Journal of Applied Pharmaceutical Science Vol. 3 (08), pp. 097-101, August, 2013 Available online at http://www.japsonline.com DOI: 10.7324/JAPS.2013.3817 ISSN 2231-3354 (cc) BY-NC-SA

ABSTRACT

Comparative Phytochemical Screening and Nutritional Potentials of the Flowers (petals) of Senna alata (I) roxb, Senna hirsuta (I.) Irwin and barneby, and Senna obtusifolia (I.) Irwin and barneby (fabaceae)

U. A. Essiett and I. E. Bassey

Department of Botany and Ecological Studies University of Uyo, Uyo.

ARTICLE INFO

Article history:

Received on: 16/03/2013 Revised on: 29/04/2013 Accepted on: 16/08/2013 Available online: 30/08/2013

Key words: Senna species, Phytochemical screening, Physicochemical analysis, Petals, Fahaceae

INTRODUCTION

Man since immemorial time has been using herb or plants products as medicine for developing immunity or resistance against cold, joint pains, fevers, and so on. Scientific data in a good number of medicinal plants investigated has been well documented (Gupta, 1994). However, only very few drugs of plant origin could reach clinical use and the National Formulatory could not adopt even a dozen of plants for medicines.

For this reason, a special effort is needed for the development of herbal drugs having therapeutic utility (Gupta, 1994). Medicinal principles are present in different parts of plants like root, stem, bark, heartwood, leaf, flower, fruits, or plant exudates. These medicinal principles are separated by different processes: the most common being extraction. Extraction is the separation of the required constituents from plant materials (Burkill, 2002). Plant derived substances has obtained greater attention in the recent years to prevent and cure human diseases as they are considered to be more bio-friendly. It is generally estimated that over 6000 plants in India and Africa are in use in tra ditional folk, and herbal medicine, representing about 75%

Comparative phytochemical screening and nutritional potentials of the flowers (petals) of S. alata, S. hirsuta and S. obtusifolia (Fabaceae) were investigated. S. alata, S. hirsuta and S. obtusifolia are plants that have been frequently used as a medicine. The phytochemical screening reveals the presence of saponins, tannins, flavonoids, and cardiac glycosides and the absence of alkaloids in all three species. Quantitative evaluation of the petals of S. alata, S. hirsuta and S. obtusifolia reveals moisture content (%) of 12.5, 13.5, 13; Ash content (%) 6,11, 9. Acid insoluble (%) 1.5, 2.5, 2; Sulphated ash (%) 5, 9, 5.3; protein (%) 5.1, 8.2, 4.1, Fats (%) 5, 3.5, 4.4; Fibre (%) 25, 40, 30; Carbohydrate (%) 53.7; 42, 40.7 respectively. The above results indicates that despite the absence of information on the phytochemical screening, quantitative parameters and nutritional potentials of the petals of these studied species they contain nutrients and mineral elements, and bioactive compounds that may be useful in nutrition and in the synthesis of various therapeutic drugs.

> medicinal needs of the third world countries (Veerachari and Bopaiah, 2011). According to Okoli et al. (2007), traditional society in Africa and else where have always used herbs to promote healing. In the recent years, researchers on Medicinal Plants have attracted a lot of attention globally. Large body of evidence has accumulated to demonstrate the promising potential of medicinal plants used in various traditional, complementary and alternate system of treatment of human diseases (Alam, 2009).

> Medicinal plants have provided the modern medicine with numerous plant-derived therapeutic agents (Evans, 2000 and Oladunmoye et al., 2009). Many plants contain a variety of phytopharmaceuticals, which have found very important applications in the field of Agriculture, Human, and Veterinary Medicine. Natural products play a dominant role in the development of novel drug leads for the treatment and prevention of disease (Newman et al., 2003; Gilani and Rahman, 2005).

> World Health Organisation estimates that 70% of population from many countries is using traditional medicine to cure various ailments (WHO, 1991). Senna alata L., Senna hirsuta L., and Senna obtusifolia L., are the woody annual herbs or undershrub herbs which are native to Africa as medicinal species with active functions and therapeutic agents (Ayo, 2010).

