Journal of Applied Pharmaceutical Science Vol. 4 (02), pp. 107-111, February, 2014 Available online at http://www.japsonline.com DOI: 10.7324/JAPS.2014.40217 ISSN 2231-3354 (CC) EY-NO-SA

In vitro antioxidant potential of the essential oil and leaf extracts of *Curcuma zedoaria* Rosc.

Atiqur Rahman^{1*}, Monira Afroz¹, Rafiquel Islam¹, Kazi Didarul Islam², M. Amzad Hossain ³ and Minkyun Na^{4*}

¹Department of Applied Chemistry and Chemical Technology, Islamic University, Kushtia 7003, Bangladesh

²Biotechnology and Genetic Engineering Discipline, Khulna University, Khulna 9208, Bangladesh

³ Natural Product Chemistry Lab., Biotechnology Research Institute (IPB), Universiti Malaysia Sabah, Kota Kinabalu, Sabah, Malaysia

⁴ College of Pharmacy, Chungnam National University, 99 Daehak-ro, Yuseong-gu, Daejeon 305-764, Republic of Korea.

ARTICLE INFO

Article history: Received on: 22/03/2013 Revised on: 26/07/2013 Accepted on: 16/01/2014 Available online: 27/02/2014

Key words:

Curcuma zedoaria Rosc.; essential oil; GC-MS; antioxidant activities; total phenolics

INTRODUCTION

Phone: +82-42-821-5925, Fax: +82-42-823-6566

ABSTRACT

In this study, we examined the chemical composition of the essential oil and tested the antioxidant potential of the oil and leaf extracts of *Curcuma zedoaria* Rosc. The chemical compositions of the oil were analysed by GC-MS. Twenty-four compounds representing 92.4% of the total oil was identified. The antioxidative potential was evaluated using two separate methods, inhibition of free radical 1, 1-diphenyl-2-picrylhydrazyl (DPPH) and superoxide radicals scavenging activities assay. In the first case, the IC₅₀ value of the oil was 14.8 ± 2.2. Among the extracts, the strongest activity was exhibited by the ethyl acetate extract (IC₅₀ = 17.56 ± 1.6 µg/ml). In the superoxide radicals scavenging activities assay, ethyl acetate extract was superior to all other extracts (IC₅₀ = $23.47 \pm 1.2 \mu$ g/ml). Furthermore, the amount of total phenolic compounds was also determined as gallic acid equivalent. Thus, the natural products produced from *C. zedoaria* may be used in food and pharmaceutical industries.

Many medicinal plants contain large amounts of antioxidants such as poly phenols, which have been widely used as additives to avoid the degradation of foods. Also, antioxidants have an important role in preventing a variety of stress-related diseases because these are closely related to the active oxygen and lipid per oxidation (Noguchi and Niki, 1999). Curcuma zedoaria Rosc. is a medicinal properties-bearing Zingiberaceae from which rhizomes are commercially exploited. Natural products from this species are widely used in perfumary, in food industry as condiment and dye, and medicine as well. In addition to the well known effect of zedoary as a stomachical, it has been recently studied by its anti tumor (Kim et al., 2000), hepatoprotective (Matsuda et al., 2001), anti inflammatory (Jang et al., 2001) and analgesic (Navarro et al., 2002) effects. In the Ayrvedic system of medicine, C. zedoaria is known as zedoary while its conventional name is aadaa or ginger. C. zedoaria grows up to 1.2 m in height. The leaves of zedoary are oblong and can be up to 81 cm long and 18 cm wide. There are many species belonging with this genus but only four species namely C. aeruginosa, C. phaeocaulis, C. pallida and C. zedoaria were recorded as medicinal species.

