

Modulatory Effect of Wheat Germ Oil on Intestinal Oxidative Stress and DNA damage Induced by Carbon tetrachloride in Mice

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ARTICLE INFO

Article history:

Received on: 14/07/2016

Revised on: 17/08/2016

Accepted on: 02/09/2016

Available online: 28/12/2016

Key words:

CCl₄; wheat germ oil; small intestine; lipid profile; DNA damage.

ABSTRACT

Background/Aim: The liver is continuously linked to the gut *via* the portal vein supply. Gastrointestinal complications are directly associated with liver cirrhosis. Carbon tetrachloride (CCl₄) caused liver toxicity is well documented in animal models, the very fewer search has been carried out on the intestinal damage in case of acute liver injury. Thus, this study aimed to investigate the intestinal alteration and subsequently the potential therapeutic role of Wheat Germ Oil (WGO) in the liver and small intestine after acute administration of CCl₄. **Material and Methods:** Mice were randomly divided into 4 groups; control group, WGO group: received corn oil orally for 2 days then WGO (1400mg/kg) orally for 8 days, CCl₄ group: received CCl₄ orally for 2 days then corn oil for 8 days, CCl₄ + WGO group: received CCl₄ for 2 days then WGO for 8 days. Serum lipid profile, serum lactate dehydrogenase (LDH), intestinal oxidative stress enzymes, histopathological and DNA fragmentation assays were estimated. **Results:** Acute dose (50%, 1 mL/kg body weight) of CCl₄-induced hyperlipidemia, hypocholesteremia, elevation in LDH, malondialdehyde (MDA), nitric oxide (NO) and intestinal DNA damage in addition to the reduction in the intestinal oxidative stress markers and an alteration in the mucosal architecture. On the contrary, WGO administration has the potency to protect not only the liver but also the small intestine in acute CCl₄-induced tissue damage. The valuable effect is chiefly attributed to its mechanism of reducing the lipid profile and suppressing the oxidative stress that caused DNA damage **Conclusion:** WGO administration could markedly improve the liver and the small intestine from the CCl₄ damage and consequently may be used as a therapeutic agent against the hepatic and intestinal toxicity.

INTRODUCTION

Carbon tetrachloride (CCl₄) is commonly used for experimental induction of liver injury in rodents to mimic the oxidative stress situations in many pathophysiological conditions (Zhu *et al.*, 2013; Go *et al.*, 2016). The single acute dose of CCl₄ causes liver toxicity that is characterized by hepatocellular necrosis and steatosis while a chronic dose of CCl₄ causes liver cirrhosis. Generation of free radicals has been observed in many tissues; liver, kidney, intestine, heart, lung, brain, and blood after CCl₄ administration (Dashti *et al.*, 1989). The free radicals generated from CCl₄ and its metabolites induced an impairment of the endoplasmic reticulum (ER) and altered the permeability

of the mitochondrial membrane, which leads to accumulation of lipids, reduction of protein synthesis and overproduction of the oxidative stress (Weber *et al.*, 2003). Oxidative stress exhibits a significant role in the pathogenesis of liver injury and other hepatic alterations through implications between free radicals generated by CCl₄ and lipid peroxidation (Gutiérrez *et al.*, 2010). The toxicity of CCl₄ is originated through its breakdown to the highly reactive trichloromethyl radical (CCl₃·) in the endoplasmic reticulum by cytochrome P₄₅₀ enzymes. CCl₃· which rapidly reacts with oxygen forming trichloromethyl peroxy radical (CCl₃OO·), that interacts with lipids causing lipid peroxidation that leads to hepatotoxicity (Risal *et al.*, 2012). The free radicals produced from CCl₄ not only affect the cellular permeability of the hepatocytes leading to the elevation of hepatic enzymes but also cause deterioration in hepatic function through an elevation in lipid profile such as total lipids, triglycerides, cholesterol and HDL-cholesterol (Essawy *et al.*, 2012).

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The gut and the liver are bidirectionally communicated together through digestion and absorption products, bile, hormones and inflammatory mediators (Bajaj *et al.*, 2012). The correlation between liver damage and the functional integrity of the intestine was documented by (Manevska 1975). As long as the gastrointestinal tract is affected, the mucosal alterations may arise, resulting in portal hypertension during liver cirrhosis (Llovet *et al.*, 1994). Oxygen free radicals generated from CCl₄ play an important role in the damage of gut epithelial cells, which may affect the gut barrier function, facilitate bacterial translocation and induce the release of endotoxin (Ramachandran *et al.*, 2002). Oxidative stress in the intestinal mucosa caused an elevation in xanthine oxidase which is an important source of free radicals in the small intestine (Watkins 2001).

Moreover, numerous herbal antioxidants could protect organs against CCl₄ induced oxidative stress by reducing lipid peroxidation and restoring the antioxidant enzymes activities. The essential oils including WGO have attracted much attention in recent years due to their unique nutritional value (Majzoobi *et al.*, 2016). WGO has a high amount of policosanols, especially octacosanol, which reduces the cholesterol levels and improves the lipid profile (Anwar *et al.*, 2015). WGO plays an important role in decreasing lipid peroxidation by activating the tocopherol redox-system (Leenhardt *et al.*, 2008), as an interesting source of natural antioxidant tocopherols, sterols, and vitamin B complex. In addition, WGO has a high vitamin E and phenolic contents which act as inhibitors of the oxidation processes in body tissues and protect cells against the damaging effects of free radicals generated from CCl₄ (Liu *et al.*, 2015).

Moreover, WGO not only prevents autoxidation of unsaturated fatty acids but also, confers DNA protective effects (Gelmez *et al.*, 2009). Although the effect of WGO on improving liver toxicity is well documented. However, very few research has been carried out on the intestinal alterations in the case of acute liver injury.

