity, antipyretic, etc. Gladiolus leaf extract showed particularly strong anticancer properties in vitro toward human leukemia and colon adenocarcinoma cells in MTT assay. Iris rhizome dry extracts have shown lower activity on human cervical adenocarcinoma cells. These plant extracts have demonstrated low toxicity toward normal human epidermal keratinocytes of HaCaT line. The studies on establishing the action mechanism of the investigated extracts will be continued. These herbal extracts have shown their potential anticancer activity in vitro studies that can be used for further drug creation for the prevention and treatment of leukemia, colon, and cervix cancers. This is the first investigation about the anticancer activity of Iridaceae plant extracts.

ACKNOWLEDGMENT

The authors would like to gratitude to Dr. Orlova (Botanical garden of the Kharkiv National University named after V.N. Karazin, Kharkiv, Ukraine) for help in determining the systematic belonging of plants.

CONFLICT OF INTEREST

The authors declared that they have no conflicts of interest.

FINANCIAL SUPPORT

None.

REFERENCES

Alireza M, Kurosh D, Banafshe H. The toxicity of saffron (*Crocus sativus* L.) and its constituents against normal and cancer cells. J Nutr Intermed Metab, 2016; 3:23–32.

Asif A, Sajad HW, Taseem AM, Shoiab B, Asrar HW, Javid MI, Qazi PH, Qadri RA. Investigating the pharmacological potential of *Iris kashmiriana* in limiting growth of epithelial tumors. Pharmacognosy J, 2013; 5:170–5.

Asowata-Ayodele AM, Afolayan AJ, Otunola GA. Ethnobotanical survey of culinary herbs and spices used in the traditional medicinal system of Nkonkobe Municipality, Eastern Cape, South Africa. S Afr J Bot, 2016; 104:69–75.

Ayoub IM, El-Shazly M, Lu MC, Singab ANB. Antimicrobial and cytotoxic activities of the crude extracts of *Dietes bicolor* leaves, flowers and rhizomes. S Afr J Bot, 2014; 95:97–101.

Basgedik B, Ugurb A, Sarac N. Antimicrobial, antioxidant, and antimutagenic activities of *Gladiolus illyricus*. J Pharm Pharmacogn Res, 2014; 2:93–9.

Bathaie SZ, Miri H, Mohagheghi MA, Mokhtari-Dizaji M, Shahbazfar AA, Hasanzadeh H. Saffron aqueous extract inhibits the chemically-induced gastric cancer progression in the wistar albino rat. Iran J Basic Med Sci, 2013; 16:27.

Bobach C, Schurwanz J, Franke K, Denkert A, Sung TV, Kuster R, Mutiso PC, Seliger B, Wessjohann LA. Multiple readout assay for hormonal (androgenic and antiandrogenic) and cytotoxic activity of plant and fungal extracts based on differential prostate cancer cell line behavior. J Ethnopharmacol, 2014; 155:721–30.

Bukhari SI, Din I, Grewal S, Dhar MK. Antiproliferative effect of saffron and its constituents on different cancerous cell lines. Pharmacogn Res, 2018; 10:291–5.

Cancer. WHO, Geneva, Switzerland, 2019. Available via https:// www.who.int/news-room/fact-sheets/detail/cancer (Accessed 19 November 2019).

Deng Y, Guo Z, Zeng Z, Wang Z. Studies on the pharmacological effects of saffron (*Crocus sativus* L.) a review. Zhongguo Zhong Yao Za Zhi, 2002; 27:565–8.

Desai AG, Qazi GN, Ganju RK, El-Tamer M, Singh J, Saxena AK, Bedi YS, Taneja SC, Bhat HK. Medicinal plants and cancer chemoprevention. Curr Drug Metabol, 2008; 9:581–91.

Finiuk N, Boiko N, Klyuchivska O, Kobylinska L, Kril I, Zimenkovsky B, Lesyk R, Stoika R. 4-Thiazolidinone derivative Les-3833 effectively inhibits viability of human melanoma cells through activating apoptotic mechanisms. Croat Med J, 2017; 58:129–39.

Gohari AR, Saeidnia S, Mahmoodabadi MK. An overview on saffron, phytochemicals, and medicinal properties. Pharmacognosy Rev, 2013; 7:61–6.

Jadouali SM, Rachid HA, Majourhat MK, Abdelatif ZB, Faouzi LA. Chemical characterization and antioxidant compounds of flower parts of Moroccan *Crocus sativus* L. J Saudi Society Agricult Sci, 2019; 18:476–80.

Kuete V, Fankam AG, Wiench B, Efferth T. Cytotoxicity and modes of action of the methanol extracts of six Cameroonian medicinal plants against multidrug-resistant tumor cells. Evid Based Compl Alt Med, 2013; 2013: Article ID 285903, 10.

Lang-Schwarz C, Melcher B, Haumaier F, Schneider-Fuchs A, Lang-Schwarz K, Krugmann J, Vieth M, Sterlacci W. Budding, tumorinfiltrating lymphocytes, gland formation: scoring leads to new prognostic groups in World Health Organization low-grade colorectal cancer with impact on survival. Hum Pathol, 2019; 89:81–9.

Li H. Krstin S, Wang S, Wink M. Capsaicin and piperine can overcome multidrug resistance in cancer cells to doxorubicin. Molecules, 2018; 2:557–68.

Ma L, Xu GB, Tang X, Zhang C, Zhao W, Wang J, Chen H. Anticancer potential of polysaccharide extracted from hawthorn (*Crataegus*) on human colon cancer cell line HCT116 via cell cycle arrest and apoptosis. J Funct Foods, 2019; 64:103677.

