

Multidrug-resistant Gram-negative bacilli producing oxacillinases and Metallo- β -lactamases isolated from patients in intensive care unit - Annaba hospital - Algeria (2014-2016)

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ABSTRACT

This study aims to investigate the prevalence and genes encoding for carbapenemases of multidrug-resistant Gram-negative bacilli isolated from the intensive care unit in the hospital of Annaba city, Algeria, over a period of 18 months. The isolates were identified using API20E, API 20NE and confirmed by Microscan WalkAway 96. Antimicrobial susceptibility testing was determined by the disk diffusion method according to clinical and laboratory standards institute guidelines. Carbapenemases production was tested using Hodge and imipenem-EDTA Combined Disc tests. Carbapenemases genes were established by Polymerase chain reaction and confirmed by sequencing. 35.52% of 152 isolates were multidrug-resistant. The majority belong to *Enterobacteriaceae* (75.92%) followed by non-fermentative Gram-negative bacilli (24.07%). The *Enterobacteriaceae* revealed high resistance to third generation cephalosporin, with the production of cephalosporinases (36.58%) and/or extended-spectrum β -lactamases (34.14%). For non-fermentative Gram-negative bacilli, they showed increased resistance to carbapenems, fluoroquinolones, and aminoglycosides. Fifteen of the MDR strains were resistant to carbapenems, 10 of them were positives for Metallo- β -lactamases and carbapenemases by phenotypic methods. The PCR results showed the presence of *bla*_{VIM-2} gene in two *Pseudomonas aeruginosa*, *bla*_{OXA-24} (*n* = 1), *bla*_{OXA-23} (*n* = 6) and *bla*_{NDM-1} (*n* = 1) in *Acinetobacter baumannii*. A surveillance of the resistance in this unit is necessary to prevent any therapeutic impasse.

INTRODUCTION

Infections in intensive care units (ICUs) are becoming not only recurrent but also increasingly complicated. They cause high morbidity and mortality over the world. These infections are usually caused by multi-drug resistant bacteria (MDR), especially Gram negative-bacilli (GNB), where their prevalence in Algeria and all over the world increases in a worrying way and represents a serious public health problem (Baba Ahmed and Arlet, 2014).

The most MDR bacteria involved are the GNB represented by *Enterobacteriaceae*; they are frequently isolated from hospitalized patients (Kaye and Pogue, 2015). *Pseudomonas aeruginosa* and *Acinetobacter baumannii* appears as the major agent causing infections in intensive care units, especially for immunocompromised patients. Their ability to develop resistance to antimicrobial agents makes treatment of these infections ever more difficult (McGowan, 2006). The infections caused by these MDR bacteria are essentially related to the frequent use of invasive procedures.

The most common resistance mechanism is the secretion of extended-spectrum β -lactamases (ESBLs) (Kaye and Pogue, 2015); these enzymes confer a resistance to the majority of β -lactams, posing then a high risk of therapeutic failure and the

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emergence of MDR. In this case, carbapenems are prescribed for the reason of their stability to most natural or acquired β -lactamases. However, the resistance for this class of antibiotics has taken place all over the world, with the emergence of *Enterobacteriaceae* producing carbapenemases (EPC) (Poirel *et al.*, 2013), carbapenem-resistant *P. aeruginosa* and *A. baumannii*. Carbapenem resistance in non-fermentative Gram-negative bacilli (NF-GNB) is due to a variety of drug resistance mechanisms such as the loss of porins OprD and efflux pumps. The most common mechanism is the acquisition of genes encoding for carbapenemases particularly, Metallo- β -lactamases (MBLs) of *P. aeruginosa* where the most common genes reported are VIM, IMP, GIM, FIM, and SPM (Rodriguez-Martinez, 2009). The *bla*_{VIM-2} is the dominant Metallo- β -lactamase in North Africa and all over the world (Hashem *et al.*, 2017). The common oxacillinases for *A. baumannii* are OXA-23, 24, 58 of which OXA-23 is the most frequent. These enzymes have been reported in clinical isolates from different hospitals in western and eastern Algeria and in other parts of the world (Touati *et al.*, 2012; Mohajeri *et al.*, 2017; Pogue *et al.*, 2013).