Thus, the aim of this study was carried out to investigate the effect of acute oral dose of CCl₄ not only on the liver injury but also on the redox balance of the intestinal mucosa by assessing different levels of the oxidative stress, histopathological changes, and the intestinal DNA damage. It also aims to evaluate the potential role of WGO in preventing the deleterious effects of the CCl₄-induced acute liver injury in mice.

MATERIALS AND METHODS

Animals and chemicals

Male C57 BL/6 mice (6 weeks old, weighing 22-25 g) were obtained from National Research Center (NRC, Giza, Egypt). Mice were randomly grouped and housed in a conventional clean facility. All the experimental procedures were carried out in accordance with international guidelines for the care and use of laboratory animals. Mice were given food and water *ad libitum* and maintained in a friendly environment with a 12 h/12 h light-dark cycle at room temperature (22°C–25°C). Mice were adapted

to laboratory conditions for 7 days before beginning of the experiment. CCl₄, WGO, and all other chemicals were purchased from Sigma-Aldrich Biotechnology (St Louis, MO, USA). Assays kits for the detection of liver lipid profile and oxidative stress markers were purchased from Biodiagnostic (Giza, Egypt).

Induction of liver injury and WGO administration

Induction of liver injury in mice was done by oral administration of CCl₄ (50%, 1 mL/kg body weight) dissolved in corn oil, the dose was chosen to induce liver injury as previously reported (Srivastava *et al.*, 2010). WGO was given to the animals by oral gavage at dose (1400 mg/kg body weight), according to previous literature (Karabacak *et al.*, 2011).

Experimental design

Mice were randomly divided into 4 groups (6 mice/group):

Group 1: **Control group**: mice received 1 mL oral administration of corn oil for 10 days.

Group 2: **WGO group**, mice received corn oil orally for 2 days then WGO (1400mg/kg b.w) for 8 days.

Group 3: **CCl₄ group**, mice received CCl₄ dissolved in corn oil orally for 2 days then received corn oil for 8 days.

Group 4: **CCl₄ + WGO group**, mice received CCl₄ dissolved in corn oil orally for 2 days then received WGO for 8 days

Animal handling

At the end of the 10th day, all the mice were euthanized after anesthesia; blood samples were collected and centrifuged at 2000 g for 20 min. Serum was stored at -20°C until used for biochemical assays. The small intestine was removed and immediately divided into two parts: the first part was stored at -80°C for biochemical studies and DNA fragmentation while the second part was suspended in 10% formal saline for fixation preparatory to histological processing.

Small intestine homogenate preparation

Small intestines were homogenized (10 % w/v) in ice-cold 0.1M Tris-HCl buffer (pH 7.4). The homogenate was centrifuged at 2000 g for 15 min at 4°C and the resultant supernatant was used for the biochemical analyses.

Biochemical analysis of serum lipid profile

The appropriate kits (Bio-Diagnostic, Dokki, Giza, Egypt) were used for the determination of total lipids (Zöllner *et al.*, 1962), triglycerides (Stein *et al.*, 1960), serum total cholesterol and high-density lipoprotein -cholesterol (Tietz *et al.*, 1995) according to the manufacturer's instructions. Then, the concentration of LDL-cholesterol was calculated according to Friedewald's formula (Friedewald *et al.*, 1972). LDL-cholesterol

(mg/dL). = Serum total cholesterol – (Triglycerides/5) – HDL cholesterol. Very low-density lipoprotein (VLDL) was calculated using the following formula: $VLDL = \text{Triglycerides}/5$.

Lactate dehydrogenase (LDH)

LDH activity as a marker of tissue injury was measured in the serum spectrophotometrically using commercially available kits (Salucea, Haansberg, Netherlands). LDH activity and expressed by U/L ultraviolet kit (Salucea, Haansberg, Netherlands) (Van der Heiden *et al.*, 1994)

Oxidative stress assessment

Malondialdehyde (MDA) assay

The tissue content of MDA, an index of oxidative stress-mediated tissue lipid peroxidation, was determined by the thiobarbituric acid (TBA) method, as described previously (Ohkawa *et al.*, 1979). The small intestine was homogenized with ice-cold 0.1M Tris-HCl buffer (pH 7.4) and the values of MDA in tissue homogenate were expressed as nmol/g tissue.

Reduced glutathione (GSH) and Glutathione-S-transferase (GST) of the small intestinal tissue

GSH and GST play a major role in protecting cells against damage from reactive oxygen species (ROS). Tissue GSH (mg/g tissue) and GST (nmol/min/g tissue) were assessed in the homogenate according to the manufacturer's instructions (Habig *et al.*, 1974; Beutler *et al.*, 1963) respectively.

Superoxide dismutase (SOD) assay

The method of measuring the SOD activity depends on the inhibition of nitro blue tetrazolium (NBT) reduction by xanthine-xanthine oxidase after addition of superoxide dismutase (Nishikimi *et al.*, 1972). SOD activity was expressed as unit/g of the intestinal tissue.

Catalase (CAT) assay

The catalase activity in the intestinal tissue was determined by the method described by Aebi (1984), in which the supernatant was incubated with H₂O₂ for one minute, and then the reaction was stopped with catalase inhibitor. The color intensity of the complex formed by chromophore and H₂O₂ was inversely proportional to the amount of catalase in the sample and measured at 510 nm. CAT activity was expressed as unit/g of the intestinal tissue (Aebi 1984)

Nitric Oxide (NO[•]) assay

Nitric oxide (NO[•]) is a free radical, an uncharged molecule with an unpaired electron. The reaction of NO[•] with oxygen or other free radicals generates reactive nitrogen species (RNS), which cause multiple biological effects. Nitrite was estimated colorimetrically with the Griess reagent (Montgomery *et*

al., 1961). The content of nitric oxide in the tissue was measured at 540 nm and expressed as umol/L.