^{*} Corresponding Author

Email: u.essiett@yahoo.com

The main medicinal uses of S. alata are as a laxative or purgative and in the treatment of skin problems. For laxative purpose usually a decoction of the leaves is drunk and less often the flowers, roots, or the stem are used (Adedayo et al., 2001). The decoction of the leaves of S. hirsuta is used against irritation of the skin in Thailand and in Laos the seeds are used as a substitute for coffee while S. obtusifolia leaves, seeds, roots and flower are used in folk medicine, primarily in Asia. It is believed to possess a laxative effect like S. alata compared to S. hirsuta as well as to be beneficial for the eyes (Oliver, 2005, Sofowora, 2008). Skin problem treated with S. alata include ring worm, favus and other mycoses, impetigo, syphilis, sores, psoriasis, herpes, chronic lichen planus, Scabies, rashes, and itching (Burkill, 1995). According to Sofowora (2008), S. obtusifolia are use as folk remedy, the seeds are often roasted, then boiled in water to produce a tea but in Java and in South-East Asia, S. hirsuta are used medicinally for treating herpes. In India, leaf decoction of S. alata are used as an expectorant in bronchitis and dyspnoea, as an astringent a mouth wash and a wash in cases of eczema (a skin condition in which areas of skin become red, rough and Sore) (Neuwinger, 2000). The seed of S. hirsuta contains a watersoluble gum, though not in commercial quantities, and it also contain bioanthraquinone which may prove medicinally important (Sangat-Roemantoyo, 1997). The root decoction of S. alata is drunk to treat painful menstruation and tattooing (Etukudo, 2003). S. obtusifolia are used as food for caterpillars of some Lepidoptera (butterflies and Moth insects). In Queensland, S. obtusifolia is a weed of pastures where it competes with pasture for light, reduce available grazing areas, nutrients and water and can rapidly exclude all other species (Annig et al., 1989). S. hirsuta is used as a forage plant and green manure (Sangat-Roemantyo, 1997). S. obtusifolia is useful against gastrointestinal condition, and its leaves and sap are used as ingredient in soaps, shampoos and lotioning in the Phillipines (Oliver, 2005). Senna (Cassia) species have been of keen interest in phytochemical and pharmacological research due to their excellent medicinal values. They are well known in folk medicine for their laxative and purgative uses (Dalziel, 1948; Abo et al., 1999; Hennebelle et al., 2009). Besides, they have been found to exhibit anti-inflammatory (Chidume et al., 2001), antioxidant (Yen et al., 1998; Yen and Chuang, 2000), hypoglycaemic (Bhakta et al., 1997), antiplasmodial (Iwalewa et al., 1990, Iwalewa et al., 1997), Larvicidal (Yang et al., 2003), antimutagenic (Silva et al., 2008; Yadav et al., 2010) and anticancer activities (Prasanna et al., 2009). They are also widely used for the treatment of wounds (Bhakta et al., 1998), skin disease such as ringworm, scabies and eczema, gastrointestinal disorder like ulcers (Dalziel, 1948, Benjamin, 1980; Abo et al., 1999; Elujoba, Abere and Adelusi, 1999; Jacob et al., 2002), and Jaundice (Pieme et al., 2006). Etukudo (2003) pointed out that S. alata can be used to treat ailment like stomach pain during pregnancy, dysentery, haemorrhoids, blood in the urine (Schistosomiasis, gonorrhoea), convulsion, heart failure, oedema, jaundice, headache, hernia, one-sided weakness or paralysis in tropical Africa. Also, Sofowora (1993) reported that the young pods of S. alata are eaten as vegetable, but only in small quantities and toasted leaves are sometimes used as a coffee, S. alata has a strong decoction made of dried leaves used as an abortifacient. Senna, the leave juice is squeezed into the eye to treat eye sties, conjunctivitis (appolo) and other minor eye problems (Fabricant and Farnsworth, 2001). Generally, the leaf, flower, root and seed are used in herbal medicine all over the world (Dennis, 1988). Smith (2009) reported that the flower and young fruit are used as curries. Phytochemical investigation of plant extracts shows the presence of active principles in the plant parts like flower, bark, leaves, root, fruits, etc. phytochemicals are non-nutritive plant chemicals that have protective or disease preventive properties. Plant produce these chemicals to protect itself but research works demonstrates that many phytochemicals can protect humans against disease (Trease and Evans, 2002; Krishnaiah et al., 2009; Essiett et al., 2010). Knowledge of chemical constituents of plants is desirable because such information will be of value for the synthesis of complex chemical substances. Plants are rich in a wide variety of secondary metabolites such as alkaloids, flavonoids, Saponins, and tannins, etc, which have been found to in-vitro have anti-microbial properties (Alam, 2009). From available literature, there is no information on the phytochemical screening, quantitative parameters and nutritive values of petals of S. alata, S. hirsuta and S. obtusifolia. The aim of this research is to establish the chemical constituents of the flowers which would eventually be useful in preparing a monograph of the plant for proper and easy identification and also at establishing some diagnostic parameters such as quantitative parameters of the crude drug (flowers) and dietary awareness of its nutritional status. The significance of the studies is to prove that S. alata, S. hirsuta and S. obtusifolia have various therapeutic uses for the synthesis of drugs and medicinal plants in the developing countries.

