High Performance Liquid Chromatography (HPLC) Fingerprinting, Mineral Composition and In Vitro Antioxidant Activity of Methanol Leaf Extract of Synsepalum dulcificum (Sapotaceae)

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ABSTRACT

This study was carried out to identify the phytochemicals and in vitro antioxidant activity of methanol leaf extract of Synsepalum dulcificum (MSD). Standard protocols were used to evaluate the total phenols, total flavonoids and total antioxidants content of the extract. Nitric oxide (NO), hydroxyl radical (OH), ABTS·+ and 2,2-diphenyl -1-picrylhydrazyl (DPPH) radicals scavenging activity, inhibition of lipid peroxidation and the ability of MSD to chelate ferrous ion as well its reductive potential were also evaluated. High Performance Liquid Chromatography (HPLC) was used to confirm the presence of polyphenols and carotenoids. Results showed the presence of flavonoids, saponins, terpenoids and cardiac glycosides in the extract. Among others, appreciable levels of potassium, calcium, sodium and magnesium were detected in the extract. The IC50 of the extract for DPPH, NO, OH−, and ABTS·+ radicals scavenging assays were 139.45 μg/ml, 119.17 μg/ml, 147.65 μg/ml, and 135.83 μg/ml respectively. It could be inferred that MSD showed appreciable in vitro antioxidant activity and could be useful in preventing and ameliorating diseases in which free radicals are implicated.

INTRODUCTION

In recent times medicinal plants have occupied a unique position among the treatment modalities of a wide variety of diseases. This might not be unconnected with the presence of biologically active molecules in such plants especially molecules that have been proven to have therapeutic potentials in several experimental disease models. Such molecules include flavonoids, tannins, steroids, terpenoids, saponins etc. These molecules have been shown to be responsible for the medicinal value of the plants in which they are found, including their antioxidant activity (Yildrim et al., 2001).

The antioxidant activity of a compound has been attributed to various mechanisms viz: prevention of chain initiation, binding of transition metal ion catalysts, decomposition of peroxides, prevention of continued hydrogen abstraction, reductive capacity and radical scavenging ability (Saumya and Mahabooob, 2011).
Moreover, The World Health Organization (WHO) stated that more than 80% of the population in developing countries use herbal and other traditional medicines to treat their common ailments (WHO, 1998). Free radicals are highly reactive chemical species regularly produced in the human system by normal biological reactions and also by various exogenous factors. Some of the radicals include superoxide $O_2^-$, nitric oxide (NO), peroxyl (ROO), and hydroxyl (OH) (Maritim et al., 2003). These ROS play an important role in degenerative or pathological processes, such as aging, cancers, coronary heart diseases, Alzheimer’s disease, neurodegenerative disorders, atherosclerosis, cataracts and inflammations (Huang et al., 2005). Living organisms have endogenous antioxidant defense systems that protect against oxidative damage by removal or repair of damaged molecules (Sun et al., 1998). The term ‘antioxidant’ refers to the activity of numerous vitamins, minerals and phytochemicals which provide protection against the damage caused by ROS (Khilfi et al., 2006). Antioxidants interfere with the oxidative processes by scavenging free radicals, chelating free catalytic metals and by acting as electron donor (Gulcin et al., 2005). More often than not, the endogenous antioxidant mechanisms may be overwhelmed in several diseased conditions thereby necessitating the supplementation of antioxidant levels through intake of dietary and plant derived antioxidants (Terao et al., 1994). Plant-based antioxidants are now preferred to the synthetic ones because of safety concerns (Grice, 1986; Wichi, 1988). These factors have inspired the widespread screening of plants for possible medicinal and antioxidant properties, the isolation and characterization of diverse phytochemicals and the development and utilization of antioxidants of natural origin (Jayaprakasha et al., 2001; Gulcin et al., 2002). It is believed that the phytochemical composition and the in-vitro antioxidant activity of a plant might give a clue to its therapeutic potential.