Histopathological preparation

The small intestines were fixed in 10% neutral buffered formalin solution and embedded in paraffin wax blocks. Sections of 5 μm thickness were stained with hematoxylin and eosin (H&E) then examined under light microscope for determination of pathological changes (Meyer 1903)

DNA fragmentation assay

Apoptosis is characterized by DNA fragments that are produced as a result of the endonucleolytic attack. According to Sambrook's protocol (Sambrook *et al.*, 1989), the intestinal tissues were homogenized in lysis buffer (10 mM Tris-HCl, pH 7.4, 10 mM EDTA and 0.2% triton-100x) for 20 min on ice before centrifugation, and digested with proteinase-K at 50°C (50 mg/ml) for 2 h. After digestion, DNA was extracted three times with phenol-chloroform and precipitated finally with absolute alcohol and 2.5 M sodium acetate (pH 4.0), then centrifuged at 5000 g for 30 min. DNA was washed three times with 70% ethanol dissolved in TE buffer (pH 8.0) and digested for 15 min at 37°C with DNase-free RNase-A (1 μg/ml). DNA was re-extracted in chloroform and reprecipitated with absolute ethanol and 2.5 M sodium acetate (pH 4.0).

A specific quantity (15 mg/lane) was loaded onto an agarose gel. DNA laddering was determined by a constant voltage (60 V) on a 1.2% agarose gel containing 0.4 mg/ml ethidium bromide. The gel was illuminated and photographed under UV light to record DNA fragmentation.

Statistical analysis

Data are expressed as mean ± SE of six mice in each group. Treated groups were compared with control and CCl₄ group by using Unpaired Student's t-test **P* < 0.05, ***P* < 0.01 vs control group, #*P* < 0.05; ##*P* < 0.01 vs CCl₄. SPSS, for Windows (USA, version 19.0) was used for the statistical analysis.

RESULTS

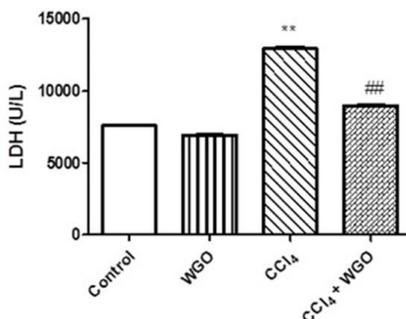
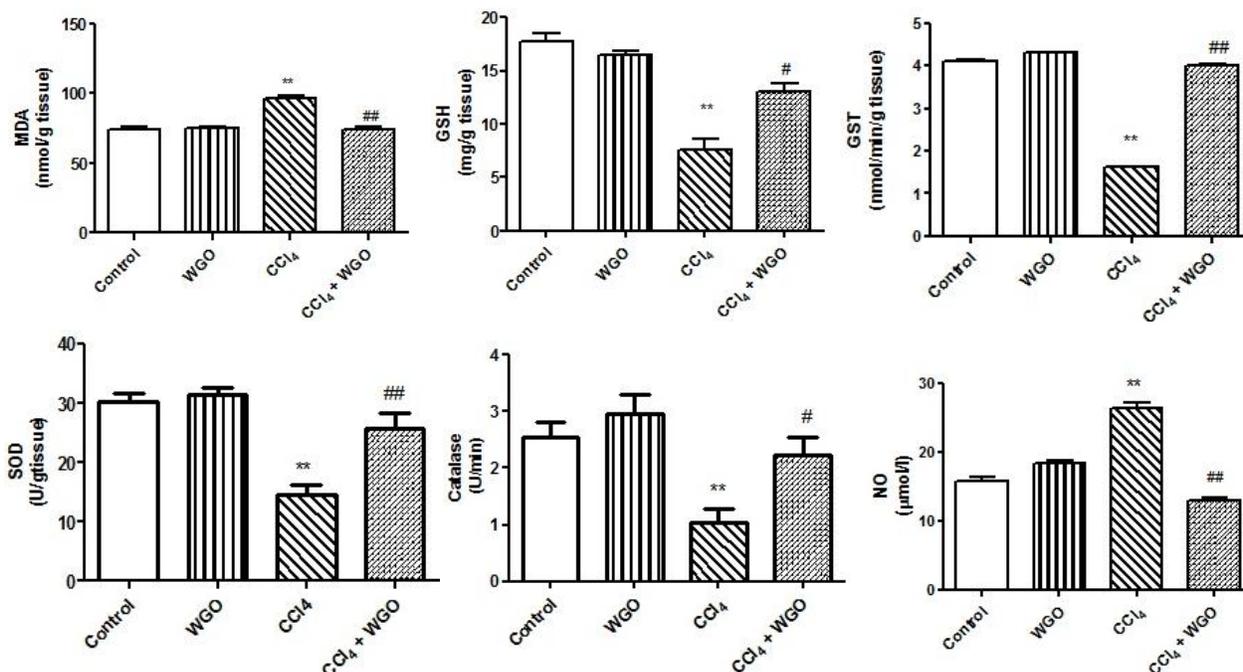
Serum lipid profile

Data recorded in Table 1 reports a significant increase (*P* < 0.01) in the serum levels of total lipids, triglycerides, total cholesterol, LDL-cholesterol and VDL-cholesterol in CCl₄ group as compared to the corresponding control group. However, CCl₄ treatment significantly decreased (*P* < 0.01) serum HDL-cholesterol in comparison to control group. On the other hand, oral administration of WGO for CCl₄ treatment group resulted in a significant decrease (*P* < 0.01) of total lipids, triglycerides, total cholesterol, LDL-cholesterol and VDL-cholesterol levels and significant increase of HDL-cholesterol as compared to the corresponding CCl₄ treatment group.