MATERIALS AND METHODS

Fresh flower (petals) of *S. alata, Senna hirsuta* and *Senna obtusifolia* were collected from a bush in Ibesikpo, Nsit atai and Uyo Local Government Area of Akwa Ibom State.

Extraction of Plant Materials

The fresh petals of *S. alata, S. hirsuta* and *S. obtusifolia* were air dried and reduced to powder with the aid of a mortar and pestle. The powdered petals were accurately weighed and then 20g each were macerated cold in 400ml of 50% ethanol and distilled water for 72 hours (3 days) at room temperature following the method suggested by Sofowora (1993). The liquid extracts were recovered by filtration using cotton wool and glass funnel. The filtrate obtained was concentrated in a vacuo at 40°C to yield a semi-solid mass. The extract obtained was accurately weighed and then used for phytochemical screening.

Phytochemical Screening

Cold extraction was carried out on the materials, which was later concentrated to dryness in *vacuo* at 40° C. The dry extract was subjected to phytochemical screening according to the

standard methods of Sofowora (1993) and Trease & Evans (2002) and Harborne (1998) to detect the presence or absence of certain bioactive compounds.

Quantative Microscopy/Proximate Analysis

The moisture content of the powdered leaves was determined by loss on drying method (African Pharmacopoeia, 1986). The ash value, acid insoluble ash, water-soluble ash and sulphated ash were determined as described (British Pharmacopoeia, 1980; African Pharmacopoeia, 1986). The water and alcohol extractive values were obtained using the method outlined (Brain and Turner, 1975; British Pharmacopoeia, 1980). The fat (lipids), crude fibre, crude protein and carbohydrate were obtained using the method outlined (Pearson, 1976; Okon, 2005; AOAC, 2000).

RESULT

Phytochemical Screening

The result of the preliminary phytochemical screeing of the petals of *S. alata*, *S. hirsuta* and *S. obtusifolia* are summarized in Table 1. The petals of *S. alata*, *S. hirsuta and S. obtusifolia* reveals the presence of Cardiac glycosides (Liebberman's test and Salkowski test) in all three species and absence of Alkaloids in all three species. However, Saponin was found to be present in *S. alata* and *S. obtusifolia* but absent in *S. hirsuta*. Tannins was present in *S. alata* but absent in *S. hirsuta*. Also, Cardiac glycoside (keller killianis test) was present in *S. alata* but absent in *S. hirsuta* and *S. obtusifolia*.

Quantitative Evaluation

The quantitative evaluation of the powdered petals of *S. alata, S. hirsuta and S. obtusifolia* are: Moisture content (%) 12.5, 13.5, 13; Ash content (%) 6, 11, 9; Acid insoluble (%) 1.5, 2.5, 2 Sulphated Ash (%) 5, 9, 5.5 respectively (Table 2).

Nutritional Analysis

The nutritional analysis of the powdered petals of *S. alata, S.hirsuta and S. obtusifolia* are: Proteins (%) 5.1, 8.2, and 4.1; Fats (%) 5, 3.5, 4.4; Crude Fibre (%) 25, 40, 30; carbohydrate (%) 53.7, 42, and 40.7 respectively (Table 3).

 Table. 1: Result of Phytochemical screening metabolites in petals of S. alata, S. hirsuta and S. Obtusifolia.

