

Median lethal dose (LD50) and cytotoxicity of Adriamycin in female albino mice

Rehab M. Mosaad¹, Amany Samir¹, Hassan M. Ibrahim^{2*}

¹Lecturer at Zoology Department, Faculty of Women for Arts, Science and Education, Ain Shams University, Cairo, Egypt.

²Researcher at Textile Research Division, National Research Centre, 33 El Bohouth st. (Former El Tahrir St.), Dokki, Giza, Egypt.

ARTICLE INFO

Article history:

Received on: 09/03/2016

Accepted on: 07/05/2016

Available online: 30/03/2017

Key words:

Median lethal dose, cytotoxicity, Adriamycin, female albino mice.

ABSTRACT

The median lethal dose of Adriamycin and its cytotoxicity in three vital organs (heart, liver and kidney) was studied. The median lethal dose (LD50) was 56.875mg/kg body. Weight (b.wt). High doses of Adriamycin induced significant increases in heart tissues enzymes (lactate dehydrogenase (LDH), alkaline phosphatase (ALP) and Creatine Kinase-MB isoenzyme (CKMB)). Hepatotoxicity appeared in significant increment in liver function enzymes (aspartate aminotransferase (AST) and Alanine aminotransferase (ALT) activities) and lipid profile (cholesterol and triglycerides) and significant reduction in total protein and albumin. Moreover, urea and creatinine have recorded significant increases in kidney tissues after interperitoneal administration of high doses of Adriamycin (100, 90 and 80 mg/kg b.wt.).

INTRODUCTION

Adriamycin is a quinone containing anticancer and antibiotic that is used to treat many solid malignancies and lymphomas (Quiles *et al.*, 2002). Adriamycin have severe toxic effects towards heart, kidney and liver so that its clinical efficacy is restricted(Quiles *et al.*, 2002; Chae *et al.*, 2005; Kim *et al.*, 2006; Zahraei and Rabbani-Chadegani, 2007; Othman *et al.*, 2008). The toxicity effect of Adriamycin on heart and liver are condensation, myofibrils, mitochondrial swelling and loss disruption in heart and lipid droplets accumulation, polymorphic mitochondria, and vacuolization of cytoplasm in liver. A nephrotic syndrome is induced by a single high-dose injection of Adriamycin (ADR). Heavy proteinuria, hypoalbuminemia, hypercoagulability, and hyperlipemia with hypercholesterolemia were used to characterize ADR(Deepa and Varalakshmi, 2005b).

Many mechanisms are considered to illustrate the cytotoxic effects of Adriamycin such as inhibition of DNA synthesis, alkylation and binding of DNA, interference of DNA strand separation and helicase activity, effects of direct membrane, cross-linking of lipid peroxidation DNA and free radical formation (Deepa and Varalakshmi, 2006).

The aim of the present study is to determine the (Median lethal dose) LD₅₀ of Adriamycin intraperitoneal injection and cytotoxicity of 80, 90, and 100 mg/kg b.wt. in female albino mice.

MATERIAL AND METHODS

Experimental animals

84 adult female albino mice were obtained from the animal house, veterinary division, NRC. Their weights ranged between (20 and -30 g). Mice were housed in plastic cages, each cage contained eight mice. Animals were kept under controlled temperature of 25 ± 2 °C for 12 hours under light and 12 hours dark cycle throughout the experiment. A commercial pelleted diet was used during the experiment. Food and water were available *ad libitum*.

* Corresponding Author

Hassan M. Ibrahim, Researcher at Textile Research Division, National Research Centre, 33 El Bohouth st. (Former El Tahrir St.), Dokki, Giza, Egypt. Email: hmaibrahim@gmail.com

Drugs

Adriamycin as Adricin was purchased from EIMC United Pharmaceuticals, Badr City, Cairo, A.R.E.

Treatment and dosage

An approximate LD50 was initially determined in pilot study by a so called "staircase method" using a small number of animals (2 for each dose) with increasing doses of Adriamycin. Five doses of 80, 90, 100, 110 and 120mg/kg b.wt., were given to 5 groups of mice (8 in each) for the determination of intraperitoneal LD50 in female mice (Table 1).