Table 1: Effect of WGO on serum lipid profile in mice treated with CCl₄.

Groups	Total Lipids (mg/dL)	Triglycerides (mg/dL)	Total cholesterol (mg/dL)	HDL-cholesterol (mg/dL)	LDL-cholesterol (mg/dL)	VLDL-cholesterol (mg/dL)
Control	591.90 ± 56.26	113.91 ± 1.63	120.52 ± 2.99	69.26 ± 0.72	32.64 ± 2.68	21.16 ± 0.53
WGO	425.43 ± 40.54	111.7 ± 2.36	111.7 ± 3.33	72.86 ± 2.15	25.66 ± 1.01	23.14 ± 0.25
CCl ₄	1108 ± 56.26**	153.70 ± 2.35**	166.20 ± 9.42**	33.90 ± 1.26**	94.23 ± 3.332**	30.73 ± 0.44**
CCl ₄ +WGO	468.5 ± 39.27##	115.63 ± 5.05##	128.80 ± 3.09##	61.33 ± 1.14##	44.35 ± 2.810##	23.13 ± 1.01##

Data are expressed as mean ± SE; (n = 6). Significant difference is indicated by * $P < 0.05$ & ** $P < 0.01$ vs control group and # $P < 0.05$ & ## $P < 0.01$ vs CCl₄.

**Fig. 1:** Effect of WGO on CCl₄-induced elevation in serum Lactate Dehydrogenase (LDH) levels as an index of general tissue damage.**Fig. 2:** Effect of WGO on CCl₄-induced changes in antioxidant markers of the small intestine.

MDA = Malondialdehyde; GSH = Glutathione Reduced; GST = Glutathione-S-transferase; SOD = superoxide dismutase; CAT = Catalase; NO = Nitric Oxide. Data are expressed as mean ± SE; (n = 6). Significant difference is indicated by * $P < 0.05$ & ** $P < 0.01$ vs control group and # $P < 0.05$ & ## $P < 0.01$ vs CCl₄.

Lactate dehydrogenase (LDH)

To evaluate the cellular injury, LDH was measured spectrophotometrically in the serum. Serum LDH was found to be elevated significantly ($P < 0.01$) after CCl₄ treatment as compared to the control group, whereas WGO administration resulted in a significant attenuation of LDH level ($P < 0.01$) (Figure 1).

Intestinal oxidative stress

The current study revealed that MDA and NO levels were increased significantly ($P < 0.01$) in CCl₄-treated mice as

compared to the control group. While, a significant decrease ($P < 0.05$) in the levels of GSH, GST, SOD and CAT were observed in the CCl₄-treated group. In contrast, the levels of MDA and NO were decreased significantly in CCl₄ + WGO group when compared to the CCl₄-treated group.

On the other hand, a significant increase in concentrations of GSH and CAT ($P < 0.05$), and GST, SOD ($P < 0.01$) were observed subsequent to the oral administration of WGO after CCl₄ treatment (Figure 2).

Histopathological examination

Sections of the small intestine for control and WGO groups showed the normal architecture of healthy tissues including the intact mucosa, cylindrical glandular epithelium and the finger-like villi structure (Figure 3 A, B).

Acute toxicity of CCl₄ has reflected the complications in the architecture of small intestine including the loose structure of the mucous membrane (L), shortened and fractured villa (S), and infiltration of the inflammatory cells into the lamina propria (F).

In addition, intestine showed cystic dilatation of some intestinal glands (C) (Figure 3 C).

DNA fragmentation

Figure (4) showed that CCl₄ produced marked intestinal DNA fragmentation below 50bp indicating the high apoptotic damaging effect of CCl₄ compared to control and WGO groups. On the other hand, much less smear of fragmented DNA was detected in CCl₄ + WGO group when compared with intact DNA in the control group.