Matabalitas	Inferences					
Metabolites	S. alata	S. hirsuta	S. obtusifolia			
Saponins	++	-	+			
Tannins	+	-	-			
Flavonoids	++	+	-			
Alkaloids			-			
Cardiac glycosides						
(a) Salkowski Test	+++	+++	+++			
(b) Keller killiani Test	+	-	-			
(c) Lieberman's Test	+	+	+			
Notos						

Notes

- Absent + Traces

++ Moderately present

+++ Abundantly present

Table. 2: Results of Quantitative Evaluation of the petals of *S. alata, S. hirsuta and S. Obtusifolia.*

Evaluation	Values (% W/V	% W/W)			
Parameters	S. alata,	S. hirsuta	S. obtusifolia		
Moisture content	12.5	13.5	13		
Ash Content	6	11	9		
cid insoluble	1.5	2.5	2		
Sulphated Ash	5	9	5.5		

Table.	3:	Result	of	Proximate	Analysis	of	Nutritional	Evaluation	of S.	alata,
S. hirsı	ıta d	and S. o	btı	ısifolia.						

Evaluation	Values (% W/W)				
Parameters	S. alata,(%)	S. hirsuta (%)	S. obtusifolia (%)		
Protein	5.1	8.2	4.1		
fats	5	3.5	4.4		
fibre	25	40	30		
carbohydrate	53.7	42	40		

DISCUSSION

Phytochemical screening of the petals of *S. alata, S.hirsuta and S. obtusifolia* reveals the presence of various bioactive compounds such as saponins, tannins, flavonoids, and cardiac glycosides (Salkowski, Keller Killiani and Lieberman's test) which are the basis of therapeutic potentials of medicinal plants. The presence of tannins as reported by Maynard (1997), is capable of lowering available protein by antagonistic competition and can therefore elicit protein deficiency syndrome, "Kwashiokor".

Saponin is responsible for its anti yeast, anti fungal, antidote, antimicrobial and antiinflamatory activities. It is also believed that the role of Saponin in plant is to protect against attack by potential pathogens (Sparg *et al.*, 2004). Flavonoids which are also known as vitamin p or plan modifier, elicit a wide of therapeutic activities as antihypertensive, antirheumatism as well as antimicrobial as identified with flavonoids (Veerachari, and Bopaiah, 2011). Essiett *et al.* (2010), reported that many plant containing flavonoids are diuretic and the antioxidants, the leaves and stems of these plants can be equally applied in each case. Cardiac glycosides were detected in the extract and this compound has been useful in the treatment of asthma (Trease and Evans, 2002). Quantitative evaluation is an important parameter in setting standard for crude drugs (Trease and Evans, 2002).

However, the values of solvent extractives can be a means of providing preliminary information on the quality of the drug. The results of the moisture content in *S. alata, S. hirsuta and S. obtusifolia* that was not high indicates less chances of microbial degradation of the drug during storage because excess moisture can result in the breakdown of important constituents by enzymatic activity and as a result may encourage the growth of yeast and fungi during storage (African Pharmacopoeia, 1986), as such the moisture content (%) of 12.5, 13.5, 13 in *S. alata, S. hirsuta and S. obtusifolia* respectively.

The general requirement for moisture content in Crude drugs was that, it should not be more than 14%, since it was normal, and implies that the plants can be stored for a longer period with lower chances of microbial attack and growth. The total ash value (%) was 6, 11, and 9 in *S. alata, S. hirsuta and S.*

obtusifolia respectively, this implies that plants have normal complexes of inorganic and organic compound (British Pharmacopoeia, 1980) but the value of Acid insoluble ash (%) are 1.5, 2.5, and 2 in *S. alata, S. hirsuta and S. obtusifolia*, this implies that the normal acid insoluble ash has a portion of ash content which was acid insoluble and hence, may be physiologically important as salt in the body when consumed. It also indicated high digestibility of plant when eaten (Ibrahim *et al.*, 2010). The sulphated ash value (%) is 5, 9, 5.5 in *S. alata, S. hirsuta and S. obtusifolia*, respectively.

This implies that *S. hirsuta* has a higher sulphated ash value than that of *S. alata* and *S. obtusifolia* and are good criteria to judge the identity and purity of crude drug. Proximate analysis of a food is the nutritional composition of that food.

