

Evaluation of Sodium Carboxymethyl Starch obtained from *Ipomoea Batatas*

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ABSTRACT

The study sought to evaluate the functional properties of sodium carboxymethyl starch obtained from *Ipomoea batatas*, in order to facilitate their exploitation as substitute excipients for the local pharmaceutical manufacturing industry. The sodium carboxymethyl starch was produced by reacting native starch with sodium hydroxide and sodium monochloroacetate in various proportions and reaction time at constant temperature. Subsequently, the starches were obtained by wet separation techniques. Physicochemical properties and proximate analysis were carried out in order to determine their suitability for pharmaceutical use. Differences in the physicochemical properties, proximate composition, and functional properties of sodium carboxymethyl starches and native *Ipomoea batatas* starch were significant. Optimum degree of substitution and reaction efficiency of 0.52 and 1.92% were achieved when sodium hydroxide and sodium monochloroacetate in a ratio 1.5 and 2.0 Mole per anhydrous glucose unite respectively in 6 hrs was used. Generally, the carboxymethyl starches had higher bulk density, tapped density, true density, hydration and swelling capacity as compared to the native starch.

INTRODUCTION

The use of natural starch and its derivatives as biodegradable polymers continue to be an area of active research despite the advent of synthetic polymers. It does remain attractive, primarily because they are inexpensive, readily available, degradable, biocompatible and non-toxic (Nikazar *et al.*, 2005; Yaacob, *et al.*, 2011). Recently, natural starch has increasingly gained importance as a backbone for new biopolymer material and functional polymer mainly synthesized by chemical modification reactions (Abdulmajid *et al.*, 2010). Carboxymethylation process of starch is one of the most versatile functionalization procedures as it provides access to biomaterials with valuable properties. The alterations take place at the molecular level with little or no change taking place in the superficial appearance of the granule. Sweet potato (*Ipomoea batatas*) is adaptable to a broad range of agro-ecological conditions and fits into low-input agriculture. It is highly productive even under adverse farming conditions (Prakash, 1994). Sweet potato is cultivated in more than 100 countries as a

valuable source of human food, animal feed and industrial raw material. The objective of this study is to evaluate sodium carboxymethyl starch obtained from *Ipomoea batatas* as an excipient for pharmaceutical applications.

MATERIALS AND METHODS

Materials

The root tubers of *Ipomoea batatas* were obtained from Sokoto central market, Sokoto state, Nigeria. All other chemicals and reagents used were of analytical grade.

Methods

Extraction

The procedure described by Alves *et al.* (2002) was adopted with little modification. The tubers were peeled and cut into cube. These cubes were then soaked in 10 liters 0.075 % of sodium metabisulphite solution overnight. This mixture was then milled, stirred and filtered using double fold clean cheesecloth which was allowed to completely settle while the starch sediments. This was followed by decanting the water and centrifuging the suspension at 4000 rpm for 5 min using Lab centrifuge (Thermo electron co. IEC FL40R, France); after centrifugation, the starch was separated from water and non water soluble constituents.

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Finally the pure starch obtained from centrifugation was air dried in a hot air oven (Nurve FN055 Oven, Germany), grinded with a grinder (Super Master Co. Ltd. SMB-3377) and sifted through Sethi standard sieves. The flour was then packed into an airtight container and stored at room temperature for further analysis.

Starch Modification

Organic slurry method of modification was used as described by Lawal, *et al.*, (2007). The native sweet potato starch (10.0 g) was suspended in 2-propanol (200 ml). 20 ml of aqueous sodium hydroxide solution was added (1.0 M, 1.5 M, and 2.0 M). The mixture was stirred at controlled temperature (40 °C) for 10 min. 80 ml Sodium monoacetate of 2.0 M conc. was added and stirring was continued up to the designated time (2 hrs, 4 hrs and 6 hrs). The pH of mixture was adjusted to about 5.0 by addition of 50 % glacial acetic acid. The carboxymethyl starch was filtered and washed with 80 % aqueous ethanol until the pH of liquid becomes neutral (7.0). The modified starch was dried at 50 °C for 6 hrs. The dried carboxymethyl starch was passed through a 100-mesh sieve.