Free radicals are responsible for aging and causing various human diseases. A study shows that antioxidant substances which scavenge free radicals play an important role in the prevention of free radical-induced diseases. By donating hydrogen radical, the primary radicals are reduced to non radical chemical compounds and are converted to oxidize antioxidant radicals (Jadhav *et al.*, 1995; Yamaguchi *et al.*, 1998). This action helps in protecting the body from degenerative disease. Widely used artificial antioxidants, such as butylated hydroxytoluene (BHT) and butylated hydroxyanisole (BHA), are very effective in their role (Chan, 1987). However, their use in food products has been falling off due to their instability, as well as due to a suspected action as promoters of carcinogenesis (Namiki, 1990; Pokorny, 1991). For this reason, there is a growing interest in the studies of natural additives as potential antioxidants.

^{*} Corresponding Author

Dr. Atiqur Rahman, E-mails: marahman12@yahoo.com

Phone: +88-071-74910-20 ext-2266, Fax: +88-071-74905, (Off.) Dr. Minkyun Na, Email : mkna@cnu.ac.kr

^{© 2014} Atiqur Rahman et al. This is an open access article distributed under the terms of the Creative Commons Attribution License -NonCommercial-ShareAlike Unported License (http://creativecommons.org/licenses/by-nc-sa/3.0/).

The plant is native to India and Indonesia. It was introduced to Europe by Arabs around the sixth century. In Bangladesh, it is found mostly in Chittagong, Dhaka, Srimangal and Dinajpur. It is also been cultivated in home yards and gardens. (Staples and Herbst, 2005). Therefore, the aim of this study was to determine the chemical composition of the essential oil from leaves of *C. zedoaria* by GC–MS and to evaluate the antioxidative properties of the essential oil and various organic extracts.

MATERIALS AND METHODS

Chemicals and Reagents

Gallic acid, kojic acid, L-ascorbic acid, L-tyrosine, Folin–Ciocalteau's phenol reagent, 1, 1- diphenyl-2-picrylhydrazyl (DPPH), butylated hydroxyanisole (BHA),tyrosinase, and xanthine oxidase (XOD) were obtained from Sigma–Aldrich (St. Louis, MO). Nitrotetrazolium blue chloride (NBT) was purchased from Fluka (Buchs, Switzerland). All other chemicals and solvents were of highest commercial grade.

Plant materials

The leaves of *Curcuma zedoaria* Rosc. were collected from Gopalgonj, Jhenaidah and Kushtia of Bangladesh in January 2010 and identified by Dr. Oliur Rahman, Bangladesh National Herbarium, Dhaka, where a voucher specimen (DACB 35212) has been deposited.

Isolation of the Essential Oil

The air-dried leaves (200 g) of *C. zedoaria* were subjected to hydro distillation for 3h using a Clevenger type apparatus. The oil was dried over anhydrous Na_2SO_4 and preserved in a sealed vial at 4°C for further analysis.

Preparation of Organic Extracts

The air-dried leaves *C. zedoaria* were first pulverized into powdered form. The dried powder (50 g) was then extracted with hexane, chloroform, ethyl acetate and methanol separately at room temperature for 7 days and the solvents were evaporated by vacuum rotary evaporator temperature at 50°C. The extraction process yielded hexane (7.3 g), chloroform (6.2 g), ethyl acetate (7.4 g) and methanol (6.5 g) extracts respectively. Solvents (analytical grade) for extraction were obtained from commercial sources (Sigma–Aldrich, St. Louis, MO, USA).

Gas Chromatography-Mass Spectrometry (GC-MS) Analysis

The GC-MS was carried out using total ion monitoring mode on a Varian 3800 gas chromatograph interfaced to a Varian Saturn ion trap 2200 GC-MS spectrometer. The temperatures of transfer line and ion source were 280°C and 275°C respectively. Ions were obtained by electron ionization mode. The VF-5 capillary column (30 m length, 0.25 mm I.D. and 0.25 μ m film thickness) was used. A 20% split injection mode was selected with a solvent delay time of 3 min with injection volume 0.2 μ l. The initial column temperature was started at 50°C for 1 min, programmed at 8°C/min to 200°C and heated until 280°C at 10°C/min. Injection port was set at 250°C. Helium was used as the carrier gas at a constant flow rate of 1.0 ml/min. Molecular ions (mass range: 40-500 m/z) were monitored for identification. The relative percentage of the oil constituents was expressed as percentage by peak area normalization. Identification of components of the essential oil was based on their retention indices, relative to a homologous series of *n*-alkane ($C_8 - C_{20}$) on the VF-5 capillary column under the same operating conditions and computer matching with the GC-MS spectra from the Wiley 6.0 MS data and literature data (Adams, 2007).