It is the estimate of the nutritive value of human food in its chemical form. The proximate analysis as shown in Table 3 shows that the protein content is relatively low in S. obtusifolia (4.1%) than in S. hirsuta (8.2%) and S. alata (5.1%) but it can contribute to the formation of hormones which controls a variety of body functions such as growth, repairs and maintenance of body protein (Mau et al. (1999). The fat content of S. alata (5%) was higher than that of S. hirsuta (3.5%) and S. obtusifolia (4.4%) and the beneficial effect of high fat content can be used for storage and transport forms of metabolic fuel. Also, high fat content can be exploited for nutritional advantage in health (Omode et al., 1995). The crude fibre content of S. hirsuta (40%) was higher than that of S. obtusifolia (30%) and S. alata (25%) The carbohydrate content was higher in S. alata (53.7%) than in S. hirsuta (42%) and S. obtusifolia (40.7%). The relatively high carbohydrate content can be used as energy sources and also it is necessary in the digestion and assimilation of other food. However, this study reveals that S. alata, S. hirsuta and S. obtusifolia contains essential nutrients for good human and animal health.

CONCLUSION

S. alata, S.hirsuta and S. obtusifolia have been distinguished on the basis of phytochemical screening, quantitative evaluation and nutritional analysis. The presence of secondary metabolites such as saponin, tannins, flavonoids, cardiac glycosides, protein, fats, fibre and carbohydrate are of great importance as a source of new useful drugs. The soil type affects chemical constituents leading to chemical races of the From these studies, it can be concluded that all plants. three species of Senna have many beneficial effects with respect to the presence of the above secondary metabolite which are likely to combat with many disease and also boost the immune the phytochemical system. However, characterization of the extracts, the identification of responsible bioactive compound and quality of standards are necessary for future study. Pharmacological activities to elicit medicinal properties of each plant should be carried out by other researchers.

REFERENCES

Abo KA, Lasaki SW and Adeyemi AA. Laxative and Antimicrobal Properties of *Cassia* species growing in Ibadan. Nig. J. Nat. Prod. Med. 1999; 3: 47-50.

Adedayo O, Anderson WA, Moo-Young M, Snieckus V, Patil PA and Kolawole DO. Phytochemistry and antibacterial activity of *Senna alata* flower. Pharmaceutical Biology. 2001; 39(6): 408-412.

African Pharmacopoeia. General Methods of Analysis Pharmacopoeia, 11(1st ed.). 1986; pp. 121-208.

Alam, S. Antimicrobial Activity of Natural Products from Medicinal Plants. Gomal Journal of medical sciences. 2009; 7(1): 72-78.

Anning P, Bishop HG, Lambert G and Sutherland. Sicklepod replaces overgraced topical pastures. Queensland Agricultural Journal. 1989; 115: 188-192.

AOAC. Association of Official Analytical Chemists International. *Officials Methods of Analysis*. 17th Ed. Gaithersburg, M. D. 2000.

Ayo RG. Phytochemcial constituents and bioactivities of the extracts of *Cassia nigricans* Vahl: A review. Journal of medicinal plants research. 2010; 4(14):1339-1348.

Benjamin TV. Investigation of *Cassia alata*, a plant used in Nigeria in the treatment of skin diseases. J. Afr. Med. Plants. 1980; 3: 135-136.

Bhakta T, Mukherjee PK, Saha K, Pal M, Saha BP. Hypoglycaemic activity of *Cassia fistula* Linn. (Leguiminosae) leaf (methanol extract) in alloxan-induced diabetic rats. J. Ethnobot. 1997; 9: 35-38.

Bhakta T, Mukherjee PK, Saha K, Pal M, Saha BP. Studies on invivo wound healing activity of *Cassia fistula Linn*. (Leguminosae) leaves in rats. Rats. Nut. Prod., Sci. 1998; 4(2): 84-87.

Brain KR and Turner TD. The practical evaluation of phytopharmaceuticals, 1975; pp. 81 – 82. Wright Scientecnica Bristol.

British Pharmacopoeia. Ash value, Acid insoluble ash, water soluble extractive and alcohol soluble extractive, Vol. II, Appendix xii, Majesty's stationary Office. London, 1980; pp. 1276-1277.

Burkill HM. The Useful Plants of West Tropical Africa, 2nd Edition, Volume 3, Families J-L. Royal Botanic Gurdens, Kew, Richmond, United Kingdom. 1995; 857p.

Burkill HM. The useful Plants of West Tropical Africa. Second Edition, Vol. 4. Families S-Z., Addenda Royal Botanic Gardens, Kew, Richmond. United Kingdom. 2002; 689p.