FT-IR Study

Both native and carboxymethyl starch sample (5 mg) were blended with solid KBr, (Merck, Germany) and about 40 mg blend was used to prepare a pellet (Hydraulic pellet press KP, Mumbai, India). The spectra were scanned from 4500-250 cm⁻¹ in a Perkin Elmer FT-IR spectrometer (Perkin Elmer, USA) under dry air at room temperature.

Determination of the Degree of Substitution of CMS

The degree of substitution (DS) was determined with flame atomic absorption spectrometry based on the sodium content of the CMS according to the method describe by Lawal, *et al* (2009). The sample (50 mg) was heated with conc. nitric acid (4 cm³) in glass vessel with hot plate. The digestion was made up to 100 cm³ with deionized water. Flame atomic absorption spectrometer was used for the analysis. The flame composition was air-acetylene while the wavelength of sodium was 589.0 nm. The degree of substitution was determined as follows:

$$DS = \frac{162\%Na}{(2300 - 80\%Na)} \dots \dots \dots (1)$$

% Na of the unmodified starch was predetermined and it was corrected in the CMS derivative.

Determination of Reaction Efficiency

Reaction efficiency was determined using the following equation as described by Lawal *et al* (2008)

$$RE = \frac{DSX100}{Dst} \dots \dots \dots (2)$$

Where: $Dst = \frac{n_{SMA}}{n_{AGU}}$, If $n_{NaOH} \geq n_{SMCA}$

$Dst = \frac{n_{NaOH}}{n_{AGU}}$, If $n_{NaOH} < n_{SMCA}$

$Nsmca$ = Number of moles of sodium

monoacetate, n_{AGU} = Number of moles of anhydroglucose

unit, n_{NaOH} = Number of moles of sodium hydroxide.

True density

The true density (D_t) of the starch was determined by the liquid displacement method using xylene as the immersion fluid as described by Ohwoauvorhua *et al* (2004) and computed according to the following equation.

$$D_t = W / [(a+w) - b] \times SG$$

Where w is the weight of the powder, SG is specific gravity of liquid, a , is Weight of bottle + liquid and b is weight of bottle + solvent + powder.

Bulk and tapped densities

A 10 g quantity of the powder sample was Placed in 50 ml clean, dry measuring cylinder and the volume V_o occupied by the sample without tapping determined. After 500 manual taps, occupied volume V_{500} was determined. The bulk and tapped densities was calculated as the ratio of the weight of weight of volume (V_o and V_{500} respectively). The Carr's index and Hausner's ratio were determined from the values of the bulk and tapped densities results obtained above.

Powder porosity

This was determined from the values of true and bulk densities when fitted into the equation according to the method of Ohwoauvorhua *et al* (2004):

$$e = 1 - B_b/D_t \times 10$$

Where B_b , is the bulk density, D_t is the true density and e is the porosity

Hydration capacity

The method of Kornblum and stoopak (1973) was used. A 1 g sample was placed in each of four 15 ml plastic centrifuge tubes and 10ml distilled water added from a 10 ml measuring cylinder and then stopped. The contents were mixed on a vortex mixer for 2 min. The mixture then allowed to stand for 10min and immediately centrifuged. The supernatant was carefully decanted and the sediment weighed. The hydration capacity was taken as the ratio of the weight of the sediment to the dry sample weight.

Swelling capacity

This was determined at the same time as the hydration determination using the method of Okhamafe *et al* (1991) and computed according to the following equation;

$$S = (V_2 - V_1)/V_1 \times 100 \%$$

Where S is the % swelling capacity, V_2 is the volume of the hydration or swollen material and V_1 is the tapped volume of the material prior to hydration.

Proximate and Elemental analysis

The proximate analysis for moisture, crude protein, crude lipid, fiber and ash content of native and retrograded starches were carried out according to the method of the AOAC (1990). The conversion factor of total nitrogen to crude protein was 6.25. Percentage total carbohydrate was determined by subtracting the

sum total of ash, crude protein, lipid and fiber from 100. The elemental analysis was carried out using an atomic absorption spectrophotometer.