Determination of antioxidant activity DPPH-Radical Scavenging Activity

DPPH radical scavenging activity of the extracts was determined by the method described by (Archana *et al.*, 2005). Various concentrations of test extracts were added to 2.9 ml of a 0.004% (w/v) methanol solution of DPPH. After 30 min of incubation period at room temperature, the absorbance was measured against a blank at 517 nm. IC₅₀ values (concentration of sample required to scavenge 50% of free radicals) were calculated from the regression equation. Synthetic antioxidant reagents, butylated hydroxyanisole (BHA) and L-ascorbic acid were used as reference positive controls. Inhibition free radical DPPH in percent (I %) was calculated in following way:

$$I\% = (A_{blank} - A_{sample} / A_{blank}) \times 100$$

Where A_{blank} is the absorbance of the control reaction (containing all reagents except the test compound), and A_{sample} is the absorbance of the test compound. IC₅₀ values (concentration of sample required to scavenge 50% of free radicals) were calculated from the regression equation.

Superoxide anion (O_2^{\bullet}) scavenging activity

Superoxide radicals were generated in vitro by the xanthine oxidase (XOD). The scavenging activity of the sample was determined using the nitro-blue tetrazolium (NBT) reduction method. In this method, O_2 reduces the yellow dye (NBT²⁺) to produce the blue formazan, which was measured spectrophotometrically at 560 nm. Antioxidants are able to inhibit the blue NBT formation (Cos *et al.*, 1998). The results were calculated as the percentage of inhibition according to the following formula:

I (%) = 100[1-(S-SB) / (C-CB)] where S, SB, C, and CB are the absorbance of the sample, the blank sample, the control, and the blank control, respectively.

Determination of total phenolics

Total phenolic constituents of the aforementioned extracts were determined by Folin-Ciocalteu reagent in alkaline medium (Lister and Wilson, 2001) and was expressed as gallic acid equivalents (GAE). The absorbance of samples was measured at 760 nm and the results were expressed in mg/g (GAE) of dry weight of samples.

RESULTS AND DISCUSSION

Chemical composition of the essential oil

The hydrodistillation of dried leaves of *C. zedoaria* Rosc. gave yellowish essential oil (yields ~ 0.8%, w/w). The identified compounds, qualitative and quantitative analytical results by GC-MS, according to their elution order on a VF-5 capillary column are shown in Table 1.

Table. 1: Chemical	composition	of the	essential	oil	of	Curcuma	zedoaria
Rosc.							

Sl. No.	Compound	RI ^a	[%] RA	Identification
1.	Cyclohexane	719	1.2	RI, MS
2.	cis-4-Heptenal	884	0.6	RI, MS
3.	Acetic acid	895	1.5	RI, MS
4.	Cyclopentylmethanol	903	tr	RI, MS
5.	l-Octen-3-ol	961	12.4	RI, MS
6.	Benzyl alcohol	1036	2.6	RI, MS
7.	Eucalyptol	1059	22.4	RI, MS
8.	Borneol	1088	1.6	RI, MS
9.	Naphthalene	1196	tr	RI, MS
10.	1,2-Dihydro-1,1,6-trimethyl-	1224	0.4	RI, MS
	naphthalene			
11.	1-Methylnaphthalene	1289	1.5	RI, MS
12.	Bornyl acetate	1292	0.8	RI, MS
13.	Cycloheptanol	1306	1.6	RI, MS
14.	Azulene	1386	1.3	RI, MS
15.	β- Elemene	1388	9.6	RI, MS
16.	Benzofuranone	1426	1.2	RI, MS
17.	γ-Elemene	1430	0.8	RI, MS
18.	α-Caryophyllene	1447	17.2	RI, MS
19.	Ledol	1530	1.6	RI, MS
20.	(-)-Spathulenol (Spathulenol)	1550	2.2	RI, MS
21.	Caryophellene oxide	1561	8.3	RI, MS
22.	Aromadendrene oxide-(II)	1664	0.8	RI, MS
23.	Hexahydrofarnesylacetone	1816	1.0	RI, MS
24.	Pentadecanoic acid	1832	1.8	RI, MS
	Total identified		92.4%	

^aRetention index relative to *n*-alkanes on VF-5 capillary column, tr: trace amount (< 0.2%).