![Fig 1: Synsepalum dulcificum leaves (George and Diejun, 2011).](image)

The miracle fruit shrub, Synsepalum dulcificum Daniell (Sapotaceae) is indigenous to tropical West Africa. The fruit berries of the plant have unusual taste modifying properties of allowing sour substances to taste amazingly sweet after the inside of the mouth has been thoroughly exposed to the fruit’s mucilaginous pulp (Brouwer et al., 1968). Miracle fruit has been studied to improve insulin resistance induced by fructose-rich chow in rats (Chen et al., 2006). Researchers also reported that the skin contributed 43.96% of free phenolic compounds with 15.91% of freeze-dried solids due to its high phenolic content. They also observed that the free flavonoid content in the skin was tremendously higher than that in the seed and pulp and that the skin contributed about 52% of total flavonoid and 15.91% of dried solids (George and Diejun, 2011). However to the best of our knowledge the antioxidant activity of the methanol extract of Synsepalum dulcificum leaves has not been evaluated, hence this study.

**MATERIALS AND METHODS**

**Chemicals**

Acetonitrile, formic acid, gallic acid, chlorogenic acid, ellagic acid and caffeic acid were purchased from Merck (Darmstadt, Germany). Quercetin, quercuritin, isoquercuritin, rutin, catechin, epicatechin and kaempferol, DPPH (2,2-diphenyl-1-picrylhydrazyl) radical, gallic acid, ascorbic acid, quercurin and Folin-Ciocalteau reagent were obtained from Sigma Chemical Co. (St. Louis, MO, USA). All other chemicals and reagents used were of analytical grade.

**Plant material and extraction**

*S. dulcificum* leaves were collected from a farmland in Iseyin, Oyo-state in South-Western Nigeria. The leaves were identified and a voucher number- UIH-22457 was obtained for the leaf at Botany Department University of Ibadan, Nigeria. The leaves were air-dried for three weeks and pulverized. 700g of the pulverized sample was extracted in 80% methanol by maceration for 72 hours. The methanol extract was concentrated in a rotary evaporator and lyophilized. A total yield of the extract (98.7g, 14.1%) was obtained and preserved for further use.

**Quantification of compounds by HPLC-DAD**

Reverse phase chromatographic analyses were carried out under gradient conditions using C$_{18}$ column (4.6 mm x 250 mm) packed with 5μm diameter particles. The mobile phase was water containing 1% formic acid (A) and acetonitrile (B), and the composition gradient was: 13% of B until 10 min and changed to obtain 20%, 30%, 50%, 60%, 70%, 20% and 10% B at 20, 30, 40, 50, 60, 70 and 80 min, respectively (Boligon et al., 2012) with slight modifications. Synsepalum dulcificum (leaves) methanolic extract was analyzed dissolved in ethanol at a concentration of 20 mg/mL. The presence of eleven antioxidants compounds was investigated, namely, gallic acid, chlorogenic acid, caffeic acid, ellagic acid, catechin, epicatechin, quercetin, quercitrin, isoquercitrin, rutin and kaempferol. Identification of these compounds was performed by comparing their retention time and UV absorption spectrum with those of the commercial standards. The flow rate was 0.7 ml/min, injection volume 50 μl and the
wavelength were 254 nm for gallic acid, 280 nm for catechin and epicatechin, 327 nm for caffeic, ellagic and chlorogenic acids, and 366 nm for quercetin, isoquercitrin, quercitrin, rutin and kaempferol. All the samples and mobile phase were filtered through 0.45 μm membrane filter (Millipore) and then degassed by ultrasonic bath prior to use. Stock solutions of standards references were prepared in the HPLC mobile phase at a concentration range of 0.030 – 0.250 mg/ml for kaempferol, quercetin, isoquercitrin, quercitrin, catechin, epicatechin and rutin; and 0.030 – 0.250 mg/ml for gallic, caffeic, ellagic and chlorogenic acids. The chromatography peaks were confirmed by comparing its retention time with those of reference standards and by DAD spectra (200 to 400 nm). All chromatography operations were carried out at ambient temperature and in triplicate. The limit of detection (LOD) and limit of quantification (LOQ) were calculated based on the standard deviation of the responses and the slope using three independent analytical curves. LOD and LOQ were calculated as 3.3 and 10 σ/S, respectively, where σ is the standard deviation of the response and S is the slope of the calibration curve (Boligon et al., 2013).