RESULTS AND DISCUSSION

The addition of carboxyl group is indicated by presence of an absorption peak band at 3875 in the FT-IR spectrum of sodium CMS which is not present in native starch (Fig. 1).

Increase in the amount of NaOH favorably increases both the DS and reaction efficiency. The limitation observed as the concentration of NaOH increases in the reaction mixture can be explain based on two competing reaction during the carboxymethylation process. Firstly, the NaOH provides the alkaline environment for the reaction as well as serving as the swelling agent to facilitate diffusion and penetration of the etherifying agent to starch granular structure and secondly, further increase in NaOH concentration causes an inactivation of sodium monochloroacetate and hence consumed in the side reaction (Lawal *et al.*, 2008).

Table 1: Parameters for Starch Modifications.

Samples	NaOH(M)	SMCA(M)	Time (hrs)	Temp.(^o C)
SP1	1.0	2.0	2.0	40
SP2	1.5	2.0	2.0	40
SP3	2.0	2.0	2.0	40
SP4	1.0	2.0	4.0	40
SP5	1.5	2.0	4.0	40
SP6	2.0	2.0	4.0	40
SP7	1.0	2.0	6.0	40
SP8	1.5	2.0	6.0	40
SP9	2.0	2.0	6.0	40

Both RE and DS (Table 2) increased with increase in reaction time within the time frame studied. The increase in the DS and RE with increase time is as a result of enhanced period of contact of etherifying reagent with the starch molecules. However no remarkable further increases were observed in both DS and RE after 4 hrs of reaction. The Swelling power of the starches increased with starch modification. This may be due to water penetrating into the starch granules as a result of the hydrophilic

nature of carboxymethyl groups. Similarly, carboxymethyl starch had high hydration capacity as compared to native starch. This may be due to addition of negatively charged carboxymethyl group to starch.

Table 2: Material Properties of Native and Sodium Carboxymethyl Starches.

Samples	Degree of Substitution	Reaction Efficiency (%)	Swelling Capacity(%)	Hydration Capacity (%)
SP1	0.07	0.41	25.00	1.80
SP2	0.41	1.52	42.86	1.87
SP3	0.14	0.40	42.86	1.85
SP4	0.14	0.78	42.86	1.84
SP5	0.11	0.41	28.57	1.81
SP6	0.10	0.26	28.57	1.80
SP7	0.16	0.88	42.86	1.86
SP8	0.52	1.92	53.85	1.96
SP9	0.14	0.38	33.33	1.81
Native	--	--	12.50	1.70

The physical properties showed little or no significant differences in bulk and flow properties between the native starch and carboxymethyl derivative but higher powder porosity of the carboxymethyl derivatives as compared to native starch (Table 3). The proximate composition is a major determinant of starch purity. It determines the presence of all other contaminants or impurities other than pure starch. Residual moisture content of starches from the native and carboxymethyl derivative were less than 15 %, which is the upper limit recommended by the British Pharmacopoeia, 2010 (Table 4). The ash content of the native starch investigated were less than 0.6 % w/w while the ash content of carboxymethyl derivatives were mostly higher (Table 4). This might be due to the impurities contributed by the reagents used during modification. The Carboxymethyl derivative had lower lipid content and higher crude protein as compared to the native starch. From the results as seen in Table 5, there was no significant difference between calcium, magnesium and phosphorus but an increase in sodium and potassium with starch modification. Presence of inorganic salts and ions of phosphorous, sodium, iodine and hydroxyl groups in starch have been reported to contribute significantly to starch granule swelling and gelatinization (Mistry and Eckhoff, 1992).