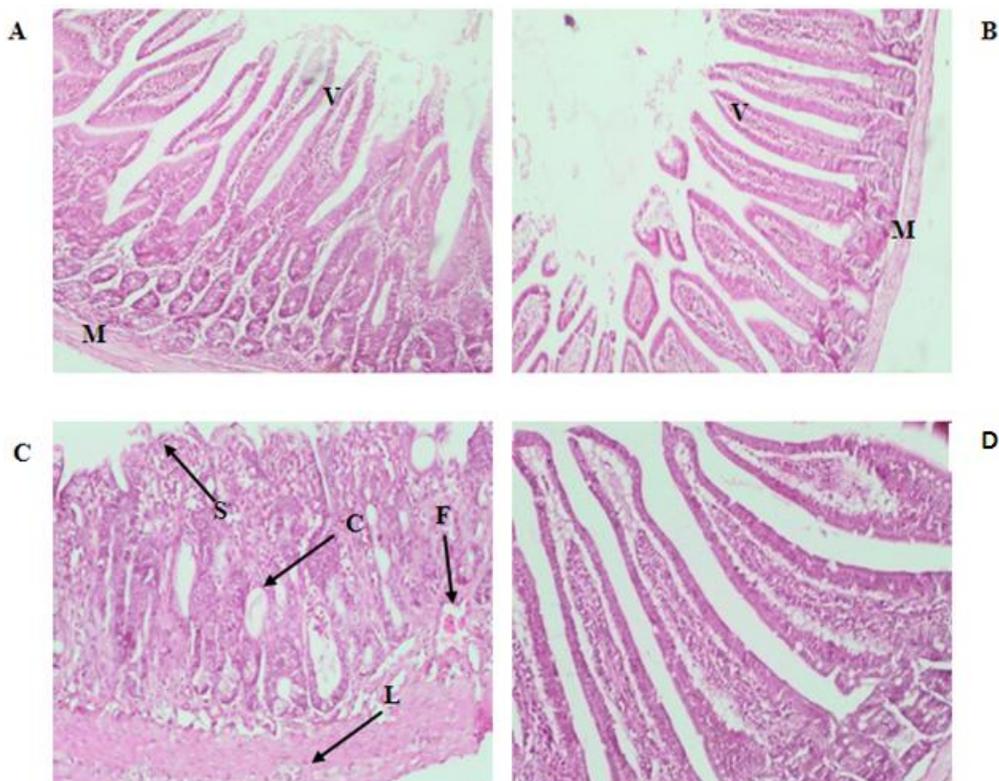


Fig. 3: Photomicrographs of intestinal tissues of mice stained with H&E, (X 200). (A and B) control and WGO groups showing the normal architecture of intestinal tissues including the intact mucosa (M), cylindrical glandular epithelium and the finger-like villi structure (V). (C): CCl₄-intoxicated mice, showed the loose structure of the mucous membrane (L), shortened and fractured villa (S), and infiltration of the inflammatory cells into the lamina propria (F). In addition, intestine showed cystic dilatation of some intestinal glands (C). (D): CCl₄ + WGO groups showed healthy morphological and the structural shape of the small intestine.

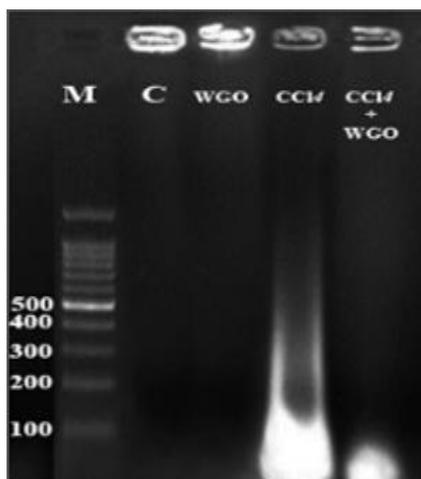


Fig. 4: DNA extracted from intestinal tissues of mice assessed by DNA fragmentation assay. Lane M: DNA ladder (low molecular weight ranges from 100-1000), C: DNA of control group, WGO: WGO-treated mice, CCl₄: CCl₄-intoxicated mice, CCl₄ + WGO: CCl₄ treated group followed by WGO administration.

CCl_4 was used for many years in experimental rodent models to induce hepatotoxicity at different levels including liver injury, fibrosis, cirrhosis, necrosis and hepatocellular carcinoma (HCC). The model of acute CCl_4 - induced necrosis in the liver has been investigated after oral administration of CCl_4 (LeSage *et al.*, 1999). CCl_4 is chemically lipophilic and diffuses simply across the cell membranes then gets distributed into the tissues (Ritesh *et al.*, 2015). In CCl_4 intoxicated mice, damage to the endoplasmic reticulum (ER) and accumulation of lipids occur in the hepatic cells (Jeon *et al.*, 2003). Boll *et al.* (2001) reported that an increase in the esterification of free fatty acids of triglycerides (TG) and phospholipids occurred due to administration of CCl_4 which in turn caused a disturbance in normal lipid homeostasis and consequently, lead to increase the levels of cholesterol synthesis.

In the present study, serum levels of total lipids, triglycerides, total cholesterol, LDL, and VLDL were significantly increased, while HDL level significantly decreased with the CCl_4 administration in comparison to control group. These findings were in agreement with Kumar *et al.* (2007), they reported a hypercholesterolemia in CCl_4 intoxicated models, that was associated with failure of the liver. This condition was due to damage of the hepatic parenchymal cells that led to disturbance of total lipid metabolism in the liver (Raju *et al.*, 2003). These effects were significantly reversed by oral administration of WGO to nearly the normal level in comparison to CCl_4 treated group. WGO is considered as a valuable source of monounsaturated and polyunsaturated fatty acids, including linoleic acid (omega-3 and omega-6) (Said *et al.*, 2008). The monounsaturated fatty acid can reduce serum TG and phospholipid levels (Jenkins *et al.*, 1999). In addition, WGO has a number of components with nutritional and health benefits, like high content of vitamin E and phytosterol. Those components might be the reason of decreasing the levels of both triglyceride and phospholipids. Moreover, WGO elevated the level of HDL-cholesterol that picks up and transports cholesterol in the blood back to the liver. That in turn leads to its elimination from the body and consequently reduces the harmful effects of LDL-cholesterol and decreases the incidence of atherosclerosis and cardiovascular diseases (Jonjala *et al.*, 2005)

Multiple organ injuries are frequently assessed by the release of lactate dehydrogenase (LDH) activity in plasma. This study showed that LDH is elevated by 1.6 folds in mice subjected to CCl_4 when compared to the control group. This finding may suggest that CCl_4 not only damages the liver but also affects other organs. Administration of WGO might be responsible for the decrease of LDH level leakage, reduce the cell death and protect against organ damage induced by CCl_4 (Zhu *et al.*, 2013). Previous study revealed that oxidative stress in the intestine of the cirrhotic liver might developed either by the generation of ROS in the mucosa or by possible inducers generated in the liver and transported into the intestinal lumen through the bile (Anup *et al.*, 1999). The present study showed that even acute dose of CCl_4 could generate marked (ROS)-induced intestinal damage through Xanthine Oxidase which is abundantly present in the intestinal mucosa (Anup *et al.*, 1999).