GNB represents one of the main causes of serious infections in the hospitals of Annaba city. Their resistance to antibacterial agents is being increasingly complicated. The aim of the present study is to investigate the prevalence of Gram-negative bacilli multidrug-resistant (MDR-GNB) and characterization of genes encoding for carbapenemases isolated from the intensive care unit (department of anesthesia-resuscitation) of Ibn Rochd hospital, Annaba, Algeria, over a period of 18 months.

MATERIALS AND METHODS

Collected data

The prospective study on MDR-GNB, isolated from various specimens was performed over 18 months (December 2014 to May 2016) from distal protected sampling (DPS), pus, rectal swab (RS) and urine taken from patients hospitalized in the intensive care unit (Ibn Rochd hospital, Annaba). The information about patients has been obtained from the patient medical records including age, sex, anterior hospitalization, clinical signs, hospitalization period and the presence or absence of risk factors.

Identification and antibiotic susceptibility testing

The GNB isolates were identified using API20E, API20NE galleries (Bio-Mérieux France) and Microscan WalkAway 96 plus (Siemens) automate. Antibiotic susceptibility was determined using the Muller-Hinton agar diffusion method, according to the recommendations of Clinical and Laboratory Standards Institute (CLSI). Eleven antibiotics were tested for *Enterobacteriaceae*: amoxicillin, amoxicillin/clavulanic acid, cefotaxime, ceftriaxone, aztreonam, ceftazidime, imipenem, ertapenem, gentamycin, amikacin, and ciprofloxacin. For NF-GNB, eleven antibiotics were also tested: piperacillin, cefotaxime, ceftriaxone, ceftazidime, aztreonam, imipenem, gentamycin, amikacin, ciprofloxacin, ticarcillin, ticarcillin/clavulanic acid.

The determination of minimum inhibitory concentrations (MIC) were carried out by automated Microscan WalkAway 96 plus, using plates adapted to each kind of bacteria. The MIC

of ertapenem and imipenem were determined in solid media, following the CLSI recommendations.

All the isolates found simultaneously resistant to two or more antibiotics of different families are considered as multidrug-resistant (according to Center for Disease Control (CDC) criteria), and are included in this study.

Phenotypic detection for extended-spectrum β -lactamases (ESBL)

The ESBL production was detected by the double-disc synergy test (DDST) and by a cloxacillin test for strains that are resistant to cefotaxime, ceftazidime and/or aztreonam. In DDST, a disc of amoxicillin/clavulanic acid (AMC 30 μ g) was placed in the center of MHA at 30 mm distance to ceftazidime (CAZ 30 μ g) and cefotaxime (CTX 30 μ g). ESBL production was detected by the appearance of keyhole effect between AMC disc and C3G (Drieux *et al.*, 2008). The cloxacillin test has been also done on MHA supplemented with 250 mg/L of cloxacillin, in order to detect any ESBL eventually masked by AmpC cephalosporinase (Drieux *et al.*, 2008).

Phenotypic detection of carbapenemases and metallo- β -lactamases (MBLs)

All the isolates resistant to imipenem and ertapenem were subjected to modified Hodge test (MHT) as was described by CLSI guidelines. A dilute suspension of 1/10 of *E. coli* ATCC 25922 (0.5 McFarland) was prepared and inoculated on MHA. An imipenem disk (10 μ g) is placed in the center of the test area. Test organism was streaked in a straight line from the edge of the disk to the edge of the plate. The plate was incubated at 35°C for 16–24 h. MHT Positive test showed a cloverleaf-like indentation of the *E. coli* 25922 growing along the test organism growth streak within the disk diffusion zone.

The combined-disc test using imipenem-EDTA was adopted for the detection of metallo- β -lactamases as previously described by Esther *et al.* (2017). Two imipenem discs (10 μ g) were placed 25 mm apart on an MHA plate, 10 μ L of EDTA was added to one of them. The plate was incubated 16-24 h at 35°C. An increase in the inhibition zone diameter greater than 6 mm around the imipenem-EDTA disc compared with that of the imipenem disc alone was considered positive for MBL production.

Molecular search for genes encoding carbapenemases and their sequencing

The search for genes encoding for carbapenemases was realized by real-time Polymerase Chain Reaction (PCR) and standard PCR, using primers and probes specific to various reference strains. The amplified PCR products obtained have been sequenced using Big Dye terminator chemistry on an ABI 3130XL automated sequencer (Applied Biosystems, Foster City, California, United States). The nucleotide and deduced protein sequences were analyzed using the ARG-ANNOT (Antibiotic Resistance Gene-ANNOTation) (Gupta *et al.*, 2014).