DPPH (1, 1, diphenyl 2-picryl hydrazyl) assay

The antioxidant activity by DPPH assay was assessed according to Molyneux method (Molyneux, 2004). To 1 ml of various concentration of the extract 1 ml of DPPH 0.1mM was added in a test tube. Tannic acid was used as the standard for comparison. After incubation for 30 mins in dark at room temperature, absorbance was recorded at 517 nm. The percent DPPH radical scavenging was calculated with the equation:

% DPPH radical scavenging = [(Absorbance control – Absorbance sample)/ Absorbance control] × 100.

ABTS radical scavenging activity

A TEAC assay was conducted based on the method of Ramos et al., (1999). The ABTS aqueous solution (7 mM) was oxidized with potassium peroxodisulfate (2.45 mM) for 16 hours in the dark at room temperature. The ABTS⁺ solution was diluted with 95% ethanol to an absorbance of 0.75 ± 0.05 at 734 nm. An aliquot (20 μL) of each sample (125 μg/mL) was mixed with 180 μL ABTS⁺ solution and the absorbance was read at 734 nm after 1 min. Trolox was used as a reference standard. The percent ABTS⁺ radical scavenging was calculated with the equation:

% ABTS⁺ radical scavenging = [(Absorbance control – Absorbance sample)/ Absorbance control] × 100.

Nitric oxide scavenging activity

The nitric oxide scavenging activity was evaluated according to the modified method of Sreejayan and Rao (1997). Various concentrations of extract was prepared. Sodium nitroprusside (2.5 mL, 10 mM) in phosphate buffered saline (PBS) was added to 0.5 mL different concentrations of extracts. The reaction mixture was incubated at 25°C for 150 min. After incubation, 0.5 mL aliquot was removed and 0.5 mL of Griess reagent: (1% (w/v) sulfanilamide, 2% (v/v) H₃PO₄and 0.1% (w/v) naphthylethylene diamine hydrochloride) was added. The absorbance was measured at 546 nm. Ascorbic acid was used as reference standard and was treated the same way as that of fractions. Sodium nitroprusside in PBS (2 mL) was used as control. The nitric oxide radicals scavenging activity of the extracts and ascorbic acid was calculated according to the following equation:

Percentage of inhibition = [Acontrol – A] / Acontrol × 100, where A is the absorbance of sodium nitroprusside in PBS and A1 is the absorbance in the presence of the fractions and ascorbic acid.

Iron chelating ability assay

The in vitro Fe²⁺ chelating ability of plant extract was assayed according to the method of Minnoti and Aust, 1987 with slight modification (Minnoti and Aust, 1987). Briefly, 900 μl of aqueous FeSO₄ (500 μM) and 150μl of extract were incubated for 5min at room temperature, 78 μl of 1,10-phenanthroline (0.25 % ,w/v , aqueous) was added. The absorbance of the orange colour solution was read at 510nm with a spectrophotometer. The principle of the assay is based on disruption of o-phenanthroline - Fe²⁺ complex in the presence of chelating agent. The in vitro of Fe²⁺ chelating ability of the sample was calculated by using the following formula:

Chelating ability (%) = (Acontrol - A sample)/ Acontrol x 100. Where;
Acontrol = The absorbance of the control (reaction mixture in the absence of sample) (FeSO₄ alone).
A sample = The absorbance of the reaction mixture (sample, FeSO₄ and 1,10-phenanthroline)

Reductive potential

Reducing power of the extracts were measured by the direct reduction of Fe³⁺−CN⁻ to Fe²⁺−CN⁻ and was determined by absorbance measurement of the formation of the Perl’s Prussian Blue complex following the addition of excess Fe³⁺−(Oyaziu, 1986), as described previously (Gulcin 2006b). Different concentrations of extracts in 0.5mL of distilled water were mixed with 1.25 mL of 0.2 M, pH 6.6 sodium phosphate buffer and 1.25 mL of potassium ferricyanide [K₄Fe(CN)₆] (1%) the mixture was incubated at 50 °C for 20 min. After 20 min incubation, the reaction mixture was acidified with 1.25 mL of trichloroacetic acid (10%). Finally, 0.5 mL of FeCl₃ (0.1%) was added to this solution and the absorbance was measured at 700 nm in a spectrophotometer. Increased absorbance of the reaction mixture indicates greater reduction capability (Gulcin, 2006a).