RI, comparison of retention index with bibliography.

Twenty-four constituents accounting for 92.4% of total oil compositions were identified. The oil contains a complex mixture consisting of mainly oxygenated monoand sesquiterpenes, and mono- and sesquiterpene hydrocarbons. The major compounds detected in the leaves oil were eucalyptol (22.4%), α- caryophyllene (17.2%), 1-octen-3-ol (12.4%), βelemene (9.6%) and caryophyllene oxide (8.3%). Besides, benzyl alcohol (2.6%), ledol (1.6%), cycloheptanol (1.6%) and pentadecanoic acid (1.8%) were also found to be the minor components of Curcuma zedoaria oil. On the other hand, the previous research shows that the oil of C. zedoaria was made up mainly of mono- and sesquiterpenoids, monoterpene hydrocarbons oxygenated monoterpenes (26%), (2.3%).sesquiterpene hydrocarbons (38%), and oxygenated sesquiterpenes (13.5%). α terpinyl acetate (8.4%), isoborneol (7%), dehydrocurdione (9%) and selina-4(15),7(11)-dien-8-one (9.4%) were the major constituents of the leaf oil (Garg et al., 2005). However, it is noteworthy that the composition of the essential oils from a particular species of plant can differ between harvesting seasons, extraction methods, and geographical sources, and that those from the different parts of the same plant can also differ widely (Burt, 2004). Several papers reported that all these compounds possess significant antioxidant activity in several model systems (Ruberto and Baratta, 2000; Cabrera and Prieto, 2010). It is also possible that the minor components might be involved in some type of synergism with the other active compounds. Also, the essential oils are, from the chemical point of view, quite complex mixtures constituted by several tens of components, and this complexity makes it often difficult to explain the aforesaid activities. If we exclude the case of some phenolic components, whose antimicrobial and antioxidant activity is well known and widely documented (Helander et al., 1998; Yanishlieva et al., 1999) and some other examples of pure compounds nothing is known about the effectiveness of most components (Aeschbach et al., 1994).

Determination of antioxidant activity Scavenging activity of DPPH radical

The DPPH is a stable free radical, which has been widely accepted as a tool for estimating free radical scavenging activities of antioxidants (Sánchez-Moreno, 2002). The effect of antioxidants on DPPH radical scavenging was thought to be due to their hydrogen donating ability. DPPH is a stable free radical and accepts an electron or hydrogen radical to become a stable diamagnetic molecule (Soares *et al.*, 1997). The reduction capability of DPPH radicals was determined by the decrease in its absorbance at 517 nm induced by antioxidants. The DPPH radical scavenging activity of the essential oil and the organic extracts are shown in Table 2.

Table. 2:	Free radical scavenging activity (DPF	'H) of the oil and extracts
of Curcun	na zedoaria R.	

Sample	IC ₅₀ (µg/ml)
Essential oil	14.8 ± 2.2
Methanol extract	25.11 ± 1.5
Hexane extract	117.79 ± 2.2
Chloroform extract	50.13 ± 1.1
Ethyl acetate extract	17.56 ± 1.6
Ascorbic acid (Control)	6.55 ± 0.9
BHA (Control)	18.27 ± 1.4
Chloroform extract Ethyl acetate extract Ascorbic acid (Control) BHA (Control)	50.13 ± 1.1 17.56 ± 1.6 6.55 ± 0.9 18.27 ± 1.4

Values are given as mean ± S.D. of triplicate experiments. BHA: butylated hydroxyanisole.