RESULTS AND DISCUSSION

Bacterial isolates

A total of 152 GNB have been isolated (13 to 81 years, mean age = 39.5). The identification allowed to class 122 strains

in the family of *Enterobacteriaceae* (80.26%) and 30 NF-GNB (19.73%). More than 35% of the isolates ($n = 54$) were MDR. This result is comparable to that obtained in India (33.5%) (Silpi *et al.*, 2016), but higher than the results obtained in the University Hospital of Marrakech, Morocco (20%) (Arsalane *et al.*, 2010). In France, many hospitals reported a high prevalence of MDR, especially in intensive care units since the 1990s (Régnier, 1996). These MDRs bacteria were isolated from urine ($n = 18$, 33.33%), various pus ($n = 21$, 38.88%), RS ($n = 8$, 14.81%) and DPS ($n = 7$, 12.96%).

The high prevalence of MDR-GNB reported worldwide is due to several factors, including the extremely vulnerable patient, the use of immunosuppressive drugs that reduce resistance to infection, frequent use of invasive devices, prolonged

hospitalization, and the abuse of broad-spectrum antibiotics (Ha YE *et al.*, 2013; Kaye and Pogue, 2015). This explains our results where all patients infected with MDR have undergone at least two invasive procedures: intubation for 88.88% of cases, catheterization and urinary catheter for 100% of them. The infected patients were hospitalized more than 10 days. On the other hand, 26.62% ($n = 16$) of patients are transferred from other services (gastrology, gynecology or traumatology), or have been hospitalized at least once in the current year. An antibiotics treatment has been noted for 90.7% of the studied cases. Ceftizoxime, cefotaxime were used in 12.24% of cases, associated with gentamycin or amikacin for 40.81% or ciprofloxacin for 10.20% of them. Carbapenems are used in association with an aminoglycoside to treat infections caused by MDR (Table 1).

Table 1: Distribution of MDR-isolated strains related to clinical characteristics of patients.

Parameter	Fermentative bacteria				No-fermentative bacteria.		
	<i>E. coli</i> $n = 14$	<i>Klebsiella</i> sp. $n = 13$	<i>Enterobacter</i> sp. $n = 7$	<i>Serratia</i> sp. $n = 4$	<i>C. freundii</i> $n = 3$	<i>P. aeruginosa</i> $n = 5$	<i>A. baumannii</i> $n = 8$
Invasive devices: n (%)							
Urinary catheter	14(100)	13(100)	7(100)	4(100)	3(100)	5(100)	8(100)
Gastric catheter	2(14.28)	00	00	00	00	1(20)	2(25)
Catheterization	14(100)	13(100)	7(100)	4(100)	3(100)	5(100)	8(100)
Intubation	12(85.71)	11(84.61)	7(100)	2(50)	3(100)	5(100)	8(100)
Ventilation	5(35.71)	9(69.23)	3(42.85)	2(50)	2(50)	5(100)	6(75)
Source: n (%)							
Urine	5(35.71)	5(38.46)	2(28.57)	1(25)	2(66.66)	1(20)	2(25)
DPS	1(7.14)	1(7.69)	2(28.57)	1(25)	00	2(40)	00
RS	4(28.57)	2(15.38)	1(14.28)	00	1(33.33)	00	00
Various pus	4(28.57)	5(38.46)	2(28.57)	2(50)	00	2(40)	6(75)
Previous antibiotic use: n (%)							
C3G	2(15.38)	2(15.38)	1(14.28)	00	1(33.33)	00	00
C3G + aminoglycoside	5(35.71)	1(7.69)	6(85.71)	1(25)	1(33.33)	1(20)	5(62.5)
C3G + fluoroquinolone	1(7.14)	1(7.69)	00	1(25)	00	1(20)	1(12.5)
Aminosides+ fluoroquinolone	1(7.14)	1(7.69)	00	1(25)	00	00	00
Carbapenems +aminoside	00	1(7.69)	00	00	00	4(80)	3(37.5)
Aminoside	2(15.38)	4(30.76)	00	00	1(33.33)	00	00

DPS = distal protected sampling; RS = rectal swab; C3G = third-generation cephalosporin.

However, other authors also report the important role of inanimate surfaces in the spread of resistant bacteria and the emergence of nosocomial infection (Zenati *et al.*, 2016).