Animals were observed for the 2, 6 and 24 hours for any toxic symptoms. After 24 hours, number of died animals was counted in each group and transformed to probits and then LD50 determined by the method of Karber (Shetty Akhila and Alwar, 2007). The cytotoxicity of 80,90 and 100 mg/kg b.wt., was determined in three tissues (heart, Liver and kidney).

Determination of biochemical parameters:

After 24hrs the surviving animals were quickly sacrificed by decapitation and heart, liver and kidney organs were collected for determination the biochemical parameters. The organs were weighed and homogenized in 10 m mol/L phosphate buffer saline (10% W/V) of PH 7.4 and centrifuged. The supernatant used for determination of LDH, ALP, CKMB in heart tissues, ALT, AST, TP, albumin, cholesterol, triglycerides in liver tissues and urea and creatinine in kidney tissues.

In heart tissue, lactate dehydrogenase (LDH) was determined according to (Young, 2000; Young and Friedman, 2001) and ALP was measured according to (Belfield and Goldberg, 1971) while creatine kinase-MB isoenzyme (CKMB) was measured in heart according to (Stein, 1998). aspartate aminotransferase (AST) and Alanine aminotransferase (ALT) activities in liver tissue were measured spectrophotometrically according to the method of (Murray, 1984; Murray, 1989), Total Protein concentration was measured by the method of (Gornall *et al.*, 1949), Albumin levels were measured according to the method of (Doumas *et al.*, 1971), The measurement of triglycerides levels using assay kit according to (Fossati and Prencipe, 1982) and cholesterol was measured according to (Richmond, 1973; Allain *et al.*, 1974). Moreover, urea content was performed in kidney tissue according to the procedure of (Fawcett and Scott, 1960) and creatinine concentration measured according to (Schirmeister *et al.*, 1964).

Statistical analysis:

Reported values for liver, heart and kidney parameters represented in the table 2 expressed as means \pm SE. Statistical analysis evaluated by one-way ANOVA. Once a significant F test obtained, LSD comparisons performed to assess the significance of differences among various treatment groups. Statistical Processor System Support "SPSS" for Windows software, Release 12.0 (SPSS, Chicago,IL) was used.

RESULTS AND DISCUSSION

Among the treated animals with high doses of Adriamycin, it was found that some animals were died immediately after few seconds. But the rest of the treated animals were drowsy and less responsive. The severity of these effects were related to the dose concentration. However, few numbers of survivors had recovered from these symptoms after 24 hours.

Table 1: Results of the lethal doses of Adriamycin for the determination of the LD50 after intraperitoneal injection in female mice (n=8)

group	Dose mg/kg	No. of animal dead	dose difference (a)	mean mortality (b)	probit (a*b)
1	control	0			
2	80	1	0		
3	90	2	10	1.5	15
4	100	4	10	3	30
5	110	8	10	6	60
6	120	8	10	8	80

LD50 = the apparent least dose lethal to all in a group- sum (a*b)/n
Where n is the number of animals in each group, (a) is the dose difference and (b) is the mean mortality =mortality in second group +1st group /2 (Table 1)

$$LD_{50} = 80 - 185/8 = 80 - 23.125 = 56.875 \text{ mg/kg b.wt.}$$

The LD₅₀ of Adriamycin in adult female mice was found to be 56.875 mg/kg b.wt. after intraperitoneal injection (Table 1).

Adriamycin, is anthracycline isolated Streptomyces, used as antibiotic and antitumor (Eliaz *et al.*, 2004; Al-Ghamdi, 2008). Waldemar *et al.* (1993) indicated that LD50 of Adriamycin was 25.7 mg/kg. While, the LD50 reported by Bertezoli *et al.* (1985) was 11.98 mg/kg in mice treated intraperitoneally. Also, (Kratz *et al.*, 2007) indicated that the LD50 of intravenous administration of Adriamycin in CD-1 mice was 12mg/kg, while it was 23.4 and 45.9 mg/kg for males and females Sprague Dawley rats respectively.