Lower IC₅₀ value indicates higher antioxidant activity. Polar extracts exhibited stronger activity than non-polar extracts. Of all samples studied, the essential oil and ethyl acetate extract had the strongest free radical-scavenging activity with an IC₅₀ value of 14.8 ± 2.2 and $17.56 \pm 1.6 \,\mu$ g/ml, respectively. The methanol (IC₅₀ = $25.11 \pm 1.5 \,\mu$ g/ml) and the chloroform extract (IC₅₀ = $50.13 \pm 1.1 \,\mu$ g/ml) showed moderate DPPH radical scavenging activity, while hexane extract showed little activity (IC50 = $117.79 \pm 2.2 \,\mu$ g/ml) as compared to positive controls L-ascorbic acid (IC₅₀ = $6.55 \pm 0.9 \,\mu$ g/ml) and BHA (IC₅₀ = $18.27 \pm 1.4 \,\mu$ g/ml). These results indicated that organic extracts of *C. zedoaria* leaves have a noticeable effect on scavenging free radical.

^bRelative area (peak area relative to the total peak area);

^cIdentification: MS, comparison of mass spectra with MS libraries.

Superoxide anion (O_2^{-}) scavenging activity

In the PMS/NADH-NBT system, superoxide anion derived from dissolved oxygen by PMS/NADH coupling reaction reduces NBT. The decrease of absorbance at 560 nm with antioxidants thus indicates the consumption of superoxide anion in the reaction mixture. The scavenging activities of the extracts on superoxide radicals are shown in Table 3.

Table. 3: Superoxide anion (O_2^{-}) scavenging activity of various leaf extracts of *Curcuma zedoaria* R.

Sample	$IC_{50} (\mu g/ml)$
Essential oil	ne
Methanol extract	34.41 ± 1.5
Chloroform extract	51.76 ± 1.7
Ethyl acetate extract	23.47±1.2
Kojic acid	8.27 ± 1.4

Values are given as mean ± S.D. of triplicate experiments. ne: not examined.

All the extracts exhibited noticeable superoxide radical scavenging activities. The highest superoxide radical scavenging activities was found in ethyl acetate extract (IC₅₀ = 23.47 ± 1.2 µg/ml). Methanol and chloroform extract exhibited IC₅₀ values 34.41 ± 1.5 and 51.76 ± 1.7 µg/ml, respectively. However, those values were significantly higher than the value of the positive control kojic acid (IC₅₀ = 8.27 ± 1.4 µg/ml). These results imply that organic extracts of *C. zedoaria* leaves are superoxide scavengers and their capacity to scavenge superoxide may contribute to their antioxidant activity.

Total phenolics content

The phenolic compounds are very important plant constituents because of their scavenging ability due to their hydroxyl groups (Hatano *et al.*, 1989). Based on the absorbance values of various extract solutions, reacted with Folin-Ciocalteu reagent and compared with the standard solutions of gallic acid equivalents, as described above, total phenolics are shown in Table 4.

Table. 4: The amount of total phenolic contents mg/g (GAE) d/w of various leaf extracts of *Curcuma zedoaria* R.

Extracts	Total phenolic (mg/g (GAE) d/w)		
Essential oil	ne		
Methanol extract	122.12 ± 1.6		
Hexane extract	40.2 ± 1.2		
Chloroform extract	61.15 ± 1.1		
Ethyl acetate extract	141.31 ± 1.5		

Values are given as mean ± S.D. of triplicate experiments. ne: not examined.

Among the organic extracts, the amount of total phenolics was higher in the ethyl acetate extract $(141.31 \pm 1.5 \text{ GAE mg/g} \text{ sample})$ as compared to methanol $(122.12 \pm 1.6 \text{ GAE mg/g} \text{ sample})$ and chloroform extracts $(61.15 \pm 1.1 \text{ GAE mg/g} \text{ sample})$. The lowest value was exhibited by the hexane extract $(40.2 \pm 1.2 \text{ GAE mg/g} \text{ sample})$. The key role of phenolic compounds as scavengers of free radicals is emphasised in several reports (Madsen *et al.*, 1996; Moller *et al.*, 1999). Phenolic antioxidants