Hydroxyl radical scavenging activity

The hydroxyl radical (‘OH) scavenging activity was measured by the method of Jin et al. (1996). The hydroxyl radical was generated in a mixture of 1.0 mL of 0.75 mM 1,10-phenanthroline, 2.0 mL of 0.2 M sodium phosphate buffer (pH 7.4), 1.0 mL of 0.75 mM FeSO₄ and 1.0 mL of H₂O₂ (0.01%, v/v). After addition of 1.0 MI sample solution, the mixture was incubated at 37°C for 30 min. Then, the absorbance of the mixture at 536 nm was measured. Deionized water and VC were used as
the blank and positive control respectively. The scavenging activity on OH was calculated by the following equation:

\[ \text{Scavenging activity (\%)} = \left( \frac{\text{Abs}_{\text{sample}} - \text{Abs}_{\text{blank}}}{\text{Abs}_{0} - \text{Abs}_{\text{blank}}} \right) \times 100 \]

where Abs0 is the absorbance of the deionized water instead of H2O2 and sample in the assay system.

**Inhibition of lipid peroxidation assay**

In vitro inhibition of lipid peroxidation was estimated according to the method of Ruberto and Baratta (2000). In this assay, egg yolk homogenate served as lipid rich medium, and FeSO4 acts as initiator of lipid peroxidation. Briefly, 50μl of plant extract was mixed with 0.25ml 10% egg yolk. This was followed by the addition of 10μl FeSO4 (0.07M, aqueous). The mixture was incubated at room temperature for 30min. This was followed by the addition of 0.75ml of glacial acetic acid (5%, v/v aqueous) and 0.75ml of thiobarbituric acid 0.8% in Sodium dodecylsulphate (SDS) 1.1%. The mixture was incubated in a boiling water bath (90°C) for 60 min, cooled and centrifuged at 3000rpm. One milliliter (1ml) of the pink colour supernant was read at 532 nm on a spectrophotometer.

In vitro inhibition of lipid peroxidation = [(Acontrol – Asample)/Acontrol] X 100

**Determination of total phenolic content**

The total phenolic content of the extract was determined according to the method described elsewhere (Hung et al., 2001). In this assay, the phenolic group present in plant extract interacts with Folin – Ciocalteau in alkaline medium using Na2CO3 solution giving a blue colour, which has maximum absorption at 685nm and correlates with total phenolic content. 0.1ml of aqueous extract of the plant of different concentrations (2 – 10%) was rapidly mixed with 0.1ml of Folin Ciocalteau reagent followed by the addition of 0.3ml sodium carbonate(15%,w/v) solution. The mixture was incubated in the dark for 30mins. The absorbance of the blue colour was read at 760 nm after 30mins on a spectrophotometer. The total phenolic content was extrapolated from a standard curve using tannic acid (graded doses, 50 – 250μg/ml) as a standard phenol.

**Determination of total flavonoid**

Total flavonoid of the leaf extract was determined using the method described by Kumaran and Karunakaran, (2007) with slight modification. Briefly, 0.5 ml of extract solution (1 mg/ml) and standard (quercetin) at different concentrations were taken in test tubes. 3.0 ml of methanol followed by 0.1 ml of 10% aluminum chloride solution was added into the test tubes. Two hundred ml of 1M potassium acetate solution was added to the mixtures in the test tubes. Furthermore, each reaction test tube was then immediately diluted with 2.8 ml of distilled water and mixed to incubate for 30 min at room temperature to complete reaction. The absorbance of pink colored solution was noted at 415 nm using a spectrophotometer against blank methanol. TFC of the extract was expressed as quercetin equivalents (QE) after calculation using the following equation: C= (cV)/m, where, C =total flavonoid contents, mg/g plant extract in QE, c = concentration of quercetin obtained from calibration curve (mg/ml), V =the volume of the sample solution (ml), m= weight of the sample (g). All tests were conducted in triplicate.