The most isolated MDR are *Enterobacteriaceae* ($n = 41$, 75.92%), especially *E. coli* and *Klebsiella* sp. ($n = 27/41$) followed by *Enterobacter* ($n = 7/41$), *Serratia* ($n = 4/41$) and *Citrobacter* ($n = 3/41$). The other MDR group constitutes of the NF-GNB, particularly *A. baumannii* ($n = 8$, 14.81%) and *P. aeruginosa* ($n = 5$, 9.25%), which represent 24.07% of MDR isolates. These results have been previously reported by other studies in Annaba city (Mellouk *et al.*, 2017). Our findings are also correlated to other studies in Gulf countries (Aly and Balkhy, 2012) and India (Silpi *et al.*, 2016). While another study in a tertiary care hospital in Riyadh city (KSA), reported that the most common isolate pathogens are *P. aeruginosa*, *A. baumannii* followed by *E. coli* and *K. pneumoniae* (Aljohani *et al.*, 2010). The multidrug-

resistant strains of *P. aeruginosa* ($n = 5$) have been isolated mainly from pus but also from urine and DPS, *A. baumannii* strains ($n = 8$) were isolated exclusively from various pus.

Antimicrobial susceptibility testing

Fermentative bacteria

The results of the antibiotic susceptibility testing for *Enterobacteriaceae* revealed an increased resistance to fluoroquinolones, exceeding 60% for ciprofloxacin, which is the main antibiotic used for the treatment of urinary infections specifically caused by *E. coli*. Resistance to the majority of tested β -lactams showed high rates: 100% (cefotaxime), 85.36% (ceftriaxone), 75.60% (amoxicillin/clavulanic acid), 68.29% (ceftazidime) and 53.65% (aztreonam). All these molecules represent the antibiotics of choice for treating infections in this department. Resistance rate

to other classes of antibiotics was 24.39%, 56.09% to amikacin and gentamycin respectively. However, the isolates remain sensitive to imipenem, colistin, and tigecycline.

Resistance to carbapenems has been recorded only for two strains with a MIC value ranging from 4 to 8 µg/ml

for ertapenem (the carbapenem that has the best sensitivity for the detection of EPCs especially OXA-48) but it was not possible to demonstrate the production of carbapenemases by biochemical tests. Details of antimicrobial susceptibility testing are demonstrated in Figure 1.

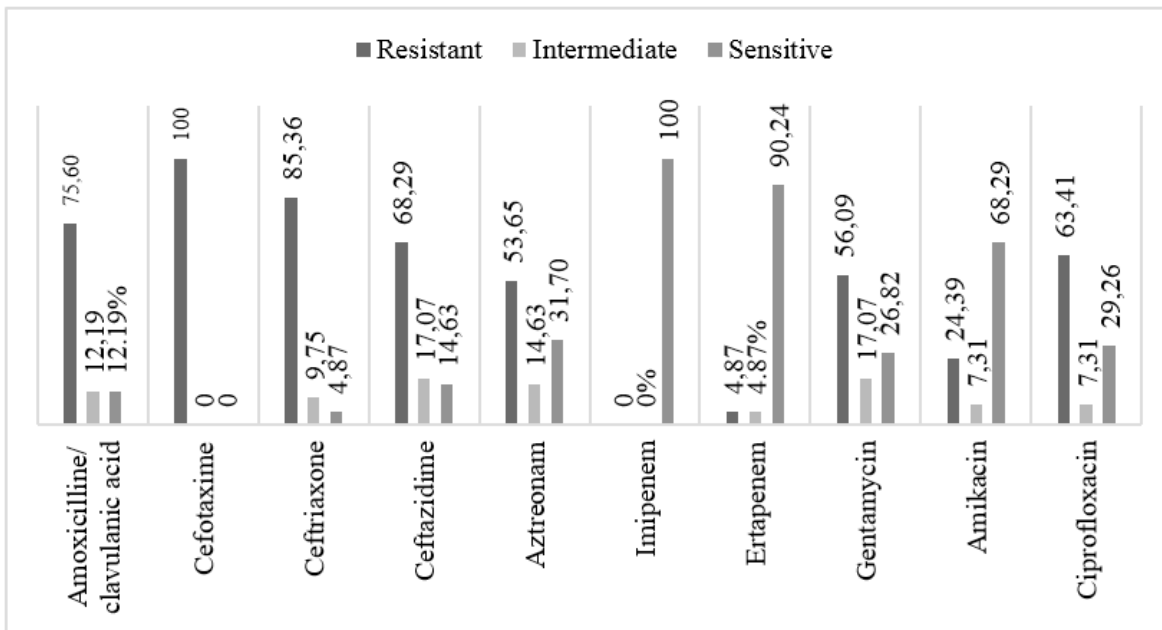


Fig. 1: Antimicrobial susceptibility of multi-drug-resistant *Enterobacteriaceae* isolates.