A single dose of the encapsulated form of Adriamycin was less toxic (LD50 of 32mg/kg) than free Adriamycin (LD₅₀ of 17 mg/kg). In addition, The LD50 of Adriamycin hydrochloride nanoliposome was 31.69 mg/kg in kuming mice (Xie *et al.*, 2013). This difference in the reported results might be due to route of administration, animal sex, method used for the estimation of LD50 and different derivative of Adriamycin.

Adriamycin as antibiotic and anticancer was successful in solid tumors, lymphomas and leukemia's treatment (Pal and Sil, 2012; Elsherbiny and El-Sherbiny, 2014). The therapeutic efficacy of Adriamycin have been severely limited by the toxic side effects of the drug on heart as well as other organs including the liver, heart, brain and kidney (Elsherbiny and El-Sherbiny, 2014; Wang *et al.*, 2014).

The data in this study present the effect of i.p. administration of three acute doses (80, 90 and 100 mg/kg b.wt.) of Adriamycin on heart, liver and kidney functions of female albino mice after 24hrs of one dose injection.

In our study, Adriamycin produced significant increases in heart tissues lactate dehydrogenase, alkaline phosphatase and Creatine Kinase-MB isoenzyme (CKMB) Table (2).

Table 2: Effect of i.p. administration of single dose Adriamycin(80, 90 and 100mg/kg b.wt.) on the heart, liver and kidney functions of female albino mice.

	Control	Adriamycin (80mg/kg b.wt.)	Adriamycin (90mg/kg b.wt.)	Adriamycin (100mg/kg b.wt.)
LDH U/g tissue	1.610±0.0368	3.389±0.125a	5.451±0.104ab	4.777±0.168ab
Alk.U/g tissue	0.062±0.002	0.172±0.006a	0.271±0.005ab	0.252±0.016ab
CKMB U/g tissue	2.284±0.079	4.414±0.171a	5.710±0.236ab	5.667±0.246ab
ALT U/g tissue	0.0149±0.0004	0.232±0.010a	0.255±0.011a	0.349±0.015abc
AST U/g tissue	0.065±0.003	0.244±0.028a	0.302±0.030a	0.436±0.008ab
TP mg/g tissue	6.005±0.195	3.198±0.170a	2.799±0.173a	2.124±0.145a
Album.mg/g tissue	14.285±0.701	10.672±0.173a	9.160±0.278a	7.228±0.163ab
TG.mg/g tissue	11.862±0.386	25.208±0.410a	25.154±0.481a	26.064±1.316a
Chol.mg/g tissue	125.570±3.782	211.76±4.646a	496.45±17.55ab	406.898±11.286abc
Urea mg/g tissue	0.424±0.016	0.551±0.019a	0.583±0.019a	0.599±0.019a
Creat.mg/g tissue	0.162±0.006	0.238±0.010a	0.374±0.011ab	0.375±0.005ab

Values are means of 8 mice ± SE. in group (1), 7 mice ± SE in group (2), 6 mice± SE in group (3) and means of 4 rats± SE in group (4) a = significant change from control, b = significant change from Adriamycin (80mg/kg) rats and c = significant change from Adriamycin (90mg/kg) rats at $p \leq 0.05$.

Ibrahim *et al.* (2009) examined the effect of accumulative Adriamycin dose (15mg/kg) and they found a significant rise in LDH and CK-MB enzyme activities which act as important markers of heart disease (Ibrahim *et al.*, 2009).

There are several mechanisms that illustrate cardiotoxicity results from Adriamycin, including membrane lipid peroxidation, mitochondrial damage, free radical generation, lipid peroxidation (LPO), and iron-dependent oxidative damage to macromolecules (Singh *et al.*, 2015). There are significant rise in malondialdehyde level (MDA) accompanied with glutathione (GSH) activity inhibition in heart followed by the activity of catalase (CAT) with a significant elevation in lactate dehydrogenase (LDH) and aspartate transaminase (AST) (Nazmi *et al.*, 2013).