are products of secondary metabolism in plants, and the antioxidant activity is mainly due to their redox properties and chemical structure, which can play an important role in chelating transitional metals, inhibiting lipoxygenase and scavenging free radicals (Decker, 1997). Some mechanisms are available for the mode of action of phenolic compounds in antioxidant activity test systems. One of them has been put forward by Ramirez-Anguiano et al., 2007. According to this group, the oxidation of diphenols to quinines is a very fast reaction, which might occur in seconds. Even when only a few quinines are formed before the preparation of the extract, they become to the low molecular weight compounds and might react spontaneously with other phenols, generating molecules like dopachrome, indolic compounds, catechol dimers and other higher polymers yielding radical scavenging degradation products. Therefore, it could be concluded that the phenolic compounds were highly involved in the antioxidant activity found in organic extracts of C. zedoaria leaves and also able to enhance or complement their activity.

CONCLUSION

This study concludes that the strong antioxidant and radical scavenging activity of the essential oil and extracts of *C. zedoaria* leaves points towards its strong protective role against oxidative diseases. The strong antioxidant activity indicates a possible use of residue oil as a natural antioxidant, food supplement and potential pharmaceutical application. However, further research is needed in order to establish the real application of *C. zedoaria* leaves essential oil or extracts in food or pharmaceuticals.

ACKNOWLEDGEMENT

We are grateful to Atomic Energy Commission Centre, Dhaka, Bangladesh for GC-MS analysis of essential oil. Besides, we are grateful to the Department of Biotechnology and Genetic Engineering Discipline, Khulna University, Bangladesh for the laboratory facilities for antioxidant assay.

REFERENCES

Adams R. P. 2007. Identification of essential oil components by gas chromatography/ Mass Spectroscopy (4th ed.), Carol stream, Illinois, Allured Publishing Corporation, USA.

Aeschbach R., Löliger J., Scott B. C., Murcia A., Butler J., Halliwell B., Aruoma O. I.. Antioxidant action of thymol, carvacrol, 6gingerol, zingerone and hydroxytyrosol. Food Chem Toxicol. 1994; 32: 31-36.

Archana B., Dasgupta, N., De B. In vitro study of antioxidant activity of Syzygium cumini fruit. Food Chem. 2005; 90: 727–733.

Burt S. Essential oils: their antibacterial properties and potential applications in foods – a review. Inter J Food Microbiol. 2004; 94: 223-253.

Cabrera A. C., Prieto J. M. Application of artificial neural networks to the prediction of the antioxidant activity of essential oils in two experimental *in vitro* models. Food Chem. 2010; 118: 141-146.

Chan H. W. S. 1987. Autoxidation of unsaturated lipids. Academic Press, London.

Cos P., Ying L.Y., Calomme M., Hu J.H., Cimanga K., Van Poel B., Pieters L., Vlietinck A.J., Vanden B. D. Structure-activity

relationships and classification of flavonoids as inhibitors of xanthine oxidase and superoxide scavengers. J Nat Prod. 1998; 61: 71–76.

Decker E. A. Phenolics: prooxidants or antioxidants? Nutr Rev. 1997; 55: 396-398.

Garg S. N., Naquvi A. A., Bansal R. P., Bahl J. R., Kumar S. Chemical composition of the essential oil from the leaves of *Curcuma zedoaria* Rosc. of Indian origin. J Essen Oil Res. 2005; 1: 29-31.

Hatano T., Edamatsu R., Mori A., Fujita Y., Yasuhara E. Effect of interaction of tannins with co-existing substances. VI. Effects of tannins and related polyphenols on superoxide anion radical and on DPPH radical. J Chem Pharm Bull. 1989; 37: 2016-2021.

Helander I. M., Alakomi H. L., Latva-Kala K., Mattila-Sandhom T., Pol I., Smid E. J., Gorris L. G. M., Von W. A. Characterisation of the action of selected essential oil components on gram-negative bacteria. J Agri Food Chem.1998; 46:3590-3595.