Chidume FC, Gamaniel K, Amos S, Akah P, Obodozie O and Wambebe C. Pharmacological activity the methanolic extract of *Cassia nigricans* leaves. Ind. J. Pharmacol. 2001; 33: 350-356.

Dennis PA. Herbal medicine among the Miskito of Eastern Nicaragua. Economic Botany. 1988; 42(1): 16-28.

Dalziel JM. Useful Plants of West Tropical Africa. Crown Agents for the Colonies, London, 1948; pp. 178-180.

Elujoba AA, Abere AT, Adelusi SA. Laxative activities of *Cassia* pods sourced from Nigeria. Nig. J. Nat. Prod. Med. 1999; 3: 51-53.

Essiett UA, Bala DN and Agbakahi JA. Pharmacognostic studies of the leaves and stem of *Diodia scandens* SW in Nigeria. Archives of Applied Science Research. 2010 2(5): 124-198.

Etukudo I. Ethnobotany: Conventional and Traditional Use of Plants. First edition, Nigeria, Verdict investment Ltd, Uyo, 2003; 191p.

Evans WC. Trease and Evans *Pharmacognosy* (14 Edition) W. B. Saunders Company Ltd., London, 2000; pp. 19-20.

Fabricant DS and Farnsworth NR. The value of plants in Traditional Medicine for Drug discovery. Environmental Health Perspective. 2001; 109(1): 69-75.

Gilani AH, Rahman AU. Trends in ethnopharmacolgy. J. Ethnopharmacol. 2005; 100: 43-49.

Gupta SS. Prospects and Perspectives of natural plants products in medicine. J. Pharmacol. 1994; 26: 1-12.

Harborne JB. *Phytochemical Methods* (3rd Ed.). Chapman and Hall Ltd., London. 1998; pp. 1-302.

Hennebelle T, Weniger B, Joseph H, Sahpaz S and Bailleul F. Senna alata. Fitoterapia. 2009; 80: 385-393.

Ibrahim J, Ajaegbu, VC and Egharevba HO. Pharmacognostic and Phytochemical Analysis of *Commelina benghalensis L*. Journal of Ethnobotanical Leaflets, 2010; 14: 610-615.

Iwalewa EO, Lege-Oguntoye L, Rai PP, Iyaniwura TT and Etkin NL. *In-vitro* antimkalarial activity of leaf extracts *Cassia Occidentalis and Guiera senegalensis* on Plasmodium yoelii senegatensis. West Afr. J. Pharmacol. Drug Res. 1990; 4: 19-21.

Iwalewa EO, Lege-Oguntoye L, Rai PP and Iyaniwura TT. *In vivo* and *in vitro* andtimalarial activities of two crude extracts of *Cassia occidentalis* leaf. Nig. J. Pharm. Sci. 1997; 5: 23-28.

Jacob DL, Odeh SO, Otsapa PBL. Preliminary in vivo studies of the anti-ulcer effects of the crude seed and leaves extracts of *Cassia ocidentalis* in albino Wistar rats. J. Med. Trop. 2002; 4(2): 15-18.

Krishnaiah D, Devi T, Bono A and Sarbatly R. Studies on phytochemical constituents of Six Malaysian Medicinal Plants. J. Med. Plants Res. 2009; 3(2): 67-72.

Mau JL, Miklus MB and Beelman RB. Shell life Studies of food and Beverages Charalambous E.d. chem. Biol. Phys. Nutr. Aspect. 1999; 57: 475-477.

Maynard LA. Animal Nutrition. McGraw Hill book company Ltd. New York, 1997; pp. 47.

Neuwinger HD. African traditional medicine: A dictionary of plant use and applications. Medpharm Scientific, Stuttgart, Germany. 2000; 589 p.

Newman DJ, Cragg GM, Snadder KM. Natural Products as sources of new drugs over the period 1981-2002. J. Nat. Prod., 2003; 66(7): 1022-1037.

Okoli RI, Agbe O, Ohaju-Obodo JO, Mensah JK. Medicinal plants used for managing some common Ailment Among Esan People of Edo State, Nigeria. Pakistan J. Nutrition. 2007; 6(5): 490-496.

Okon EU. Handbook of basic food and beverage analysis. Etovin Publishers, AKS. Nigeria. 2005; Pp. 53-70.