Adriamycin myocardial impairment involve myocyte apoptosis via oxidative free radical formation (Deepa and Varalakshmi, 2006). Also Adriamycin increase radicals of superoxide and NO in heart tissue then form toxic free radicals their (Prahalthan *et al.*, 2005).

Adriamycin can impair normal liver by generating oxidative stress, decreases in the antioxidation capacity of the liver and the liver weight (Wang *et al.*, 2014).

In the present study, Adriamycin induced significant increases ($p < 0.05$) in liver tissues enzymes (ALT and AST) and significantly decreases in liver total protein and albumin contents. In addition, the treatment of single different doses of Adriamycin showed significant increases in liver triglycerides and cholesterol contents (Table 2).

Single i.p. dose of 15mg/kg of Adriamycin caused liver toxicity appeared in an increased in alanine transaminase (ALT) and aspartate transaminase (AST) which attributed to oxidative stress that appears in significant decrease in GSH level and increase in MDA (El-Moselhy and El-Sheikh, 2014). Moreover, a single i.p. dose of 10mg/kg b.wt. into male rats caused elevations in amino transferases and these changes were associated with increases in peroxidation of lipid and protein oxidation as well as decrease in glutathione levels and glutathione-s-transferase while glutathione peroxidase and catalase activity were elevated in liver Adriamycin treated rat (Othman *et al.*, 2008). Adriamycin intravenous injection dose of (7.5 mg/kg) rise the lipids levels on

heart, liver and kidney (cholesterol, triglycerides and LDL cholesterol), and decrease in HDL cholesterol levels (Deepa and Varalakshmi, 2005a; Deepa and Varalakshmi, 2006).

In kidney tissues, the data showed significant increases in urea and creatinine content after the treatment with 15 mg/kg Adriamycin i.p. single dose for 5 days, which cause renal toxicity (El-Moselhy and El-Sheikh, 2014). Repeated injection doses from Adriamycin lead to renal dysfunctions involving elevation in levels of urea and creatinine in blood and albumin in urine which attributed to oxidative stress mechanisms (Elsherbiny and El-Sherbiny, 2014). Also, Adriamycin induced renal toxicity was monitored in kidney via increase in MDA and decrease in GSH levels (Ibrahim *et al.*, 2009; Nazmi *et al.*, 2013). Decreases in GSH cause cellular electrolytes disrupt which lead to renal dysfunctions and renal toxicity (Pal and Sil, 2012). The high levels of oxygen free radicals produced by Adriamycin metabolism cause pathologic alterations in kidneys including, increased level of lipid peroxidation products and apoptosis (Dzięgiel *et al.*, 2002; Chmielewska *et al.*, 2015).

CONCLUSION

In this paper, the median lethal dose (LD₅₀) and cytotoxicity of Adriamycin were evaluated in female albino mice. The median lethal dose was 56.875 mg/kg. Cytotoxicity of 80, 90 and 100mg/kg were examined by the biochemical analysis of heart, liver and kidney tissues. After 24 hrs of one dose, the data recorded sever effects as significant changes in heart and liver enzymes, hepatic total protein and albumin, hepatic lipid profile and kidney function.

ACKNOWLEDGEMENT

The authors gratefully acknowledges National Research Centre, for financial support and for facilities provided through project ID: 10050309.

Conflict of Interests: There are no conflicts of interest.

REFERENCES

Al-Ghamdi SS. N-(2-Hydroxypropyl)-Methacrylamide-Attached Doxorubicin Induces Cytotoxicity to Prostate Cancer Cell Line DU145. Toxicology mechanisms and methods, 2008; 18 (5): 419-424.