Jadhav S.J., Nimbalkar S.S., Kulkarni, A.D., Madhavi D.L. 1995. Lipid oxidation in biological and food system. New York.

Jang M.K., Sohn D.H., Ryu J.H. A curcuminoid and sesquiterpenes as inhibitors of macrophage TNF-alpha release from *Curcuma zedoaria*. Planta Med. 2001; 67: 550-552.

Kim K.I., Kim J.W., Hong B.S., Shin D.H., Cho Y., Kim H.K., Yang H.C. Antitumor, genotoxicity and anticlastogenic activities of polysaccharide from *Curcuma zedoaria*. J Mol Cell. 2000; 10: 392-398.

Lister, E. Wilson P. 2001. Measurement of total phenolics and ABTS assay for antioxidant activity, Crop Research Institute, Lincoln, Personal Communication, New Zealand.

Madsen H.L., Nielsen B.R., Bertelsen G., Skibsted L.H. Screening of antioxidative activity of spices. A comparison between assays based on ESR spin trapping and electrochemical measurement of oxygen consumption. Food Chem. 1996; 57: 331-337.

Matsuda H., Morikawa T., NIinomiya K., Yoshikawa M. Hepatoprotective constituents from Zedoariae rhizoma: absolute stereostructures of three new carabrane type sesquiterpenes, curcumenolactones A, B and C. Bio-organ Med Chem. 2001; 9: 909-916.

Moller J.K.S., Madsen H.L., Altonen T., Skibsted L.H. Dittany (*Origanum dictamnus*) as a source of water-extractable antioxidants. Food Chem. 1999; 64: 215-219.

Namiki M. Antioxidants/antimutagens in food. Criti Rev Food Sci Nutri. 1990; 29: 273-300.

Navarro D.D., Souza M.M., Neto, R.A., Golin V., Niero R.,Yunes R.A., Delle monache F., Cechinel V. Phytochemical analysis and analgesic properties of *Curcuma zedoaria* grown in Brazil. Phytomed. 2002; 9: 427-432.

Noguchi N., Niki E. 1999. Diet Nutrition and Health. (20th ed). Papas M. P., CRC Press, Florida.

Pokorny J. Natural antioxidant for food use. Trend Food Sci Tech. 1991; 9: 223-227.

Ramirez-Anguiano A.C., Santoyo S., Reglero G., Soler-Rivas C. Radical scavenging activities, endogenous oxidative enzymes and total phenols in edible mushrooms commonly consumed in Europe. J Sci Food Agri. 2007; 87: 2272-2278.

Ruberto G., Baratta M.T. Antioxidant activity of selected essential oil components in two lipid model systems. Food Chem. 2000; 69, 167-174.

Sánchez-Moreno C. Review: Methods used to evaluate the free radical scanvenging activity in foods and biological systems. Food Sci Tech Inter. 2002; 8: 121-137.

Soares J.R., Dins T.C.P., Cunha A P., Ameida L.M. Antioxidant activity of some extracts of *Thymus zygis*. Free Rad Res. 1997; 26: 469-478.

Staples G.W., Herbst D.R. 2005. A Tropical Garden Flora: Plants cultivated in the Hawaiian. Islands and other tropical places. Bishop Museum Press, Honolulu, Hawai'i.

Yamaguch T., Takamura H., Matoba T., Terao J. HPLC method of evaluation of the free radical scavenging activity of foods by using 1,1-diphenyl-2-picrylhydrazyl. Bioci Biotec Biochem. 1998; 62: 1201-1204.

Yanishlieva N.V., Marinova E.M., Gordon M.H., Raneva V.G. Antioxidant activity and mechanism of action of thymol and carvacrol in two lipid systems. Food Chem.1999; 64: 59-66.

How to cite this article:

Atiqur Rahman, Monira Afroz, Rafiquel Islam, Kazi Didarul Islam, M. Amzad Hossain and Minkyun Na. *In Vitro* Antioxidant Potential of the Essential Oil and Leaf Extracts of *Curcuma zedoaria* Rosc. J App Pharm Sci, 2014; 4 (02): 107-111.