Oladunmoye MK, Adetuyi FC, Akinyosoye FA. Effect of *Cassia hirsuta* (L) extract in DNA profile of some Microorganisms. Afr. J. Biotechnol. 2009; 8(3):447-450.

Oliver B. Medicinal plants in Nigeria. Nigeria College of Arts. In linking-hub.elsevier.com/retrieve/pi. February 23. 2005.

Omode AA, Fatoki OS and Olaogun KA. Phytochemical properties of some under exploited and non-conventional oil seeds. J. Agric. Food Chem. 1995; 43(11): 2850-2853.

Pearson D. Laboratory techniques in food analysis. The Butterworth Group, London, 1976; pp. 22 – 25.

Prasanna R, Harish CC, Pichai SD and Gunasekaran P. Anticancer effect of *Senna auriculata* leaf extract in vitro through cell cycle arrest and induction of apoptosis in human breast and largnx cancer cell lines. Cell Biol. Int. 2009; 33: 127-134.

Pieme CA, Penlap VN, Nkegoum B, Taziebou CL, Tekwu EM, Etoa FX and Ngongang J. Evaluation of acute and subacute toxicities of aqueous ethanolic. 2006.

Sangat-Roemantyo H. *Senna hirsuta* (L.) Irwin and Barneby. In Faridah HI and Van der Maesen LJG (eds.): Plant Resources of South-East Asia No. 11. Auxilliary Plant. Prosea Foundation, Bogor, Indonesia, 1997; pp. 231-232.

Silva CR, Monteiro MR, Rocha NM, Ribeiro AF, Caldeira-de-Araujo A, Leitao AC, Bezerra RJAC and Padula M. Assessment of antimutagenic and genotoxic potential of *Senna (Cassia angustifolia* vahi) aqueous extract using *in vitro* assays. *Toxicol. In vitro*, 2008; 22:212-218.

Smith YRN. Determination of Chemical Composition of Sennasiamea (Cassia leaves) Pakistan Journal of Nutrition, 2009; 8(2): 119-121.

Sofowora A. Medicinal plants and traditional medicine in Africa. Spectrum Books Limited, Ibadan, Nigeria, 1993; pp. 150-153.

Sofowora A. Medicinal Plants and Tradition Medicine in Africa. 3rd Ed., Spectrum Books Limited. Ibadan, Nigeria. 2008; Pp. 199-204.

Sparg SG, Light ME and Stadan JV. Biological Activities and Distribution of Plant Saponins. Journal of Ethnopharmacology. 2004; 94: 219-243.

Trease GE and Evans WC. Pharmacognosy (13th Edition). Bailliere Tindall, London. 1989; 171: 204-205.

Trease GE and Evans WC. Pharmacognosy. Harcourt Publishers Ltd., London, 2002; 72p.

Veerachari U and Bopaiah AK. Preliminary phytochemical evaluation of the leaf extract of five *Cassia* species. J. Chem. Pharm. Res. 2011; 3(5): 574-583.

World Health Organization. "Guideline for Assessment of the herbal Medicines" Programme on Traditional. WHO, Geneva, 1991; pp. 56-91.

Yadav JP, Arya V, Yadav S, Panghal M, Kumar S and Dhankhar S. *Cassia occidentalis* L.: A review on its ethnobotany, phytochemical and pharmacological profile. Fitoterapia, 2010; 81:223-230.

Yang Y, Lim M and Lee H. Emodin isolated from *Cassia* obtusifolia species. J. Agric. Food Chem. 2003; 51: 7629-763.

Yen GC, Chen HW and Duh PH. Extraction and Identification of an antioxidative component from *juemingzi (Cassia tora* L.) J. Agric. Food Chem. 1998; 46: 820-824.

Yen GC and Chuang DY. Antioxidant properties of water extracts from *Cassia tora* L. in relation to the degree of roasting. J. Agric Food Chem. 2000; 48: 2760-2765.

How to cite this article:

U. A. Essiett and I. E. Bassey. Comparative Phytochemical Screening and Nutritional Potentials of the Flowers (petals) of *Senna alata* (1) roxb, *Senna hirsuta* (1.) Irwin and barneby, and *Senna obtusifolia* (1.) Irwin and barneby (fabaceae). J App Pharm Sci, 2013; 3 (08): 097-101.