- Allain CC, Poon LS, Chan CS, Richmond W and Fu PC. Enzymatic determination of total serum cholesterol. *Clinical chemistry*, 1974; 20 (4): 470-475.
- Belfield A and Goldberg D. Colorimetric determination of alkaline phosphatase activity. *Enzyme*, 1971; 12 (5): 561-568.
- Chae H-J, Kim H-R, Kim D-S, Woo E-R, Cho Y-G and Chae S-W. Saeng-Ji-Hwang has a protective effect on adriamycin-induced cytotoxicity in cardiac muscle cells. *Life sciences*, 2005; 76 (18): 2027-2042.
- Chmielewska M, Symonowicz K, Pula B, Owczarek T, Podhorska-Okolow M, Ugorski M and Dziegiel P. Expression of metallothioneins I and II in kidney of doxorubicin-treated rats. *Experimental and Toxicologic Pathology*, 2015; 67 (4): 297-303.
- Deepa P and Varalakshmi P. Beneficial cardio-renal effects of a low-molecular-weight heparin-derivative on adriamycin-induced glycosaminoglycanuria and tissue lipid abnormalities. *Toxicology*, 2005a; 211 (1): 77-85.
- Deepa P and Varalakshmi P. Biochemical evaluation of the inflammatory changes in cardiac, hepatic and renal tissues of adriamycin-administered rats and the modulatory role of exogenous heparin-derivative treatment. *Chemico-biological interactions*, 2005b; 156 (2): 93-100.
- Deepa P and Varalakshmi P. Influence of a low-molecular-weight heparin derivative on the nitric oxide levels and apoptotic DNA damage in adriamycin-induced cardiac and renal toxicity. *Toxicology*, 2006; 217 (2): 176-183.
- Doumas BT, Watson WA and Biggs HG. Albumin standards and the measurement of serum albumin with bromocresol green. *Clinica chimica acta*, 1971; 31 (1): 87-96.
- Dziegiel P, Suder E, Surowiak P, Jethon Z, Rabczyński J, Januszewska L, Sopol M and Zabel M. Role of exogenous melatonin in reducing the nephrotoxic effect of daunorubicin and doxorubicin in the rat. *Journal of pineal research*, 2002; 33 (2): 95-100.
- El-Moselhy MA and El-Sheikh AA. Protective mechanisms of atorvastatin against doxorubicin-induced hepato-renal toxicity. *Biomedicine & Pharmacotherapy*, 2014; 68 (1): 101-110.
- Eliaz RE, Nir S, Marty C and Szoka FC. Determination and modeling of kinetics of cancer cell killing by doxorubicin and doxorubicin encapsulated in targeted liposomes. *Cancer Research*, 2004; 64 (2): 711-718.
- Elsherbiny NM and El-Sherbiny M. Thymoquinone attenuates Doxorubicin-induced nephrotoxicity in rats: Role of Nrf2 and NOX4. *Chemico-biological interactions*, 2014; 223: 102-108.
- Fawcett J and Scott J. A rapid and precise method for the determination of urea. *Journal of clinical pathology*, 1960; 13 (2): 156-159.
- Fossati P and Prencipe L. Serum triglycerides determined colorimetrically with an enzyme that produces hydrogen peroxide. *Clinical chemistry*, 1982; 28 (10): 2077-2080.
- Gornall AG, Bardawill CJ and David MM. Determination of serum proteins by means of the biuret reaction. *J. biol. Chem.*, 1949; 177 (2): 751-766.
- Ibrahim MA, Ashour OM, Ibrahim YF, El-Bitar HI, Gomaa W and Abdel-Rahim SR. Angiotensin-converting enzyme inhibition and angiotensin AT 1-receptor antagonism equally improve doxorubicin-induced cardiotoxicity and nephrotoxicity. *Pharmacological research*, 2009; 60 (5): 373-381.
- Kim D-S, Kim H-R, Woo E-R, Kwon D-Y, Kim M-S, Chae S-W and Chae H-J. Protective effect of calceolarioside on adriamycin-induced cardiomyocyte toxicity. *European journal of pharmacology*, 2006; 541 (1): 24-32.