**Determination of mineral composition**

Copper, zinc, magnesium, lead, manganese, calcium and iron content of MSD were analyzed using atomic absorption spectrophotometer (Buck 210 VGD), while sodium and potassium content were analyzed using flame photometer (Gulfex medical and scientific, England).

**Statistical analysis**

All statistical analyses were performed using Microsoft excel package 2013 version. Results are expressed as mean± standard deviation. Independent sample t-test was used for data analysis. Linear regression analysis was used to calculate the IC50 values.

**DISCUSSION**

Before the advent of modern medicine herbs and herbal products were the mainstay of treatments around the world. The phytochemicals present in commonly consumed plant foods are normally non-toxic and have the potential for preventing chronic diseases. The plant extracts encompass high concentration of flavonoids and phenolic compounds. As potent antioxidants, flavonoids are especially important for protection against human diseases. The multiple properties of these phytochemicals have made them more attractive, as they can modulate various aspects of diseases like lipid peroxidation involved in atherogenesis, thrombosis, carcinogenesis, hepatotoxicity and a variety of disease conditions (Tiwari, 2001). Table 1 shows the phytochemical composition of the methanol extract of S. dulcificum leaves. Flavonoids, saponins, terpenoids and cardiac glycosides were present in the extract. Our result as presented in table 2 also showed that the total phenols and total flavonoids were estimated to be 58.67 mg/g TAE and 13.01 mg/g QE respectively.

| Table 1: Phytochemical screening of methanol leaf extract of Synsepalum dulcificum. |
|------------------|------------------|
| Phytochemical     | Result           |
| Saponins          | +ve              |
| Tannins           | -ve              |
| Flavonoids        | +ve              |
| Alkaloids         | -ve              |
| Terpenoids        | +ve              |
| Anthraquinones    | -ve              |
| Cardiac glycosides| +ve              |
| Steroids          | -ve              |

<table>
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<tr>
<th>Table 2: Total phenol and total flavonoids content of methanol leaf extract of Synsepalum dulcificum.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total phenol</td>
</tr>
<tr>
<td>Total flavonoids</td>
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</table>
The evaluation of the antioxidant activity of medicinal plants has become a key component of the processes employed in unraveling the therapeutic potentials of such plants. Researchers opined that no single assay accurately reflects the mechanism of action of all radical sources or all antioxidants in a complex system (Prior et al., 2005) and that two or more methods should be employed in order to evaluate the total antioxidant activity (Ilhami et al., 2005) due to various oxidative processes.

DPPH radical scavenging activity is usually used to estimate the in-vitro antioxidant activity of natural compounds or plant extracts (Philips et al., 2010). As shown in figure 3 The DPPH scavenging activity of methanolic extract of Synsepalum dulcificum leaves compared favorably with tannic acid at all the concentrations tested. At the highest concentration- 400μg/ml the extract showed 91.84% inhibition while the standard antioxidant showed 93.53% inhibition of DPPH radical. The result also showed that the extract exhibited a dose-dependent inhibition of DPPH radical.

Chelating agents form σ-bonds with a metals and are effective as secondary antioxidants because they reduce the redox potential, thereby stabilizing the oxidized form of the metal ion (Kumaran and Karunakaran, 2006). Figure 6 shows the percentage iron-chelating activity of our extract. The result shows that though the EDTA which was used as standard showed a better iron-chelating ability than the extract at all the doses tested, the extract still showed appreciable activity in this regard. At 400 μg/ml the extract and standard showed 48.86% and 60.52% iron-chelating activity respectively.