CCl_4 rapidly reaches the liver where it metabolized by cytochrome P_{450} . It generates the highly reactive free radical CCl_3 that initiates progression of lipid peroxidation of endoplasmic reticulum cell membrane that consequently, causes the formation of malondialdehyde (MDA) that in turn leads to damage in different organs (Ganie *et al.*, 2011). The increase in MDA level in the small intestine leads to failure of the antioxidant mechanisms to prevent the generation of excessive free radicals through reduction of glutathione and superoxide dismutase in liver injury after administration of CCl_4 (Ramachandran *et al.*, 2002).

GSH is a tripeptide thiol that is abundant in the tissue and plays a vital role in protecting cells against peroxides and free radicals generated from toxic substances including CCl_4 (Endo *et al.*, 2013).

GST plays a significant role in cell defense against ROS-mediated injury by detoxification of lipid hydro- peroxides that have been developed due to oxidative damage (Yang *et al.*, 2001). The antioxidant enzymes such as CAT, SOD and non-enzymatic electron receptors such as GSH are affected and used as indexes to evaluate the level of oxidative stress. Ramachandran *et al.* (2002) mentioned that the reason of intestinal oxidative stress in liver treated with CCl_4 are the increase in mucosal xanthine oxidase activity in conjunction with the decrease in the activity of antioxidant enzymes SOD and CAT. This was found in harmony with (Yang *et al.*, 2001) who informed that injection with CCl_4 induced significant reduction in GST that can initiate lipid peroxidation and DNA damage. The current study showed a significant increase in intestinal NO level after oral administration of CCl_4 that reflects the imbalance between pro-oxidant and antioxidant activity which led to tissue injury (Hamouda *et al.*, 2016).

In this study, acute dose of CCl_4 caused a significant reduction in GSH, GST, SOD and CAT levels in the intestinal mucosa, along with a significant elevation in MDA and NO levels. The administration of WGO suppressed the generation of free radicals and increased the levels of intestinal endogenous antioxidants. The ameliorative effect of WGO is due to its high content of vitamin E that has the potency to inactivate the reactive free radicals, thus avoiding the propagation of the radical chain reaction (Ismael *et al.*, 2014). WGO contains a high percentage of unsaturated (81%) and numerous saturated fatty acids 64% that has anti-inflammatory properties and can decrease oxygen free radicals and nicotinamide adenine dinucleotide phosphate (NADPH) oxidase activity (Abd-El-Hameed *et al.*, 2013). In addition, the phenolic compounds in WGO exhibit an antioxidant influence that could reduce the pro-oxidative state and give powerful antioxidant protection to different organs in the body (Durak *et al.*, 2010).

CCl_4 induced damage by over production of lipid peroxidation and generation of reactive oxygen species that attack and damage DNA leading to DNA hydroxylation. Several studies proved that the administration of CCl_4 caused apoptosis and DNA damage in the liver (Wiseman *et al.*, 1996; Aksit *et al.*, 2014). Our study demonstrated that CCl_4 administration caused marked

oxidative DNA damage in the small intestine through overproduction of ROS. WGO administration caused significant reduction of DNA fragmentation as a result of the ameliorative effect of WGO against ROS induced by CCl₄ treatment.

This study showed morphological alterations in the architecture of intestinal tissue which reflect the pathophysiologic complications of the small intestine in response to acute administration of CCl₄. Our findings showed a change of the intestinal epithelial barrier as a shortening of villi, cystic dilatation of glandular cells, infiltration of immune cell. The administration of WGO restored the integrity of the epithelial barrier and the healthy architecture of the small intestine through the reduction of the ROS and lipid peroxidation.

CONCLUSION

The results of the present study demonstrate that CCl₄ caused liver hypercholesterolemia and hyperlipidemia as well as an increase in the atherosclerosis index. CCl₄ caused damage to other organs such as small intestine through increasing the ROS, NO, and oxidative stress. WGO administration has the potency to protect not only the liver but also the small intestine in acute CCl₄-induced tissue damage. The valuable effect is chiefly attributed to its mechanism of reducing the lipid profile and suppressing the oxidative stress that caused DNA damage.

Financial support and sponsorship: Nil.

Conflict of Interests: There are no conflicts of interest.