Enterobacter sp. and *S. marcescens* isolated strains are relatively few but they showed the highest rates of resistance for the majority of tested antibiotics.

Algeria considered as a country with a high epidemic of ESBL, where their prevalence in intensive care unit has increased from 43.73% in 2014 to 44.16 % in 2015 according to Algerian network for surveillance the resistance of bacteria to antibiotics report's (AARN, 2014-2015). In our study *Enterobacteriaceae* producing ESBL is detected for 11.47% of a total of strains ($n = 14/122$; 34.14% for MDR isolates) which is comparatively lower than Tlemcen (Algeria) (32.1%) (Ahmed *et al.*, 2012) and Burkina Faso (58%) (Ouedraogo *et al.*, 2016). Many studies reported that *E. coli* is the main ESBL producing followed by *K. pneumoniae*, which is clearly increasing (Ouedraogo *et al.*, 2016). Other study revealed the predominance of *K. pneumoniae* (Dropa *et al.*, 2009). Our findings revealed that the number of *E. coli* producing ESBL ($n = 6$, 42.85%) is the same as that of *K. pneumoniae* ($n = 6$, 42.85%) followed by *Serratia sp.* ($n = 2$, 14.28%). These ESBL producing *Enterobacteriaceae* comes from urine samples (6/14), rectal swabs (3/14), DPS (3/14) and pus (2/14).

The question that arises is how to identify the grouped cases of cross-transmission or linked to an environmental reservoir. In this case, molecular typing on positive ESBL isolates from infected or colonized patients is needed to determine the clonal distribution (Ha YE *et al.*, 2013).

The existence of ESBL bacteria is more commonly associated with the resistance to other classes of antibiotics such as

fluoroquinolones, as shown in our study where 58.33% of ESBLs are resistant to ciprofloxacin. Plasmid gene transfer (Qnr) has been reported in *Enterobacteriaceae* isolates simultaneously producing ESBL, by several authors in Annaba city, Algeria (Meradi *et al.*, 2011). The lowest rates of co-resistance are recorded for ertapenem (8.33%), whereas highest level noted for amikacin (33.33%).

The production of ESBL is associated with the elaboration of cephalosporinases for 17.07% of strains, revealed by the cloxacillin test. Only 36.58% ($n = 15$) of studied MDR strains were found to be AmpC β -lactamases producers.

Non-fermentative – Gram-negative bacilli

P. aeruginosa and *A. baumannii* are the most NF-GNB studied because of their higher isolation rates and their increased resistance to almost all available antimicrobials. The isolates obtained in the current study showed different levels of resistance to the antimicrobial agents. The MIC values are high for the β -lactams family, ranging from 16 to 128 µg/ml for imipenem and meropenem, > 16 µg/ml for aztreonam, > 64 µg/ml for ticarcillin, > 32 µg/ml for ceftazidime, > 16 µg/ml for cefepime, > 32 µg/ml for amikacin, > 2 µg/ml for ciprofloxacin. They had sensitivity only to polymyxin-E (2 µg/ml) and tigecycline (*A. baumannii* MIC = 1-2 µg/ml), which limits then the antibiotherapeutic possibilities and increases the costs of care and mortality rate (Al Johani *et al.*, 2010).

The emergence of imipenem-resistant *A. baumannii* (IRAB) has become a universal concern, as these molecules are often the only effective treatment against these strains (Otéo *et al.*,

2007). In our study the imipenem resistance rate is 80% ($n = 8/10$), which is similar to results found in Iran (>80%) (Mohajeri *et al.*, 2017) and Algeria (75.2%) (Khorsi *et al.*, 2015), significantly higher than those reported in Saudi Arabia (5.4%) (Memish *et al.*, 2012).

P. aeruginosa has a sensitivity rate of 75% to imipenem ($n = 15/20$); this value is comparable to that reported in Egypt (73.5%) (Hashem *et al.*, 2017). Higher than that reported in Iran (4%) (Saffari *et al.*, 2016), a little higher than that reported in another study in Annaba city (50%) (Meradji *et al.*, 2016) and a study in western Algeria between 2009 and 2012 (39.32%) (Sefraoui *et al.*, 2014).