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The reducing capacity of a compound may serve as a significant indicator of its potential antioxidant activity. Most antioxidants and pharmacologically therapeutic agents used in the treatment of oxidative stress related diseases have been shown to
have strong reducing power (Amin and Razieh, 2007). The result in figure 7 shows that the methanol extract of *Synsepalum dulcificum* leaves showed an appreciable reducing power when compared to quercetin which was used as the standard antioxidant. At 400μg/ml the extract and standard showed absorbance values of 1.21 and 1.80 respectively.

Lipid peroxidation in biological systems has long been thought to be a toxicological phenomenon that can lead to various pathological consequences (Hochestein and Attalah, 1988). Lipid peroxidation is caused by the generation of free radicals from a variety of sources including organic hydroperoxides, redox cycling compounds and iron-containing compounds. The TBARS assay has been used to measure the degree of lipid peroxidation. TBA reacts specifically with malondialdehyde (MDA), a secondary product of lipid peroxidation to give a red chromogen, which may then be determined spectrophotometrically (Giri et al., 2010). In this study, it was observed that methanol extract of SD leaves showed a moderate inhibition of lipid peroxidation. Our result as presented in figure 9 shows that the extract has a dose dependent inhibition of lipid peroxidation with the highest concentration tested – 400μg/ml showing a percentage inhibition of 48.79% while ascorbic acid had a percentage inhibition of 74.65% at the same concentration.
The IC₅₀ of the extract for DPPH, NO, OH, and ABTS⁺ radicals scavenging assays were 139.45 μg/ml, 119.17 μg/ml, 147.65 μg/ml, and 135.83 μg/ml respectively, while its IC₅₀ for iron chelation and inhibition of lipid peroxidation were 345.03 μg/ml and 164.19 μg/ml respectively.

HPLC/DAD fingerprint should be considered to evaluate the quality of herbal medicines all over the world considering the fact that multiple constituents are present in herbal medicines and its product (Coppens et al., 1983). Our results as shown in tables 3 and 4 show that the extract has a high level of polyphenols in the order flavonoids (85.96 mg/g / 8.58%) > phenolic acids (63.5 mg/g / 6.33%) > tannins (23.87 mg/g / 2.38%).

Table 3: Composition of methanol leaf extract of Synsepalum dulcificum.

<table>
<thead>
<tr>
<th>Compounds</th>
<th>Synsepalum dulcificum LOD µg/mL</th>
<th>LOD µg/mL</th>
<th>LOQ µg/mL</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gallic acid</td>
<td>6.18 ± 0.03¹</td>
<td>0.61</td>
<td>0.024</td>
</tr>
<tr>
<td>Catechin</td>
<td>5.95 ± 0.01¹</td>
<td>0.59</td>
<td>0.015</td>
</tr>
<tr>
<td>Chlorogenic acid</td>
<td>20.93 ± 0.01³</td>
<td>2.09</td>
<td>0.007</td>
</tr>
<tr>
<td>Caffeic acid</td>
<td>14.82 ± 0.00³</td>
<td>1.48</td>
<td>0.035</td>
</tr>
<tr>
<td>Ellagic acid</td>
<td>21.57 ± 0.01³</td>
<td>2.15</td>
<td>0.028</td>
</tr>
<tr>
<td>Epicatechin</td>
<td>17.92 ± 0.01³</td>
<td>1.79</td>
<td>0.021</td>
</tr>
<tr>
<td>Rutin</td>
<td>30.65 ± 0.03³</td>
<td>3.06</td>
<td>0.042</td>
</tr>
<tr>
<td>Isoquercitrin</td>
<td>15.03 ± 0.02²</td>
<td>1.50</td>
<td>0.010</td>
</tr>
<tr>
<td>Quercitrin</td>
<td>12.76 ± 0.01³</td>
<td>1.27</td>
<td>0.019</td>
</tr>
<tr>
<td>Quercetin</td>
<td>21.90 ± 0.01³</td>
<td>2.19</td>
<td>0.026</td>
</tr>
<tr>
<td>Kaempferol</td>
<td>5.62 ± 0.01¹</td>
<td>0.56</td>
<td>0.013</td>
</tr>
</tbody>
</table>

Results are expressed as mean ± standard deviations (SD) of three determinations. Averages followed by different letters differ by Tukey test at p < 0.01.