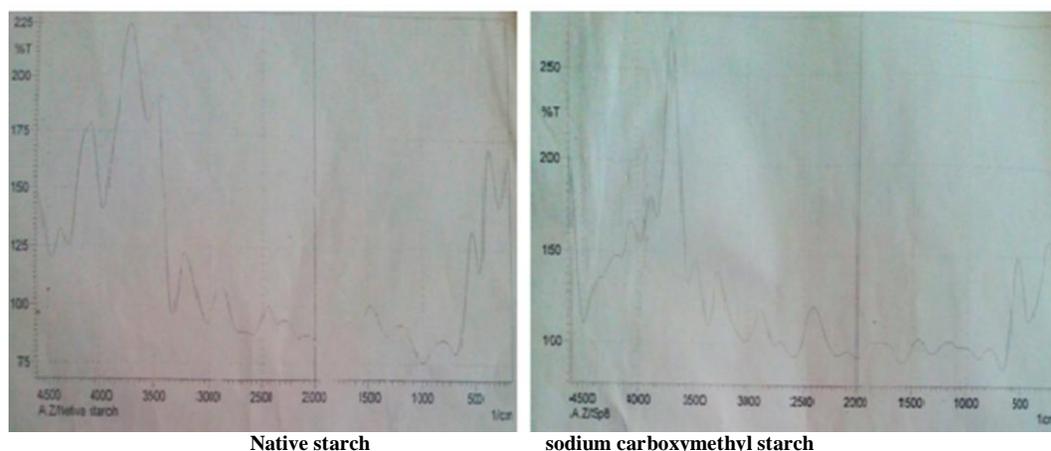


Fig. 1: FTIR Spectra of native and sodium carboxymethyl starch.

Table 3: Physicochemical Parameters of Native and Sodium Carboxymethyl Starches.

Samples	Bulk Density (g/ml)	Tapped Density (g/ml)	Carr's Index	Hausner's Ratio	True Density (g/cm ³)	Powder Porosity
SP1	0.57	0.79	0.22	1.38	3.28	8.26
SP2	0.55	0.82	0.27	1.50	3.14	8.26
SP3	0.53	0.76	0.23	1.50	2.54	7.92
SP4	0.51	0.75	0.24	1.47	2.50	7.95
SP5	0.58	0.84	0.26	1.41	2.71	7.95
SP6	0.58	0.80	0.22	1.39	3.14	8.16
SP7	0.55	0.83	0.28	1.50	2.86	8.07
SP8	0.43	0.81	0.38	1.87	1.95	7.77
SP9	0.65	0.91	0.26	1.41	2.83	7.72
Native	0.61	0.84	0.23	1.31	2.50	7.55

Table 4: Proximate Analysis of Native and sodium Carboxymethyl Starches.

Samples	Moisture Content (%)	Ash (%)	Nitrogen (%)	Crude Protein (%)	Lipid (%)	Fiber (%)	Carbohydrate (%)
SP1	5.05	0.10	0.57	3.59	0.05	0.05	96.21
SP2	5.50	0.33	0.57	3.59	0.45	0.05	95.58
SP3	4.00	0.86	0.38	2.36	0.05	0.05	96.68
SP4	4.75	0.86	0.50	3.24	0.30	0.05	95.55
SP5	4.70	0.93	0.55	3.41	0.05	0.05	95.56
SP6	5.50	0.96	0.52	3.24	0.05	0.05	95.70
SP7	4.30	0.36	0.50	3.24	0.50	0.05	95.85
SP8	4.55	0.81	0.50	3.24	0.15	0.05	95.75
SP9	4.35	0.58	0.60	3.76	0.10	0.05	95.51
Native	9.25	0.05	0.42	2.63	0.55	0.05	96.72

Table 5: Elementary Analysis of Native and Sodium Carboxymethyl Starches.

Samples	Sodium (mg/Kg)	Potassium (mg/Kg)	Calcium (mg/Kg)	Magnesium (mg/Kg)	Phosphorus (mg/Kg)
SP1	132.0	110.5	0.35	0.25	2.2
SP2	515.0	55.0	0.35	0.20	2.2
SP3	222.0	36.5	0.40	0.25	2.1
SP4	212.0	29.8	0.35	0.25	2.6
SP5	180.0	75.0	0.35	0.25	2.0
SP6	160.5	30.0	0.40	0.30	2.6
SP7	240.0	40.0	0.35	0.25	2.3
SP8	615.0	90.0	0.45	0.20	2.3
SP9	212.0	47.0	0.25	0.20	2.0
Native	31.3	45.0	0.25	0.30	2.1

CONCLUSION

Sodium carboxymethyl starch derived from *Ipomoea batatas* would be a suitable alternative to other native and modified starches commonly used in pharmaceutical preparations.

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