- Kratz F, Ehling G, Kauffmann H and Unger C. Acute and repeat-dose toxicity studies of the (6-maleimidocaproyl) hydrazine derivative of doxorubicin (DOXO-EMCH), an albumin-binding prodrug of the anticancer agent doxorubicin. *Human & experimental toxicology*, 2007; 26 (1): 19-35.
- Murray R. Alanine aminotransferase. *Clinical Chemistry: Theory, Analysis, and Correlation*, 2nd ed. Kaplan LA, Pesce AJ, eds. St. Louis: The CV Mosby Company, 1989: 895-898.
- Murray R. Aspartate aminotransferase. *Clinical Chemistry: Theory, analysis and correlation*. Kaplan LA, Pesce AJ (Ed), CV Mosby Company, 1984: 1105-1108.
- Nazmi AS, Ahmad SJ, Pillai KK, Akhtar M, Ahmad A and Najmi AK. Protective effects of Bombyx mori, quercetin and benazepril against doxorubicin induced cardiotoxicity and nephrotoxicity. *Journal of Saudi Chemical Society*, 2013:
- Othman AI, El-Missiry MA, Amer MA and Arafa M. Melatonin controls oxidative stress and modulates iron, ferritin, and transferrin levels in adriamycin treated rats. *Life sciences*, 2008; 83 (15): 563-568.
- Pal S and Sil PC. A 43 kD protein from the leaves of the herb *Cajanus indicus* L. modulates doxorubicin induced nephrotoxicity via MAPKs and both mitochondria dependent and independent pathways. *Biochimie*, 2012; 94 (6): 1356-1367.
- Prahalathan C, Selvakumar E and Varalakshmi P. Lipoic acid ameliorates adriamycin-induced testicular mitochondriopathy. *Reproductive Toxicology*, 2005; 20 (1): 111-116.
- Quiles JL, Huertas JR, Battino M, Mataix J and Ramírez-Tortosa MC. Antioxidant nutrients and adriamycin toxicity. *Toxicology*, 2002; 180 (1): 79-95.
- Richmond W. Preparation and properties of a cholesterol oxidase from *Nocardia* sp. and its application to the enzymatic assay of total cholesterol in serum. *Clinical chemistry*, 1973; 19 (12): 1350-1356.
- Schirmeister J, Willmann H and Kiefer H. Plasma creatinine as rough indicator of renal function. *Deutsche medizinische Wochenschrift* (1946), 1964; 89: 1018.
- Shetty Akhila J and Alwar M. Acute toxicity studies and determination of median lethal dose. *Curr. Sci*, 2007; 93: 917-920.
- Singh P, Sharma R, McElhanon K, Allen CD, Megyesi JK, Beneš H and Singh SP. Sulforaphane protects the heart from doxorubicin-induced toxicity. *Free Radical Biology and Medicine*, 2015; 86: 90-101.
- Stein W. Creatine kinase (total activity), creatine kinase isoenzymes and variants. *Clinical laboratory diagnostics*. Frankfurt: TH-Books Verlagsgesellschaft, 1998: 71-80.
- Wang B, Ma Y, Kong X, Ding X, Gu H, Chu T and Ying W. NAD⁺ administration decreases doxorubicin-induced liver damage of mice by enhancing antioxidation capacity and decreasing DNA damage. *Chemico-biological interactions*, 2014; 212: 65-71.
- Xie M, Chen Y and Wu L. Preparation of doxorubicin-hydrochloride nanoliposomes by ethanol injection-pH gradient method and their safety evaluation. *Journal of Nanoscience and Nanotechnology*, 2013; 13 (1): 216-221.
- Young DS. 2000. Effects of drugs on clinical laboratory tests. AACC press.
- Young DS and Friedman RB. 2001. Effects of disease on clinical laboratory tests. 2. Listing by disease. AACC-Press.
- Zahraei Z and Rabbani-Chadegani A. A comparison of the effect of anticancer drugs, idarubicin and adriamycin, on soluble chromatin. *European journal of pharmacology*, 2007; 575 (1): 28-33.

How to cite this article:

Mosaad RM, Samir A, Ibrahim HM. Median lethal dose (LD50) and cytotoxicity of Adriamycin in female albino mice. *J App Pharm Sci*, 2017; 7 (03): 077-080.