Table 4: Carotenoids compositions of methanol leaf extract Synsepalum dulcificum.

<table>
<thead>
<tr>
<th>Carotenoids</th>
<th>S. dulcificum mg/g</th>
<th>S. dulcificum %</th>
<th>LOD µg/mL</th>
<th>LOQ µg/mL</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tocopherol</td>
<td>2.06 ± 0.01¹</td>
<td>0.005</td>
<td>0.001</td>
<td>0.015</td>
</tr>
<tr>
<td>β-Carotene</td>
<td>2.93 ± 0.04¹</td>
<td>0.010</td>
<td>0.010</td>
<td>0.030</td>
</tr>
<tr>
<td>Lycopene</td>
<td>0.57 ± 0.01³</td>
<td>0.005</td>
<td>0.001</td>
<td>0.010</td>
</tr>
</tbody>
</table>

Results are expressed as mean ± standard deviations (SD) of three determinations. Averages followed by different letters differ by Tukey test at p < 0.01.

Among the flavonoids rutin was the most abundant followed by quercetin, isoquercitrin, quercitrin and kaempferol in that order. Ellagic acid is the most abundant among the phenolic acids present in the extract followed by chlorogenic acid, caffeic acid, and gallic acid in that order. For tannins our result also shows that the extract contains a higher level of epicatechin than catechin. The presence of carotenoids was also established in the extract. Figure 2 shows the chromatogram indicating the peaks of the polyphenols that were detected in the extract. HPLC is a sensitive, accurate and specific method for the separation and identification of phenolic acids, flavonoid glycosides and aglycones in plants. (Praven, 2012; Ana et al., 2009). Moreover, many sources indicate good correlations between phenolics content and antioxidant capacity evaluated by different techniques, including HPLC (Benzie and Strain, 1999; Brand-Williams et al., 1995), and Praven, (2012) asserted that plants exhibited medicinal properties due to the presence of flavonoids in it. We therefore opine that the presence of these plant polyphenols may contribute in part to the significant antioxidant properties of MSD as observed in this study.

The medicinal value of the plants lies in the chemical substances that produce a definite physiological action on human body (Hill, 1952). The distribution of minerals in common medicinal plants has important application for the health of people in addition to the basic need of developing countries.

Therefore there is need to evaluate the local herbs for mineral and nutrient composition, so as to determine their potential as indigenous source of medicine (Rahila et al., 1994). Our result in table 5 shows that presence of some micro and macro elements. Potassium, calcium, magnesium and sodium are present in appreciable amounts while lead, a heavy metal is present in trace quantity.

Table 5: Mineral composition of methanol leaf extract of Synsepalum dulcificum.

<table>
<thead>
<tr>
<th>Metals</th>
<th>mg/100g</th>
</tr>
</thead>
<tbody>
<tr>
<td>Copper</td>
<td>1.10</td>
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<tr>
<td>Zinc</td>
<td>4.16</td>
</tr>
<tr>
<td>Magnesium</td>
<td>291.0</td>
</tr>
<tr>
<td>Lead</td>
<td>0.01</td>
</tr>
<tr>
<td>Manganese</td>
<td>8.70</td>
</tr>
<tr>
<td>Calcium</td>
<td>830.0</td>
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<tr>
<td>Iron</td>
<td>26.30</td>
</tr>
<tr>
<td>Sodium</td>
<td>102.0</td>
</tr>
<tr>
<td>Potassium</td>
<td>1850.0</td>
</tr>
</tbody>
</table>

CONCLUSION

From our study it could be safely concluded that the methanolic extract of Synsepalum dulcificum leaves has appreciable in vitro antioxidant activity and mineral content. This is a clear pointer to the possibility of the plant having potential health benefits especially in diseases in which oxidants have been established as the culprit. To confirm this possibility however, more research needs to be carried out using models of the diseases in question.
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Conflicts of Interest: There are no conflicts of interest.

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