Majolo F. Delwing LK de Oliveira B, Marmitt DJ, Bustamante-Filho IC, Goettert MI. Medicinal plants and bioactive natural compounds for cancer treatment: important advances for drug discovery. Phytochem Lett, 2019; 31:196–207.

Mousavi SH, Moallem SA, Mehri S, Shahsavand S, Nassirli H, Malaekeh-Nikouei B. Improvement of cytotoxic and apoptogenic properties of crocin in cancer cell lines by its nanoliposomal form. Pharm Biol, 2011; 49:1039–45.

Munyemana F, Mondego AP, Cumbane P. Qualitative phytochemical screening and antimicrobial activity evaluation of the bulb extracts of *Gladiolus psittacinus* Hook (*Iridaceae*). Int Net Env Manag Conflicts, 2013; 2:14–31.

Mykhailenko O, Ivanauskas L, Bezruk I, Lesyk R, Georgiyants V. Comparative investigation of amino acids content in the dry extracts of *Juno bucharica, Gladiolus hybrid Zefir, Iris hungarica, Iris variegata* and *Crocus sativus* raw materials of Ukrainian flora. Sci Pharm, 2020, 88:1–13.

Mykhailenko O, Kovalyov V, Goryacha O, Ivanauskas L, Georgiyants V. Biologically active compounds and pharmacological activities of species of the genus *Crocus*: a review. Phytochemistry, 2019; 162:56–89.

Parizadeh MR, Gharib FG, Abbaspour AR, Afshar JT, Ghayour-Mobarhan M. Effects of aqueous saffron extract on nitric oxide production by two human carcinoma cell lines: Hepatocellular carcinoma (HepG2) and laryngeal carcinoma (Hep2). Avicenna J Phytomed, 2011; 1:43–50.

Ramakrishna A, Ravishankar GA. Influence of abiotic stress signals on secondary metabolites in plants. Plant Signal Behav, 2011; 6:1720–31.

Samarghandian S, Borji A. Anticarcinogenic effect of saffron (*Crocus sativus* L.) and its ingredients. Pharmacognosy Res, 2014; 6:99–107.

Sánchez-Vioque R, Santana-Méridas O, Polissiou M, Vioque J, Astraka K, Alaizd M, Herraiz-Peñalvera D, Tarantilis PA, Girón-Calle J. Polyphenol composition and *in vitro* antiproliferative effect of corm, tepal and leaf from *Crocus sativus* L. on human colon adenocarcinoma cells (Caco-2). J Funct Foods, 2016; 24:18–25.

Sawadogo WR, Schumacher M, Teiten MH, Dicato M, Diederich M. Traditional west African pharmacopeia, plants and derived compounds for cancer therapy. Biochem Pharmacol, 2012; 84:1225–40.

Shinwari KJ, Rao PS. Thermal-assisted high hydrostatic pressure extraction of nutraceuticals from saffron (*Crocus sativus*): Process optimization and cytotoxicity evaluation against cancer cells. Innov Food Sci Emerg Technol, 2018; 48:296–303.

Singab ANB, Ayoub IM, El-Shazly M, Korinek M, Wu TY, Cheng YB, Chang FR, Wu YC. Shedding the light on *Iridaceae*: ethnobotany, phytochemistry and biological activity. Ind Crop Prod, 2016; 92:308–35.

Solowey E, Lichtenstein M, Sallon S, Paavilainen H, Solowey E, Lorberboum-Galski H. Evaluating medicinal plants for anticancer activity. Sci World J, 2014; 2014:721402–14

Sun Y, Xu HJ, Zhao YX, Wang LZ, Sun LR, Wang Z, Su XF. Crocin exhibits antitumor effects on human leukemia HL-60 cells *in vitro* and *in vivo*. Evid Based Complement Alternat Med, 2013; 3:690164.

Verjee S, Garo E, Pelaez S, Fertig O, Hamburger M, Butterweck V. Saffron flower extract promotes scratch wound closure of keratinocytes and enhances VEGF production. Planta Med, 2017; 83:1176–83.

Wang X, Chen Y, Fang Z. *In-vitro* photothermal therapy using plant extract polyphenols functionalized graphene sheets for treatment of lung cancer. J Photochem Photobioi B: Biology, 2019; 204:111587–601.

Wollenweber E, Stevens JF, Klimo K, Knauft J, Frank N, Gerhauser C. Cancer chemopreventive *in vitro* activities of isoflavones isolated from *Iris germanica*. Planta Med, 2003; 69:15–20.

Xie GY, Zhu Y, Shu P, Qin XY, Wu G, Wang Q, Qin MJ. Phenolic metabolite profiles and antioxidants assay of three *Iridaceae* medicinal plants for traditional Chinese medicine "She-gan" by on-line HPLC–DAD coupled with chemiluminescence (CL) and ESI-Q-TOF-MS/ MS. J Pharmaceut Biomel Anal, 2014, 98:40–51.

Zheng J, Zhou Y, Li Y, Xu DP, Li S, Li HB. Spices for prevention and treatment of cancers. Nutrients, 2016; 8:495–530.

How to cite this article:

Mykhailenko O, Lesyk R, Finiuk N, Stoika R, Yushchenko T, Ocheretniuk A, Vaschuk V, Mishchenko V, Georgiyants V. *In vitro* anticancer activity screening of *Iridaceae* plant extracts. J Appl Pharm Sci, 2020; 10(07):059–063.