Among these strains 43.33% ($n = 13/30$) are resistant to carbapenems, only two strains of *P. aeruginosa* ($n = 2/20$, 10%) are positive for production of carbapenemases by both MHT and MBL screen, these enzymes are detected in 80% ($n = 8/10$) of

A. baumannii isolates. Both MHT and MBL screen were positive only for one strain of them. No strain of them produced ESBL.

Resistance gene determination

Several studies have identified a variety of oxacillinases in carbapenem-resistant *A. baumannii*. The bla_{OXA-23} gene was reported worldwide, the bla_{OXA-24} , bla_{OXA-72} , and bla_{OXA-58} genes were described later (Pogue *et al.*, 2013). In Algeria, bla_{OXA-23} and bla_{OXA-24} were reported by Touati *et al.* from patients hospitalized in the intensive care units (2012), bla_{OXA-24} and bla_{OXA-72} reported in Setif, bla_{OXA-23} in Tizi-Ouzou (Bakour *et al.*, 2012), bla_{OXA-23} and bla_{OXA-24} in Tlemcen, Oran and Algiers (Mesli *et al.*, 2013; Khorsi *et al.*, 2015). The bla_{OXA-23} gene was founded in six strains of *A. baumannii* resistant to imipenem (60%) while the bla_{OXA-24} was described only for one isolate (10%) Table 2.

Table 2: Genes encoding carbapenemases of *P. aeruginosa* and *A. baumannii* isolates.

Strains	Sex/Age	Source	Diagnostic	Antibiotics use	CMI/IMP (µg/ml)	Genes
<i>P. aeruginosa</i>	M/27	pus	traumatic brain injury	- ceftizoxime, gentamycine - imipenem, amikacin	128	VIM-2
<i>P. aeruginosa</i>	M/50	DPS	polytrauma	- ceftizoxime, gentamycine - imipenem, gentamicine vancomycine	128	VIM-2
<i>A. baumannii</i>	M/36	DPS	peritonitis	- ceftizoxime, gentamycine - imipenem, gentamicine Vancomycine	16	OXA-23
<i>A. baumannii</i>	M/25	pus	traumatic brain injury	- ceftizoxime, metronidazol - cefotaxime, ciprofloxacin, gentamycine	16	OXA-23
<i>A. baumannii</i>	M/19	pus	hemorrhagic pulmonary contusion	- ceftizoxime, gentamicine	32	OXA-23
<i>A. baumannii</i>	F/41	pus	polytrauma	- cefotaxime, gentamicine	16	OXA-23
<i>A. baumannii</i>	F/53	DPS	escarre	- ceftizoxime, ciprofloxine, metronidazol	16	OXA-23
<i>A. baumannii</i>	M/65	AT	insufficiency renal	- imipenem	16	OXA-23
<i>A. baumannii</i>	F/32	AT	post-surgery coma	- ceftizoxime, gentamycine - tienam, gentamicine vancomycine	16	OXA-24
<i>A. baumannii</i>	M/21	AT	polytrauma	- ceftizoxime, metronidazol - cefotaxime, ciprofloxacin, gentamycine	64	NDM-1

DPS = distal protected sampling; AT = aspiration tracheal.

New Delhi MBL (NDM-1) has been mostly detected in *K. pneumoniae* and *E. coli* clinical isolates and lower in *Pseudomonas sp.* and *Acinetobacter sp.* The first description of this enzyme in *A. baumannii* in Algeria was reported by Boulanger *et al.* (2012), Bogaerts *et al.* (2012) and then reported by Bakour *et al.* (2012), Mesli *et al.* (2013), and Khorsi *et al.* (2015). Recently, this gene was detected for the first time in Algeria in *Acinetobacter nosocomialis* isolates in Ouargla city (Yagoubat *et al.*, 2017); in this study, the bla_{NDM-1} gene was only detected in one isolate of *A. baumannii* (Table 2).

In this study, two *P. aeruginosa* strains harbor bla_{VIM-2} (MBL) (Table 2), which is the most frequently isolated gene in

this bacteria, previously identified in Annaba (Touati *et al.*, 2013; Meradji *et al.*, 2016) and Oran (Sefraoui *et al.*, 2014). The bla_{VIM-4} gene was reported by Mellouk *et al.* (2017) and Meradji *et al.* (2016) in the burns department. Recapitulative of carbapenemases genes detected in non-fermentative bacteria in Algeria is presented in Table 3.