REFERENCES

- Abd-El-Hameed AMA, Soliman HA, and El-Reheem ESA. Protective Role of Wheat Germ Oil in Clozapine-Induced Oxidative Stress and Biochemical Alterations in Liver of male albino rats, *Journal of American Science*, 2013; 9.
- Aebi H. Catalase *in vitro*, *Methods in enzymology*, 1984; 105: 121-26.
- Aksit H, and Bildik A. Determination of DNA damage in experimental liver intoxication and role of N-acetyl cysteine, *Cell biochemistry and biophysics*, 2014; 70: 1119-25.
- Anup R, Aparna V, Pulimood A, Balasubramanian KA, Sussana P, and Watkins P. Surgical stress and the small intestine: role of oxygen free radicals, *Surgery*, 1999; 125: 560-69.
- Anwar M, and Mohamed N. Amelioration of liver and kidney functions disorders induced by sodium nitrate in rats using wheat germ oil, *Journal of Radiation Research and Applied Sciences*, 2015; 8: 77-83.
- Bajaj JS, Hylemon PB, and Younossi Z. The intestinal microbiota and liver disease, *The American Journal of Gastroenterology Supplements*, 2012; 1: 9-14.
- Beutler E, Duron O, and Kelly BM. Improved method for the determination of blood glutathione, *The Journal of laboratory and clinical medicine*, 1963; 61: 882-88.
- Boll M, Lutz W, Becker E, and Stampfl A. Mechanism of carbon tetrachloride-induced hepatotoxicity. Hepatocellular damage by reactive carbon tetrachloride metabolites, *Zeitschrift für Naturforschung C*, 2001; 56: 649-59.
- Dashti H, Jéppsson B, Hägerstrand I, Hultberg B, Srinivas U, Abdulla M, and Bengmark S. Thioacetamide-and carbon tetrachloride-induced liver cirrhosis, *European surgical research*, 1989; 21: 83-91.
- Durak D, Kalender S, Uzun F, Demir F, and Kalender Y. Mercury chloride-induced oxidative stress and the protective effect of vitamins C and E in human erythrocytes *in vitro*, *Afr J Biotechnol*, 2010; 9: 488-95.
- Endo H, Niioka M, Kobayashi N, Tanaka M, and Watanabe T. Butyrate-producing probiotics reduce nonalcoholic fatty liver disease progression in rats: new insight into the probiotics for the gut-liver axis, *PLoS One*, 2013; 8: 6388.
- Essawy AE, Abdel-Moneim AM, Khayyat LI, and Elzergy AA. *Nigella sativa* seeds protect against hepatotoxicity and dyslipidemia induced by carbon tetrachloride in mice, *Journal of Applied Pharmaceutical Science*, 2012; 2: 21.
- Friedewald WT, Levy RI, and Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge, *Clinical chemistry*, 1972; 18: 499-502.
- Ganie SA, Haq E, Hamid A, Qurishi Y, Mahmood Z, Zargar BA, Masood A, and Zargar MA. Carbon tetrachloride induced kidney and lung tissue damages and antioxidant activities of the aqueous rhizome extract of *Podophyllum hexandrum*, *BMC complementary and alternative medicine*, 2011; 11: 17.
- Gelmez N, Kıncaç NS, and Yener ME. Optimization of supercritical carbon dioxide extraction of antioxidants from roasted wheat germ based on yield, total phenolic and tocopherol contents, and antioxidant activities of the extracts, *The Journal of Supercritical Fluids*, 2009; 48: 217-24.
- Go J, Kim JE, Koh EK, Song SH, Sung JE, Lee HA, Lee YH, Lim Y, Hong JT, and Hwang DY. Protective Effect of galloytannin-enriched Extract isolated from *Galla rhois* against CCl₄-induced hepatotoxicity in ICR mice, *Nutrients*, 2016; 8: 107.
- Gutiérrez R, Alvarado JL, Presno M, Pérez-Veyna O, Serrano CJ, and Yahuaca P. Oxidative stress modulation by *Rosmarinus officinalis* in CCl₄-induced liver cirrhosis, *Phytotherapy Research*, 2010; 24: 595-601.
- Habig WH, Pabst MJ, and Jakoby WB. Glutathione S-transferases the first enzymatic step in mercapturic acid formation, *Journal of biological Chemistry*, 1974; 249: 7130-39.
- Hamouda AF, Sameeh MY, and Shrourou RM. Effect of Avocado (*Persea Americana*), Cabbage (*Brassica Oleracea*) and Ginger (*Zingiber Officinale*) on Rat Liver and Thyroid Injuries Induced by CCl₄ (Carbon Tetrachloride), *Journal of Pharmacy and Pharmacology*, 2016; 4: 108-18.
- Ismael OE, Hashish EA, and Ali HA. Lipid Profile and Lipogenesis Following Corn Oil, Truffle Oil or Wheat Germ Oil Administration in Albino Rat, *Global Veterinaria*, 2014; 12: 461-69.
- Jenkins DJ, Kendall CW, Vuksan V, Augustin LS, Mehling C, Parker T, Vidgen E, Lee B, Faulkner D, and Seyler H. Effect of wheat bran on serum lipids: influence of particle size and wheat protein, *Journal of the American College of Nutrition*, 1999; 18: 159-65.
- Jeon TI, Hwang SG, Park NG, Jung YR, Im Shin S, Choi SD, and Park DK. Antioxidative effect of chitosan on chronic carbon tetrachloride induced hepatic injury in rats, *Toxicology*, 2003; 187: 67-73.
- Jonnala R, Dunford N, and Irmak S. 2005. "Policosanol, tocopherol and phytosterol composition of wheat extracts as affected by extraction methods. Institute of Food Technologists Annual Meeting, New Orleans." In *IFT Annual Meeting*, July, 15-20.
- Karabacak M, Kanbur M, Eraslan G, and Sarıca ZS. The antioxidant effect of wheat germ oil on subchronic coumaphos exposure in mice, *Ecotoxicology and environmental safety*, 2011; 74: 2119-25.
- Kumar SS, Kumar BR, and Mohan GK. Antihepatotoxic activity of *Trichosanthes cucumerina* on carbon tetrachloride induced liver damage in rats, *Pharmacologyonline*, 2007; 3: 461-69.
- Leenhardt F, Fardet A, Lyan B, Gueux E, Rock E, Mazur A, Chanliaud E, Demigné C, and Révész C. Wheat germ supplementation of a low vitamin E diet in rats affords effective antioxidant protection in tissues, *Journal of the American College of Nutrition*, 2008; 27: 222-28.
- LeSage GD, Benedetti A, Glaser S, Marucci L, Tretjak Z, Caligiuri A, Rodgers R, Phinzy JL, Baiocchi L, and Francis H. Acute carbon tetrachloride feeding selectively damages large, but not small, cholangiocytes from normal rat liver, *Hepatology*, 1999; 29: 307-19.