The strains of *P. aeruginosa* resistant to imipenem but non-productive of carbapenemases may be attributed to OprD mutations, but this mechanism is not limited only to Carbapenem-resistant isolates, but also for the isolates with MIC of only 0.06 to 4 µg/ml of imipenem or meropenem. The phenomenon of efflux ad can be also mentioned.

Table 3: Carbapenemases genes of *Acinetobacter* sp. and *Pseudomonas aeruginosa* detected in Algeria.

Species	Genes	Year	Lieux	References	Presentstudy
<i>P. aeruginosa</i>	VIM-2	2013	Annaba	Touati <i>et al.</i> , 2013	
	VIM-2	2014	Oran	Sefraoui <i>et al.</i> , 2014	
	VIM-2	2016	Annaba	Meradji <i>et al.</i> , 2016	VIM-2
	VIM-4	2016	Annaba-Skikda	Mellouk <i>et al.</i> , 2017	
	VIM-4	2016	Annaba	Meradji <i>et al.</i> , 2016	
<i>A. baumannii</i>	OXA-23	2010, 2012, 2013, 2014, 2015	Annaba, Tizi-Ouzou, Setif, Tlemcen, Sidi Bel-Abbes, Oran, Algiers, Bejaia	Touati <i>et al.</i> , 2012 Mellouk <i>et al.</i> , 2017 Bakour <i>et al.</i> , 2012 Kempf <i>et al.</i> , 2014 Khorsi <i>et al.</i> , 2015 Mugnier <i>et al.</i> , 2010	OXA-23
	OXA-24	2012, 2013, 2014, 2015	Setif, Tlemcen, Sidi-Bel Abbes, Oran, Tizi-Ouzou, Alger	Bakour <i>et al.</i> , 2014 Kempf <i>et al.</i> , 2014 Khorsi <i>et al.</i> , 2015 Mesli <i>et al.</i> , 2013	OXA-24
	NDM-1	2012, 2013, 2014, 2015, 2016	Oran, Alger, Annaba	Bulanger <i>et al.</i> , 2012 Bogaerts <i>et al.</i> , 2012 Bakour <i>et al.</i> , 2014 Khorsi <i>et al.</i> , 2015 Mesli <i>et al.</i> , 2013 Mellouk <i>et al.</i> , 2017	NDM-1
	OXA-58	2010, 2012	Tlemcen, Annaba	Drissi <i>et al.</i> , 2010 Touati <i>et al.</i> , 2012	
	OXA-94	2012	Oran	Bulanger <i>et al.</i> , 2012	
	OXA-51	2012, 2014, 2015	Setif, Tizi-Ouzou, Algiers, Bejaia	Bakour <i>et al.</i> , 2012 Khorsi <i>et al.</i> , 2015	
	OXA-23 et OXA-58	2012	Annaba	Touati <i>et al.</i> , 2012	
	OXA-23 et OXA-72	2012	Setif	Bakour <i>et al.</i> , 2012	
	OXA-23 et OXA-24	2012	Tlemcen, Setif, Sidi-Bel Abbes, Oran, Tizi-Ouzou	Kempf <i>et al.</i> , 2012	
	NDM-1 et OXA-23	2014	Algiers	Bakour <i>et al.</i> , 2014	
<i>Acinetobacter nosocomialis</i>	NDM-1	2016	Ouargla	Yagoubat <i>et al.</i> , 2017	

CONCLUSION

The intensive care unit in Ibn Rochd hospital shows a high prevalence of MDR (more than a third of isolates are MDR). This situation is probably due to the higher rate, excessive and inappropriate use of antimicrobials; making infections more difficult to treat and increases the cost and duration of hospitalization. *E. coli*, *K. pneumoniae*, *P. aeruginosa* and *A. baumannii* are the most frequent incriminated infections MDR-GNB agent in this structure harboring a various genes encoding for oxacillinases and metallo- β -lactamases. The situation is worrying that could lead to outbreaks of epidemic resistance because they are carried by mobile genetic elements easily transferable between Gram-negative species. Some measures must be taken, to slow down the rising problem of MDR especially; rational use of antibiotics and infection control to prevent any potential epidemic risk.

CONFLICT OF INTEREST

None.

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