Liu X, Zheng X, Song Z, Liu X, kumar Koppurapu N, Wang X, and Zheng Y. Preparation of enzymatic pretreated corn gluten meal hydrolysate and *in vivo* evaluation of its antioxidant activity, *Journal of Functional Foods*, 2015 18: 1147-57.

Llovet J, Bartoli R, Planas Re, Cabre E, Jimenez M, Urban A, Ojanguren I, Arnal J, and Gassull M. Bacterial translocation in cirrhotic rats. Its role in the development of spontaneous bacterial peritonitis, *Gut*, 1994; 35: 1648-52.

Majzoobi M, Ghiasi F, and Farahnaky A. Physicochemical assessment of fresh chilled dairy dessert supplemented with wheat germ, *International Journal of Food Science & Technology*, 2016; 51: 78-86.

Manevska B. [Enzymatic changes in the mucous membrane of the small intestine in tetrachlormethane-induced experimental liver cirrhosis], *Eksperimentalna meditsina i morfologija*, 1975; 15: 107-11.

Meyer A. An attempt at analysis of the neurotic constitution, *The American Journal of Psychology*, 1903; 14: 90-103.

Montgomery H, and Dymock J. Colorimetric determination of nitric oxide, *Analyst*, 1961; 86: 414-17.

Nishikimi M, Rao NA, and Yagi K. The occurrence of superoxide anion in the reaction of reduced phenazine methosulfate and molecular oxygen, *Biochemical and biophysical research communications*, 1972; 46: 849-54.

Ohkawa H, Ohishi N, and Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction, *Analytical biochemistry*, 1979; 95: 351-58.

Raju K, Anbuganapathi G, Gokulakrishnan V, Raj Kapoor B, Jayakar B, and Manian S. Effect of dried fruits of *Solanum nigrum* LINN against CCl₄-induced hepatic damage in rats, *Biological and Pharmaceutical Bulletin*, 2003; 26: 1618-19.

Ramachandran A, Prabhu R, Thomas S, Reddy JB, Pulimood A, and Balasubramanian KA. Intestinal mucosal alterations in experimental cirrhosis in the rat: role of oxygen free radicals, *Hepatology*, 2002; 35: 622-29.

Risal P, Hwang PH, Yun BS, Yi H-K, Cho BH, Jang KY, and Jeong YJ. Hispidin analogue davallialactone attenuates carbon tetrachloride-induced hepatotoxicity in mice, *Journal of natural products*, 2012; 75: 1683-89.

Ritesh K, Suganya A, Dileepkumar H, Rajashekar Y, and Shivanandappa T. A single acute hepatotoxic dose of CCl₄ causes oxidative stress in the rat brain, *Toxicology Reports*, 2015; 2: 891-95.

Said UZ, Saada HN, Mahdy E-SM, Shousha WG, and Shedid SM. Wheat germ oil attenuates gamma radiation-induced skeletal muscles damage in rats. *Egyptian Journal of Biochemistry & Molecular Biology*, 2008; 26: 101-18.

Sambrook J, Fritsch EF, and Maniatis T. 1989. *Molecular Cloning: A Laboratory Manual*, 2nd ed., (Cold spring harbor laboratory press, New York).

Srivastava A, and Shivanandappa T. Hepatoprotective effect of the root extract of *Decalepis hamiltonii* against carbon tetrachloride-induced oxidative stress in rats, *Food Chemistry*, 2010; 118: 411-17.

Stein Y, and Shapiro B. Uptake and metabolism of triglycerides by the rat liver, *Journal of lipid research*, 1960; 1: 326-31.

Tietz PS, Holman RT, Miller LJ, and LaRusso NF. Isolation and characterization of rat cholangiocyte vesicles enriched in apical or basolateral plasma membrane domains, *Biochemistry*, 1995; 34: 15436-43.

Van der Heiden C, Bais R, Gerhardt W, Lorentz K, and Rosalki S. Approved recommendation on IFCC methods for the measurement of catalytic concentration of enzymes. Part 8. IFCC method for lactate dehydrogenase, *Eur J Clin Chem Clin Biochem*, 1994; 32: 639-55.

Watkins P. Role of xanthine oxidase in small bowel mucosal dysfunction after surgical stress (*Br J Surg* 2000; 87: 1094-1101), *British Journal of Surgery*, 2001; 88: 152-53.

Weber LW, Boll M, and Stampfl A. Hepatotoxicity and mechanism of action of haloalkanes: carbon tetrachloride as a toxicological model, *Critical reviews in toxicology*, 2003; 33: 105-36.

Wiseman H, and Halliwell B. Damage to DNA by reactive oxygen and nitrogen species: role in inflammatory disease and progression to cancer, *Biochemical Journal*, 1996; 313: 17.

Yang Y, Cheng J-Z, Singhal SS, Saimi M, Pandya U, Awasthi S, and Awasthi YC. Role of glutathione S-transferases in protection against lipid peroxidation overexpression of HGSTA2-2 in K562 cells protects against hydrogen peroxide-induced apoptosis and inhibits JNK and caspase 3 activation, *Journal of biological Chemistry*, 2001; 276: 19220-30.

Zhu K-X, Guo X, Guo X-N, Peng W, and Zhou H-M. Protective effects of wheat germ protein isolate hydrolysates (WGPIH) against hydrogen peroxide-induced oxidative stress in PC12 cells, *Food research international*, 2013; 53: 297-303.

Zöllner N, and Kirsch K. Colorimetric method for determination of total lipids, *Fur Gesampte Exp Med*, 1962; 135: 545.

How to cite this article:

Hanan Saleh. Modulatory Effect of Wheat Germ Oil on Intestinal Oxidative Stress and DNA damage Induced by Carbon tetrachloride in Mice. *J App Pharm Sci*, 2016; 6 (12): 067-074.